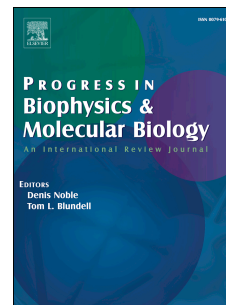


Journal Pre-proof

The role of DNA demethylation in induction of stem cells

Mohammad H. Ghazimoradi, Shirin Farivar



PII: S0079-6107(19)30216-0

DOI: <https://doi.org/10.1016/j.pbiomolbio.2019.12.005>

Reference: JPBM 1507

To appear in: *Progress in Biophysics and Molecular Biology*

Received Date: 20 September 2019

Revised Date: 27 December 2019

Accepted Date: 31 December 2019

Please cite this article as: Ghazimoradi, M.H., Farivar, S., The role of DNA demethylation in induction of stem cells, *Progress in Biophysics and Molecular Biology* (2020), doi: <https://doi.org/10.1016/j.pbiomolbio.2019.12.005>.

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2019 Published by Elsevier Ltd.

The role of DNA demethylation in induction of stem cells

Mohammad H. Ghazimoradi¹, Shirin Farivar^{1*},

1. Genetics, Stem Cells, Department of Cell and Molecular Biology, Faculty of Life Sciences and Biotechnology, Shahid Beheshti University, Tehran, 1983963113, Iran.

* corresponder email: s_farivar@sbu.ac.ir

Key words: DNA demethylation, induction of stem cells, pluripotent stem cells, embryo development, zygote

Abstract

DNA methylation is an epigenetic factor, which plays important roles in embryo and many other diseases development. This factor determines gene expression, and when half of them have CpG islands, DNA methylation and its enzyme effectors have been under the vast studies. Whole genome DNA demethylation is a crucial step of embryogenesis and also cell fate determination in embryos. Therefore, demethylation agents were used as a tool for dedifferentiation and transdifferentiation. Although many of these efforts have been successful, but using this method gave us a vast spectral cell type which is confusing. In this article, we briefly reviewed DNA methylation, and its role in embryogenesis and gene expression. In addition to that, we introduce studies that used this action as a direct method in induction of stem cells and cell fate decision.

Introduction

It has been shown that, epigenetic factors determine transcriptome of cells. These factors determine cell functions and identities [1]. Cell fate determination is among the most significant aspects influenced by these factors [2]. DNA methylation occurring on cytosine in mammals is a major epigenetic factor, which its presence and absence may pose control over 50% of genes, especially developmental-related genes [3]. It has been hypothesized that; DNA demethylation is one of the most significant events in initiation of

embryo development after fertilization, which has been named as Zygote Gene Activation (ZGA) [4]. This procedure induces expression of many stemness-related genes. Recently, many studies showed role of DNA demethylation and other epigenetic factors in changing cell fate [2]. It has been proved that, DNA demethylation could change fate of specific somatic cells to another somatic identity in a process named as transdifferentiation [5]. This phenomenon can reprogram somatic cells directly to stem cells [6]. DNA methylation enzyme effectors can establish new fates in differentiation process of stem cells as well [7]. Although it is believed that, disruption of epigenome or reactivation of developmental process in demethylation process could be a potential answer to these phenomena, but related findings have shown that, different roles of DNA demethylation are very confusing to determine. Furthermore, the basic concept and application of this field have been hindered because of its reproducibility and cell fate heterogeneity.

DNA methylation and its role in cell fate decision

DNA methylation appears on fifth position of cytosine by DNA methyltransferase1 (Dnmt1), Dnmt3a, and DNMT3b in mammals. Substrates of these enzymes are single cytosine or CG (CpG) content of the genome [8, 9]. In general, DNA methylation is known for its role in repressing and condensing chromatin, but it plays a variety of other roles based on its position in chromosomes and genes [10-12]. DNA demethylation will lead to gene transcripts upregulation [13]. DNA hypermethylation causes downregulation of the gene in promoters or enhancers [14, 15]. This factor might interfere with direct protein interaction to the DNA sequences like promoters and enhancers [16]. There is also a strong evidence for cross talking between DNA methylation and histone modifications. In general, H3K4 methylation, H3K27 acetylation, and DNA demethylation have a robust correlation with upregulation, and establish of each other's in-situ. H3, H1 deacetylation, H3K36 methylation, Ubiquitination, and DNA methylation on Promoters have a firm relationship with each other and also gene silencing. It has also been hypothesized that body methylation gene's silenced antisense transcript, which aborts mRNA degradation or vice versa [17, 18].

Dnmts are the enzymes, which were responsible for DNA methylation. Dnmt1 is known for maintenance of DNA methylation in cell division by recognizing hemimethylated CpG in PCNA and methylation of them [19]. It has been shown that the passive demethylation of genome by inhibition of Dnmt1 induces hypomethylation and Concentration of Dnmt1 will rise in the cell cycle phase of S/G1, due to methylation of new synthesis DNA, and cell fate maintenance [20]; other Dnmt family members are Dnmt3a and b, which most of their roles are in de novo methylation and establishing new pattern [21], It is remarkable that expression of Dnmt3a and Dnmt3b in order to lose pervious patterns will downregulate before the differentiation [22].

One of the most significant roles and changes in DNA methylation content of cells are in embryogenesis and induction of stem cells. The genome will go under heavy whole genome DNA demethylation after fertilization. In this notion, Dnmt1, Dnmt3a, and Dnmt3b will not be effective on the nucleus [23] and relocate the cytosol causing DNA demethylation of parental genomes by destabilizing and dilution of DNA methylation after every cell division, which is called passive demethylation. Tet1, Tet2, and Tet3 enzyme are another mechanism for demethylation of the genome (active demethylation), which catalyzes methyl cytosine to intermediated compounds [23]. It has been shown that many of embryogenesis and organogenesis genes, which are crucial for preimplantation and development of the zygote, are controlled and upregulated by demethylation [24]. This wave of upregulation includes many genes, and demethylation inducing dedifferentiates many cells to stemness lineage artificially [25-27].

After pervious stages and in initiation of organogenesis, Dnmt3a, and Dnmt3b upregulate and present in the nucleus, and de novo wave of methylation occurs subsequently. This de novo wave establishes the right pattern of epigenome, in accordance with specific fate of cells during this time of events, the majority of embryonic genes will heavily methylated and silenced (Figure1) [28].

Demethylation studies methods

Many researchers devote their time to investigate the importance of DNA demethylation in embryo development and its role in gene expression. There are different approaches to study about DNA demethylation. Many studies have been performed based on chemical component, which targets DNMTs. The most famous one amongst these agents is azacitidine (AZA-c) [29], which is a cytosine analog. This component is studied in stem cell and cancer researches extensively. This agent has a nitrogen atom in the fifth position instead of carbon, so there is no extra valance for methyl group to bond, according to this change [30]. This substance cannot be methylated and DNMTs captured in this area and it will be degraded promptly [31], and this procedure dilutes the DNMTs concentration. There are some upside downs for this agent. First, this agent makes the chromosomes unstable, and induces double strand break and mutation. Demethylation leads to apoptosis and senescing in somatic cells, which in addition to the AZA effect, will be catastrophic [32]. Another disadvantage of this agent is related to Dnmts variants, as some of them are not catalytic and as a result they cannot be affected.

Another way to study about demethylation is by genome editing methods based on knock downing Dnmts, which is a great approach to study demethylation, but most of these methods are permanent for genome editing [33], so cannot be tolerated by cells and creatures. Using siRNA because of numerous variants of Dnmts and knock downing efficiency may not be as effective as it needs to be, although some great progress had been made for this problem [34, 35].

The final approach is using array and next generation sequencing between embryonic stem cells and somatic cells as a comparative and predictive approach for demethylation studies [36]. Although all above mentioned methods for methylation have their own advantages and disadvantages, and also new chemical components have been discovered with a lower side effect for cells which has not been tackled on differentiation and transdifferentiation studies. The most important substance of these components is 5-aza-2-deoxycytidine; this substance is more tolerable for cells and animals subjected to these kinds of studies [37]. Another approach for investigation the role of DNA demethylation in induction of stem cells and transdifferentiation is the over expression of Tet enzymes in cells. Therefore, some researchers have

been performed to obtain pluripotent cells and muscle differentiation [38]. Although all above mentioned Tet enzymes play a significant role in demethylation of embryonic stem cells, it has been proven that Tet2 enzyme is contributed in stem cells inducing [39].

DNA demethylation and induction of stem cells

These surveys use the fact that embryo-related genes are heavily methylated, and demethylation leads to reactivation of these genes, in early embryo development. As a result, somatic cells demethylation might dedifferentiate cells to embryonic stem cells.

It has also been shown that demethylation upregulates many embryo-related genes (Figure1), and they are not only including epiblast markers like *Nanog*, and *Pou5f1* [40,41,42,43], but also it will reactivate many genes related to Trophoblast lineage like *Cdx2*, *Emoes*, *Elf5*, *Fgfr* [42,44], and primitive endoderm lineage like *Gata4* and 6 [45]. It is remarkable that genes like *Nanog*, *Cdx2* and *Gata4* have transcription factors, which are upregulated when the right network will be established in cells. It is considerable that because of these genes' role establishing totipotent stem cells and developing the embryos, and also providing the development potential to downstream lineage for totipotent stem cells happened [46]. It has been shown that demethylation of somatic cells, reprograms these cells to pluripotent stem cells and Trophoblast stem cells [41, 44]. However, the totipotent phenotype, despite existence of numerous studies with these substances has not been observed and obtained.

Other important genes that are reactivated by demethylation are *Hox* cluster, which are essential for hematopoiesis and embryo development (table1). Induction of these genes by demethylation would result in hematopoietic production of cells and their progenies [45].

Some interesting genes have been expressed after demethylation which is considerable, including *Dnmt3l* [46], which are only expressed in embryonic cells. Other examples are genes related to telomerase activity and elongation. It is shown that the transcript of the hTERT increases after demethylation [47]. The

demethylation increases expression of *12qF1* and its cluster, which will influence chimera formation, and will cause survival in pluripotent stem cells [48] (Tbale1). It is notable that the other demethylation substances, such as RG108 and decitabine, have been administered for inducing the stem cells, which lead to upregulation of *Nanog* and *Oct4* [49].

Role of DNA Demethylation in Differentiation and Transdifferentiation

Obviously, after fertilization and generation of totipotent stem cells (the cells which can generate embryonic and extra embryonic tissues) and their development to pluripotent (the cells which could generate all of three germ layers of cells including mesoderm, ectoderm, and endoderm) and multipotent cells (the cells which could rise to various cells from a defined germ layer), epigenetic pattern becomes more and more defined and restricted. It is notable that, in this process, stemness-related genes become strongly silenced (by DNA methylation and ubiquitination of histones) and other required genes will be activated [50, 51]. Eventually, when developmental stages come to an end, each type of cell has its own remarkable epigenetic pattern [52-56]. Significant role of epigenetics in differentiation can be elucidated by knock down of epigenetic enzyme effectors before induction of differentiation. After knock down of *Hdac 1or Dnmts* genes and withdraw of LIF from ES cells (the factors which are used rationally in ESc culture and are the main factors for self renewal of stem cells and maintenance of their stemness) normally leading to differentiation of these cells in three-germ layer cells, these cells hold their identity and cannot undergo differentiation steps nor cell death. This result is replicated by induction of stem cells with different simulators of differentiation [57-61].

DNA demethylation is applied for transdifferentiation purposes in disruption of epigenetic pattern or enhancing transdifferentiation process including transdifferentiation of fibroblast to other types of cell like adipocytes, myocytes, cardiocyte, and chondrocyte [62-64]. Heterogeneity of obtained cells has been reported as one of the most important observations of these surveys. Different types of cell will be obtained after DNA demethylation at low concentration. This issue has hindered progress related to this subject in the literature. In this essence, many researchers use this substance as an initiator of

intermediates, and promote it using other factors like growth factors in order to obtain desired cell fate. However, heterogeneity of cell fate remains an enigmatic problem due to obtaining different cell types (Table2).

Controversy between transdifferentiation and dedifferentiation

DNA somatic genome demethylation reactivated many genes (Table1). We now know that DNA methylation is the main obstacle and determining factor for repressing the genes [65]. Although we mentioned that many genes related to embryogenesis and development undergo the DNA demethylation, but the other genes play pivotal roles in cell fate, and other processes regulate by this mechanism as well. This issue may raise conflict in determination of cell's fate. DNA demethylation induced many different cell types from source cell type in agreement with this notion, which raised the question that how cell type will be determined. There can be some explanation about this subject (Figure2), one of the explanations to this controversy is that DNA demethylation dedifferentiated cells to stemness, and also lack of maintain factors differentiates these stem cells or partially reprograms stem cells to somatic cells. As a proof, withdrawing LIF or feeder layer from stem cell's culture leads to differentiation of stem cells to other somatic cells [66]. Second hypothesis in cell fate studies by DNA demethylation is that researchers use supporting proteins for establishing a specific cell fate, as this reagent can activate many genes[41,44,47], but in some cases if the DNA demethylation reactivated receptors and growth factors, it can be participated to determining cell's fate directly. Supporting proteins can give an advantage to particular cell subpopulation and also increase their population, if DNA demethylation differentiates somatic cells to wide spectral of cell types, and this is another hypothesis. The last explanation is based on demethylation induction, embryogenic activation, and organogenesis genes, which have been inhibiting role on each other and based on differences, cell fate will be determined in order of loci demethylation (which seems to be random), and their conflicts (Figure2).

Effect of demethylation agents on *in-vivo* generation of stem cells

The use of commercial name vidaza or decitabine, or names for AZA-c and 5-AZA-2-deoxycitidine, are wide spread for patients' treatment and clinical researches [67]. The main areas are cancers in which this drug can be considered. Although we do not know the exact mechanisms, many cancers have been occurred due to disruptions in epigenetics patterns, which can be erased by these agents. As a treatment for the disease, consuming this drug has a considerable effect, which is an improvement of the target tissue. The most accessible examples of this effect can be observed in leukemia and increasing number of hematopoietic stem cells [68].

Although curing the cancers, and blood cancers, especially leads to death caused by low number of blood cell counts, rather than cancer and organ malfunction [69], using this drug improves the level of hematopoietic and blood cell numbers in the majority of cases, and it can be due to induction of hematopoietic stem cells from mesenchymal cells in bone marrow. Actually, using epigenetics drugs in order to induce stem cells in *in-vivo* tests has been done [60, 70].

Cancer Stem Cells and DNA Methylation

Cancer Stem Cells (CSCs) are cancer cells that are capable of unlimited self-renewal and differentiation [71]. CSCs are one of the major theories related to development and progression of cancer. The significant role of epigenetics in initiation of CSCs rises when almost 25% of patients with Acute Myeloid Leukemia (AML) have mutation in their *DNMT3A*. These mutations occur in catalytic part of this enzyme [72]. Although many studies supported role of demethylation in CSCs, other investigations have indicated significant roles of DNA hypermethylation. This phenomenon rises from role of DNA methylation in gene expression [73-77]. It has been demonstrated that, hypomethylation of oncogenes leads to upregulation of these genes, resulting in initiation of cancer progression. Accordingly, hypermethylation of tumor suppressors could result in uncontrollable growth of cells as examples for these phenomena are found in P53, P21, and RB cancer suppressor genes locus [78-81]. Wnt and Hedgehog pathways and its related genes are significant genes influenced by this factor. These pathways have crucial roles in differentiation and formation of stem cells in embryo. It has been shown that, hypermethylation of Wnt genes pathway

such as *WIF-1* and *SFRP-1* will result in abnormal upregulation followed by progression of cancers through suppression of corresponding pathway. Hg pathway has also been reported to suffer from aberrant DNA methylation pattern. It has been elucidated that, hypomethylation of Shh promoter is linked with development of various types of cancer. Also, it has been shown that, aberrant DNA methylation and other epigenetic factors can initiate development of CSCs and EMT phenotype. In this regard, many DNA methylation modulators such as azacitidine and decitabine have been suggested for cancer elimination, which could lead to silencing of cancer-related genes and a significant decrease in the tumorigenicity of CSCs in many cases [82-84].

Conclusion

DNA demethylation studies, because of its roles in cell fate determination and diseases such as autoimmune diseases and cancers are a curial step in cellular and developmental biology. DNA methylation is a remarkable step in embryonic stem cells induction. It is a roadblock to dedifferentiation research, and a part of the natural way for stem cells inducing in embryos. Although there are many studies on the direct effect of epigenetic reprogramming on induction stem cells, previous studies have focused on its effect as an enhancer of induction. This neglect could be a direct affection of heterogeneity of reprogrammed cells, even in single study. The most reasonable answer to this questionable mixture could be induction of stem cells and future differentiation of reprogrammed stem cells. The differentiation of reprogrammed cells could be due to used medium and the disrupted transcriptome. Another raising matter due to this research is induction of totipotent stem cells which has not been detected over 50 years of research. This subject can be explained by the degree of demethylation which reaches its maximum level in blastula stage and presumably affects cells in these stages as pluripotent stem cells and trophoblast lineage. As it has been seen, this process could be complex. But advantages of this field is efficiency of induced stem cells;

dedifferentiation and first source transdifferentiation to a wide spectral of cell types by a natural way which could eliminate many obstacles in stem cells field.

ACKNOWLEDGMENTS

We also are grateful to Dr Ehsan Zolghadr from Alabama University for his considerable efforts in editing this manuscript. We should appreciate Dr Sadeg Safari efforts for designing the art works. We also would thank to Dr Siamak Ebrahimi for his valuable efforts to this review.

Author Disclosure Statement

All the authors declared no financial and non-financial competing interests exist.

References

1. Jaenisch, R., & Bird, A. (2003). Epigenetic regulation of gene expression: how the genome integrates intrinsic and environmental signals. *Nature genetics*, 33(3s), 245.
2. Paksa, A., & Rajagopal, J. (2017). The epigenetic basis of cellular plasticity. *Current opinion in cell biology*, 49, 116-122.
3. Bell, J. T., Pai, A. A., Pickrell, J. K., Gaffney, D. J., Pique-Regi, R., Degner, J. F., ... & Pritchard, J. K. (2011). DNA methylation patterns associate with genetic and gene expression variation in HapMap cell lines. *Genome biology*, 12(1), R10.
4. Minami, N., Suzuki, T., & Tsukamoto, S. (2007). Zygotic gene activation and maternal factors in mammals. *Journal of Reproduction and Development*, 53(4), 707-715.
5. Cho, Y. D., & Ryoo, H. M. (2018). Trans-differentiation via Epigenetics: A New Paradigm in the Bone Regeneration. *Journal of bone metabolism*, 25(1), 9-13.

6. Gomes, K. M. S., Costa, I. C., Santos, J. F. D., Dourado, P. M. M., Forni, M. F., & Ferreira, J. C. B. (2017). Induced pluripotent stem cells reprogramming: Epigenetics and applications in the regenerative medicine. *Revista da Associação Médica Brasileira*, 63(2), 180-189.
7. Gomes, K. M. S., Costa, I. C., Santos, J. F. D., Dourado, P. M. M., Forni, M. F., & Ferreira, J. C. B. (2017). Induced pluripotent stem cells reprogramming: Epigenetics and applications in the regenerative medicine. *Revista da Associação Médica Brasileira*, 63(2), 180-189.
8. Hotchkiss, R. D. (1948). The quantitative separation of purines, pyrimidines, and nucleosides by paper chromatography. *Journal of Biological Chemistry*, 175(1), 315-332.
9. Li, E., & Zhang, Y. (2014). DNA methylation in mammals. *Cold Spring Harbor perspectives in biology*, 6(5), a019133.
10. Jones, P. A. (2012). Functions of DNA methylation: islands, start sites, gene bodies and beyond. *Nature Reviews Genetics*, 13(7), 484-492.
11. Messerschmidt, D. M., Knowles, B. B., & Solter, D. (2014). DNA methylation dynamics during epigenetic reprogramming in the germline and preimplantation embryos. *Genes & development*, 28(8), 812-828.
12. Jones, P. A. (1999). The DNA methylation paradox. *Trends in Genetics*, 15(1), 34-37.
13. Laurent, L., Wong, E., Li, G., Huynh, T., Tsigos, A., Ong, C. T., ... & Wei, C. L. (2010). Dynamic changes in the human methylome during differentiation. *Genome research*, 20(3), 320-331.
14. Han, H., Cortez, C. C., Yang, X., Nichols, P. W., Jones, P. A., & Liang, G. (2011). DNA methylation directly silences genes with non-CpG island promoters and establishes a nucleosome occupied promoter. *Human molecular genetics*, 20(22), 4299-4310.
15. Lee, D. S., Shin, J. Y., Tonge, P. D., Puri, M. C., Lee, S., Park, H., ... & Kim, J. (2014). An epigenomic roadmap to induced pluripotency reveals DNA methylation as a reprogramming modulator. *Nature communications*, 5.

16. Thomson, J. P., Skene, P. J., Selfridge, J., Clouaire, T., Guy, J., Webb, S., ... & Turner, D. J. (2010). CpG islands influence chromatin structure via the CpG-binding protein Cfp1. *Nature*, 464(7291), 1082-1086.
17. Panning, B. and Jaenisch, R. (1996) DNA hypomethylation can activate Xist expression and silence X-linked genes. *Genes Dev.* 10, 1991–2002.
18. Dodge, J. E., Okano, M., Dick, F., Tsujimoto, N., Chen, T., Wang, S., ... & Li, E. (2005). Inactivation of Dnmt3b in mouse embryonic fibroblasts results in DNA hypomethylation, chromosomal instability, and spontaneous immortalization. *Journal of Biological Chemistry*, 280(18), 17986-17991.
19. Estève, P. O., Chin, H. G., Smallwood, A., Feehery, G. R., Gangisetty, O., Karpf, A. R., ... & Pradhan, S. (2006). Direct interaction between DNMT1 and G9a coordinates DNA and histone methylation during replication. *Genes & development*, 20(22), 3089-3103.
20. He, S., Sun, H., Lin, L., Zhang, Y., Chen, J., Liang, L., ... & Wang, F. (2017). Passive DNA demethylation preferentially up-regulates pluripotency-related genes and facilitates the generation of induced pluripotent stem cells. *Journal of Biological Chemistry*, 292(45), 18542-18555.
21. Robertson, K. D. (2001). DNA methylation, methyltransferases, and cancer. *Oncogene*, 20(24), 3139.
22. Li, J. Y., Pu, M. T., Hirasawa, R., Li, B. Z., Huang, Y. N., Zeng, R., ... & Xu, G. L. (2007). Synergistic function of DNA methyltransferases Dnmt3a and Dnmt3b in the methylation of Oct4 and Nanog. *Molecular and cellular biology*, 27(24), 8748-8759.
23. Bourc'his, D., Xu, G. L., Lin, C. S., Bollman, B., & Bestor, T. H. (2001). Dnmt3L and the establishment of maternal genomic imprints. *Science*, 294(5551), 2536-2539.
24. Okano, M., Xie, S., & Li, E. (1998). Dnmt2 is not required for de novo and maintenance methylation of viral DNA in embryonic stem cells. *Nucleic acids research*, 26(11), 2536-2540.

25. Cedar, H., & Bergman, Y. (2012). Programming of DNA methylation patterns. *Annual review of biochemistry*, 81, 97-117.
26. Wu, H., & Zhang, Y. (2014). Reversing DNA methylation: mechanisms, genomics, and biological functions. *Cell*, 156(1), 45-68.
27. Farthing, C. R., Ficz, G., Ng, R. K., Chan, C. F., Andrews, S., Dean, W., ... & Reik, W. (2008). Global mapping of DNA methylation in mouse promoters reveals epigenetic reprogramming of pluripotency genes. *PLoS Genet*, 4(6), e1000116.
28. Gonzalez, M., & Li, F. (2012). DNA replication, RNAi and epigenetic inheritance. *Epigenetics*, 7(1), 14-19.
29. Gonzalez, M., & Li, F. (2012). DNA replication, RNAi and epigenetic inheritance. *Epigenetics*, 7(1), 14-19.
30. Santi, D. V., Norment, A., & Garrett, C. E. (1984). Covalent bond formation between a DNA-cytosine methyltransferase and DNA containing 5-azacytosine. *Proceedings of the National Academy of Sciences*, 81(22), 6993-6997.
31. Schermelleh, L., Spada, F., Easwaran, H. P., Zolghadr, K., Margot, J. B., Cardoso, M. C., & Leonhardt, H. (2005). Trapped in action: direct visualization of DNA methyltransferase activity in living cells. *Nature methods*, 2(10), 751-756.
32. Pali, S. S., Van Emburgh, B. O., Sankpal, U. T., Brown, K. D., & Robertson, K. D. (2008). DNA methylation inhibitor 5-Aza-2'-deoxycytidine induces reversible genome-wide DNA damage that is distinctly influenced by DNA methyltransferases 1 and 3B. *Molecular and cellular biology*, 28(2), 752-771.
33. Liao, J., Karnik, R., Gu, H., Ziller, M. J., Clement, K., Tsankov, A. M., ... & Pop, R. (2015). Targeted disruption of DNMT1, DNMT3A and DNMT3B in human embryonic stem cells. *Nature genetics*, 47(5), 469.

34. Leu, Y. W., Rahmatpanah, F., Shi, H., Wei, S. H., Liu, J. C., Yan, P. S., & Huang, T. H. M. (2003). Double RNA interference of DNMT3b and DNMT1 enhances DNA demethylation and gene reactivation. *Cancer research*, 63(19), 6110-6115.
35. Fang, X., Poulsen, R. R., Wang-Hu, J., Shi, O., Calvo, N. S., Simmons, C. S., ... & Wendler, C. C. (2016). Knockdown of DNA methyltransferase 3a alters gene expression and inhibits function of embryonic cardiomyocytes. *The FASEB Journal*, 30(9), 3238-3255.
36. Farthing, C. R., Ficz, G., Ng, R. K., Chan, C. F., Andrews, S., Dean, W., ... & Reik, W. (2008). Global mapping of DNA methylation in mouse promoters reveals epigenetic reprogramming of pluripotency genes. *PLoS Genet*, 4(6), e1000116.
37. "Decitabine". 2016 National Center for Biotechnology Information. Retrieved September 24.
38. Bagci, H., & Fisher, A. G. (2013). DNA demethylation in pluripotency and reprogramming: the role of tet proteins and cell division. *Cell stem cell*, 13(3), 265-269.
39. Doege, C. A., Inoue, K., Yamashita, T., Rhee, D. B., Travis, S., Fujita, R., ... & Levine, R. L. (2012). Early-stage epigenetic modification during somatic cell reprogramming by Parp1 and Tet2. *Nature*, 488(7413), 652.
40. Messerschmidt, D. M., Knowles, B. B., & Solter, D. (2014). DNA methylation dynamics during epigenetic reprogramming in the germline and preimplantation embryos. *Genes & development*, 28(8), 812-828.
41. Tsuji-Takayama, K., Inoue, T., Ijiri, Y., Otani, T., Motoda, R., Nakamura, S., & Orita, K. (2004). Demethylating agent, 5-azacytidine, reverses differentiation of embryonic stem cells. *Biochemical and biophysical research communications*, 323(1), 86-90.
42. Huang, X., Han, X., Uyunbilig, B., Zhang, M., Duo, S., Zuo, Y., ... & Li, J. (2014). Establishment of bovine trophoblast stem-like cells from in vitro-produced blastocyst-stage embryos using two inhibitors. *Stem cells and development*, 23(13), 1501-1514.

43. Shi, J., Shi, W., Ni, L., Xu, X., Su, X., Xia, L., ... & Zhu, J. (2013). OCT4 is epigenetically regulated by DNA hypomethylation of promoter and exon in primary gliomas. *Oncology reports*, 30(1), 201-206.
44. Ng, R. K., Dean, W., Dawson, C., Lucifero, D., Madeja, Z., Reik, W., & Hemberger, M. (2008). Epigenetic restriction of embryonic cell lineage fate by methylation of Elf5. *Nature cell biology*, 10(11), 1280-1290.
45. Oda, M., Kumaki, Y., Shigeta, M., Jakt, L. M., Matsuoka, C., Yamagiwa, A., ... & Okano, M. (2013). DNA methylation restricts lineage-specific functions of transcription factor Gata4 during embryonic stem cell differentiation. *PLoS genetics*, 9(6), e1003574.
46. Dietrich, J. E., & Hiiragi, T. (2007). Stochastic patterning in the mouse pre-implantation embryo. *Development*, 134(23), 4219-4231.
47. Harris, D. M., Hazan-Haley, I., Coombes, K., Bueso-Ramos, C., Liu, J., Liu, Z., ... & Singh, S. (2011). Transformation of human mesenchymal cells and skin fibroblasts into hematopoietic cells. *PLoS One*, 6(6), e21250.
48. Kim, H., Park, J., Jung, Y., Song, S. H., Han, S. W., Oh, D. Y., ... & Kim, T. Y. (2010). DNA methyltransferase 3-like affects promoter methylation of thymine DNA glycosylase independently of DNMT1 and DNMT3B in cancer cells. *International journal of oncology*, 36(6), 1563.
49. Taylor, S. M., & Jones, P. A. (1979). Multiple new phenotypes induced in 10T12 and 3T3 cells treated with 5-azacytidine. *Cell*, 17(4), 771-779.
50. Cedar, H., & Bergman, Y. (2012). Programming of DNA methylation patterns. *Annual review of biochemistry*, 81, 97-117.
51. Suelves, M., Carrió, E., Núñez-Álvarez, Y., & Peinado, M. A. (2016). DNA methylation dynamics in cellular commitment and differentiation. *Briefings in functional genomics*, 15(6), 443-453.
52. Wu, H., Coskun, V., Tao, J., Xie, W., Ge, W., Yoshikawa, K., ... & Sun, Y. E. (2010). Dnmt3a-dependent nonpromoter DNA methylation facilitates transcription of neurogenic genes. *Science*, 329(5990), 444-448.

53. Lichtenstein, M., Keini, G., Cedar, H., & Bergman, Y. (1994). B cell-specific demethylation: a novel role for the intronic κ chain enhancer sequence. *Cell*, 76(5), 913-923.
54. Keller, G. O. R. D. O. N., Kennedy, M. A. R. I. O. N., Papayannopoulou, T. H. A. L. I. A., & Wiles, M. V. (1993). Hematopoietic commitment during embryonic stem cell differentiation in culture. *Molecular and cellular biology*, 13(1), 473-486.
55. McCool, K. W., Xu, X., Singer, D. B., Murdoch, F. E., & Fritsch, M. K. (2007). The role of histone acetylation in regulating early gene expression patterns during early embryonic stem cell differentiation. *Journal of Biological Chemistry*, 282(9), 6696-6706.
56. Dovey, O. M., Foster, C. T., & Cowley, S. M. (2010). Histone deacetylase 1 (HDAC1), but not HDAC2, controls embryonic stem cell differentiation. *Proceedings of the National Academy of Sciences*, 107(18), 8242-8247.
57. Jackson, M., Krassowska, A., Gilbert, N., Chevassut, T., Forrester, L., Ansell, J., & Ramsahoye, B. (2004). Severe global DNA hypomethylation blocks differentiation and induces histone hyperacetylation in embryonic stem cells. *Molecular and cellular biology*, 24(20), 8862-8871.
58. Jackson, M., Krassowska, A., Gilbert, N., Chevassut, T., Forrester, L., Ansell, J., & Ramsahoye, B. (2004). Severe global DNA hypomethylation blocks differentiation and induces histone hyperacetylation in embryonic stem cells. *Molecular and cellular biology*, 24(20), 8862-8871.
59. Sheaffer, K. L., Kim, R., Aoki, R., Elliott, E. N., Schug, J., Burger, L., ... & Kaestner, K. H. (2014). DNA methylation is required for the control of stem cell differentiation in the small intestine. *Genes & development*, 28(6), 652-664.
60. Potdar, P. D., & Prasanna, P. (2013). Differentiation of human dermal mesenchymal stem cells into cardiomyocytes by treatment with 5-azacytidine: concept for regenerative therapy in myocardial infarction. *ISRN Stem Cells*, 2013.
61. Assis, R. I., Wiench, M., Silvério, K. G., da Silva, R. A., da Silva Feltran, G., Sallum, E. A., ... & Andia, D. C. (2018). RG108 increases NANOG and OCT4 in bone marrow-derived mesenchymal cells through global changes in DNA modifications and epigenetic activation. *PloS one*, 13(12), e0207873.
62. Devereux, T. R., Horikawa, I., Anna, C. H., Annab, L. A., Afshari, C. A., & Barrett, J. C. (1999). DNA methylation analysis of the promoter region of the human telomerase reverse transcriptase (hTERT) gene. *Cancer research*, 59(24), 6087-6090.
63. Kim, K., Doi, A., Wen, B., Ng, K., Zhao, R., Cahan, P., ... & Yabuuchi, A. (2010). Epigenetic memory in induced pluripotent stem cells. *Nature*, 467(7313), 285-290.

64. Jones, P. A., & Taylor, S. M. (1980). Cellular differentiation, cytidine analogs and DNA methylation. *Cell*, 20(1), 85-93.
65. Lee, D. S., Shin, J. Y., Tonge, P. D., Puri, M. C., Lee, S., Park, H., ... & Kim, J. (2014). An epigenomic roadmap to induced pluripotency reveals DNA methylation as a reprogramming modulator. *Nature communications*, 5.
66. Cartwright, P., McLean, C., Sheppard, A., Rivett, D., Jones, K., & Dalton, S. (2005). LIF/STAT3 controls ES cell self-renewal and pluripotency by a Myc-dependent mechanism. *Development*, 132(5), 885-896.
67. Kaminskis, E., Farrell, A. T., Wang, Y. C., Sridhara, R., & Pazdur, R. (2005). FDA drug approval summary: azacitidine (5-azacytidine, Vidaza™) for injectable suspension. *The oncologist*, 10(3), 176-182.
68. . Derissen, E. J., Beijnen, J. H., & Schellens, J. H. (2013). Concise drug review: azacitidine and decitabine. *The oncologist*, 18(5), 619-624.
69. Price, V. E., & Greenfield, R. E. (1958). Anemia in cancer. In *Advances in cancer research* (Vol. 5, pp. 199-290). Academic Press
70. Chandrakanthan, V., Yeola, A., Kwan, J. C., Oliver, R. A., Qiao, Q., Kang, Y. C., ... & Villanueva, J. E. (2016). PDGF-AB and 5-Azacytidine induce conversion of somatic cells into tissue-regenerative multipotent stem cells. *Proceedings of the National Academy of Sciences*, 113(16), E2306-E2315.
71. Wainwright, E. N., & Scaffidi, P. (2017). Epigenetics and cancer stem cells: unleashing, hijacking, and restricting cellular plasticity. *Trends in cancer*, 3(5), 372-386.
72. Voigt, P., & Reinberg, D. (2013). Genomic and epigenomic landscapes of adult de novo acute myeloid leukemia The Cancer Genome Atlas Research Network. *N. Engl. J. Med.*, 368, 2059-2074.
73. Lu, R., Wang, P., Parton, T., Zhou, Y., Chrysovergis, K., Rockowitz, S., ... & Wang, G. G. (2016). Epigenetic perturbations by Arg882-mutated DNMT3A potentiate aberrant stem cell gene-expression program and acute leukemia development. *Cancer cell*, 30(1), 92-107.

74. Yang, L., Rodriguez, B., Mayle, A., Park, H. J., Lin, X., Luo, M., ... & Zhang, X. (2016). DNMT3A loss drives enhancer hypomethylation in FLT3-ITD-associated leukemias. *Cancer cell*, 29(6), 922-934.
75. Mayle, A., Yang, L., Rodriguez, B., Zhou, T., Chang, E., Curry, C. V., ... & Goodell, M. A. (2015). Dnmt3a loss predisposes murine hematopoietic stem cells to malignant transformation. *Blood*, 125(4), 629-638.
76. Koya, J., Kataoka, K., Sato, T., Bando, M., Kato, Y., Tsuruta-Kishino, T., ... & Kurokawa, M. (2016). DNMT3A R882 mutants interact with polycomb proteins to block haematopoietic stem and leukaemic cell differentiation. *Nature communications*, 7, 10924.
77. Russler-Germain, D. A., Spencer, D. H., Young, M. A., Lamprecht, T. L., Miller, C. A., Fulton, R., ... & Ley, T. J. (2014). The R882H DNMT3A mutation associated with AML dominantly inhibits wild-type DNMT3A by blocking its ability to form active tetramers. *Cancer cell*, 25(4), 442-454.
78. Herrero, A. B., Rojas, E. A., Misiewicz-Krzeminska, I., Krzeminski, P., & Gutiérrez, N. C. (2016). Molecular mechanisms of p53 deregulation in cancer: an overview in multiple myeloma. *International journal of molecular sciences*, 17(12), 2003.
79. Teramen, H., Tsukuda, K., Tanaka, N., Ueno, T., Kubo, T., Ando, M., ... & Miyoshi, S. (2011). Aberrant Methylation of p 21 Gene in Lung Cancer and Malignant Pleural Mesothelioma. *Acta medica Okayama*, 65(3), 179-184.
80. Schroeder, M., & Mass, M. J. (1997). CpG Methylation Inactivates the Transcriptional Activity of the Promoter of the Human p53 Tumor Suppressor Gene. *Biochemical and biophysical research communications*, 235(2), 403-406.
81. Stirzaker, C., Millar, D. S., Paul, C. L., Warnecke, P. M., Harrison, J., Vincent, P. C., ... & Clark, S. J. (1997). Extensive DNA methylation spanning the Rb promoter in retinoblastoma tumors. *Cancer research*, 57(11), 2229-2237.
82. Klarmann, G. J., Decker, A., & Farrar, W. L. (2008). Epigenetic gene silencing in the Wnt pathway in breast cancer. *Epigenetics*, 3(2), 59-63.
83. Suzuki, H., Watkins, D. N., Jair, K. W., Schuebel, K. E., Markowitz, S. D., Chen, W. D., ... & Toyota, M. (2004). Epigenetic inactivation of SFRP genes allows constitutive WNT signaling in colorectal cancer. *Nature genetics*, 36(4), 417.
84. Cui, W., Wang, L. H., Wen, Y. Y., Song, M., Li, B. L., Chen, X. L., ... & Mi, X. Y. (2010). Expression and regulation mechanisms of Sonic Hedgehog in breast cancer. *Cancer science*, 101(4), 927-933.
85. Wang, X., Gao, H., Ren, L., Gu, J., Zhang, Y., & Zhang, Y. (2014). Demethylation of the miR-146a promoter by 5-Aza-2'-deoxycytidine correlates with delayed progression of castration-resistant prostate cancer. *BMC cancer*, 14(1), 308.
86. Kim, H., Park, J., Jung, Y., Song, S. H., Han, S. W., Oh, D. Y., ... & Kim, T. Y. (2010). DNA methyltransferase 3-like affects promoter methylation of thymine DNA glycosylase

independently of DNMT1 and DNMT3B in cancer cells. *International journal of oncology*, 36(6), 1563-1572.

Legends

Table 1: List of embryogenesis and organogenesis reactivation genes in demethylation studies by AZA.

Table 2: cell types list, which has been activated by DNA demethylation.

Figure1: Demonstration of dynamic factors in different cleavage stages. Green line refers to the methylation amount, blue is demonstrating transcriptome of zygote, and correlation of its timing versus methylation, and red line illustrates oocyte transcriptome degradation.

Figure2 demonstrated different predicted answers to dedifferentiating and transdifferentiation controversy. First pathways are explaining that demethylation dedifferentiates with another cell line randomly. The second way demonstrates a two-stage hypothesis, which at first is somatic cells reprogram to stem cells, and then they differences with another lineage due to the absence of maintaining factors. Third way shows that maybe there is an internal conflict in transcription, and the outcomes are based on their concentration and network.

Table1.

genes	function	reference
<i>Nanog</i>	Key transcription factor of pluripotent stem cells and totipotent stem cells	41
<i>Pou5f1</i>	Key transcription factor of pluripotent stem cells and	41,55

	totipotent stem cells	
<i>Lifr</i>	Stem cells receptors	41
<i>Elf5</i>	transcription factor	44
	of trophoblast stem cells	
<i>Cdx2</i>	Key transcription factor	44
	of trophoblast stem cells and totipotent stem cells	
<i>Fgfr4</i>	Stem cells receptors	44
<i>Gata4</i>	Key transcription factor	45
	of pluripotent stem cells and totipotent stem cells	
<i>Gata6</i>	Stem cells receptors	45
<i>Hox cluster</i>	transcription factor	47
	of hematopoietic stem cells	
<i>C-kit</i>	Stem cells receptors	47
<i>Mir 146a</i>	A key miRNA in pluripotent network and regulation of OCT4	85
<i>F12q8</i>	A non coding RNA	63

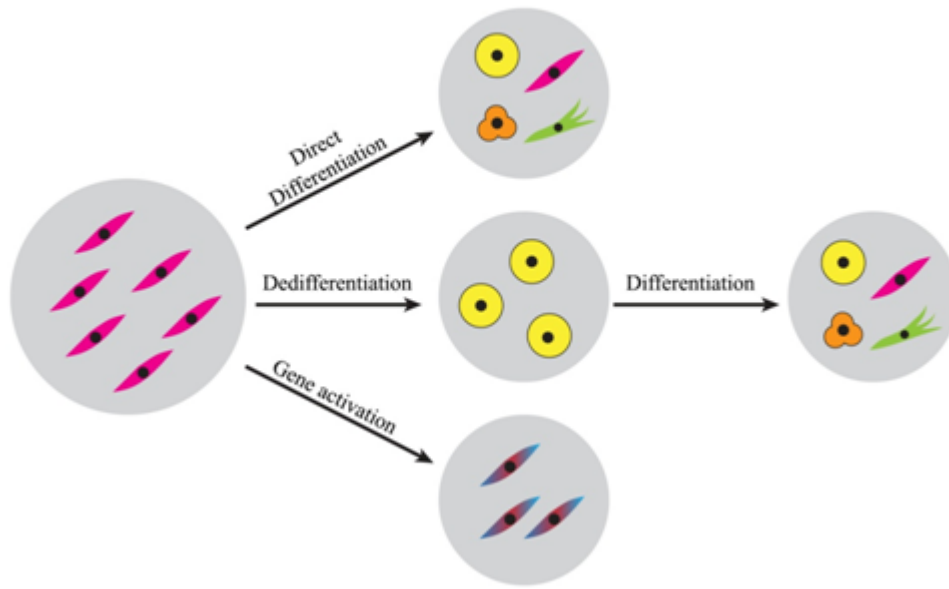
	related to pluripotent and chimer engrafting	
<i>Dnmt3l</i>	A epigenetic enzyme in first step of per implantation	33
<i>hTERT</i>	Increasing telomeres	62

Table2

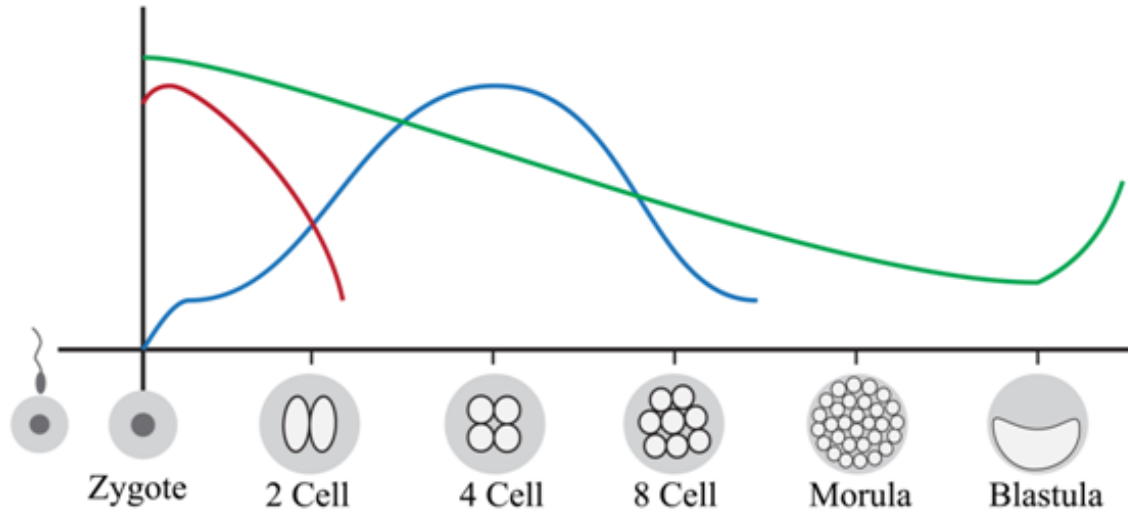
First fate	Second fate	Differentiation agent	reference
fibroblast	Pluripotent stem cell	AZA,AZA and TSA, AZA and LIF	41
	Trophoblast stem cells(MEGA)	AZA, AZA and FGF4	44
Fibroblast and mesenchymal cell	Hematopoietic cells	AZA, AZA and SCF	47
fibroblast	Adipocytes	AZA	65
	Chondrocyte	AZA	64,65
	myocyte	AZA	64,65
	Cardiomyocyte	AZA	66
	Monocyte and	AZA,AZA and SCF	47

lymphocyte

Journal Pre-proof



Journal Pre-proof



Journal ↑

- Epigenetics factors have undeniable duties in Developmental process of the preimplantation embryo.
- These factors are determining factors in regulation of natural stem cells induction.
-
- Many researchers have been administrating the reprogramming of epigenetics as a stem cell induction method.
- This method might be an answer to many problematic issues in fields of dedifferentiation.

Journal Pre-proof