



Synthesis of gum acacia-silver nanoparticles based hydrogel composites and their comparative anti-bacterial activity

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Abstract

An interpenetrating polymer network (IPN) containing gum acacia (GA), poly(methacrylic acid) (MAA), and poly(acrylic acid) (AA) was developed using a two-step aqueous polymerization method. Firstly, semi-IPNs were produced by radical polymerization of MAA chains onto GA in the presence of ammonium persulfate as a free radical initiator and N, N'-methylene-bisacrylamide (MBA) as a cross-linking agent using a microwave heating. To obtain a semi-IPN with a higher swelling percentage, several reaction parameters such as initiator, monomer, and crosslinker concentrations were varied. The percentage swelling (%S) was highly dependent upon the reaction conditions. The optimal reaction conditions for maximal %S were 2.55×10^{-2} mol/L initiator concentration, 12 mL solvent, 0.424×10^{-3} mol/L of monomer, and 2.16×10^{-2} mol/L cross-linker concentration, according to the findings. GA-g-poly(MAA) was the name to given to the semi-IPN. Second, IPN was created by grafting AA chains onto a GA-g-poly(MAA) matrix that had been optimized. The IPN was named as a GA-g-poly(MAA-IPN-AA). The reduction of silver ions to silver nanoparticles (AgNPs) was carried out by heating the mixture of flower extract of *Koelreuteria apiculata* under microwave radiation. Finally, the as-prepared semi-IPN and IPN samples were used as templates for the loading of AgNPs. XRD, FTIR, SEM, and TGA were used to characterize the synthesized semi-IPN, IPN, and their composites with AgNPs. GA, GA-g-poly(MAA), GA-g-poly(MAA-IPN-AA), and their composites with AgNPs are tested for antibacterial activity against five common bacteria strains: *Escherichia coli*, *Micrococcus luteus*, *Pseudomonas aeruginosa*, *Rhizobium species*, and *Staphylococcus aureus*. All of the bacteria strains were shown to have a noticeable zone of inhibition when IPN and their composite with AgNPs were used. When compared to other bacteria strains, *Pseudomonas aeruginosa* was observed to be more vulnerable to the tested samples. The obtained results demonstrate that the synthesized systems are suitable for application as antibacterial agents.

Keywords Biopolymer · Silver nanoparticles · Composite hydrogels · Graft-copolymerization · Microwave irradiation · Antimicrobial properties

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Introduction

Hydrogels are three-dimensional (3D) networks formed by the crosslinking of hydrophilic polymers [1, 2]. They might swell while maintaining their network structure by absorbing significant amounts of water or biological fluids. The inclusion of hydrophilic groups such as -OH, -CONH, -CONH₂, -COOH, and -SO₃H along the polymer chain contributes to their ability to absorb a large amount of water [1]. Polysaccharide hydrogel networks have received a lot of attention in recent years [3–5]. The physico-chemical properties of these hydrogels frequently differ significantly from those of the macromolecular constituents. Hydrogel characteristics can also be altered utilizing a range of physical and chemical crosslinking techniques [3, 6]. Various researchers have examined a number of polymeric systems to create hydrogels [7]. The polymer composition can be classified as natural polymeric hydrogels, synthetic polymer hydrogels, and or a combination of both [7]. Because of their non-toxicity, low cost, permeability, and biocompatibility, hydrogels have gotten a lot of attention in a variety of sectors [8–10].

Semi-interpenetrating polymer networks (semi-IPNs) and interpenetrating polymeric networks (IPNs) have developed as new materials for waste-water treatment, agricultural and horticultural applications, diapers and pad industries and biomedical and pharmaceutical applications [11–13]. The likelihood of integrating the beneficial features of each polymeric constituent of IPNs or semi-IPNs, resulting in a novel system with qualities that differ from the two separate components, has piqued interest in these structures [12, 14]. Semi-IPNs or IPNs develops the possibilities to tailor the features of the subsequent materials via selecting accurately the initial components of the IPN and semi-IPN frameworks [12]. Natural polysaccharides are used as a popular material for the synthesis of semi-IPN and IPNs hydrogels due to their favourable polymer system, abundance in nature, biocompatibility, and the likelihood to change their chemical structure through simple and efficient reactions [12, 15].

Gum acacia (GA) is a non-cytotoxic, hemostatic, nonhemolytic, antibacterial, antioxidant, biocompatible, cost-effective and anti-inflammatory natural polysaccharide [16–18]. GA is extracted from the stems and branches of *Acacia Senegal* trees, and its chemical composition varied depending on the tree's age, climate, and local soil conditions [19, 20]. It is normally composed of D-galactose, L-arabinose, L-rhamnose, D-glucuronic acid, and 4-O-methylglucuronic acid units which are joined by β -glycosidic chains [16–18]. In 2015, the potential applications of natural polymer GA were studied [21]. Several authors have described the synthesis of stimuli-responsive hydrogels based on GA [16, 22, 23].

Silver (Ag) is well-known for its antimicrobial and disinfectant properties, and it has a wide range of uses in medication and pharmaceuticals [24–26]. While silver nanoparticles (AgNPs) are gaining a lot of attention due to their ability to discharge Ag particles in a controlled way, which has a tremendous antibacterial effect on a wide range of microscopic organisms. AgNPs can agglomerate into groups due to their high surface to volume ratio, which limits their application. AgNPs in graft copolymeric systems represent a promising solution for stability and dispersal issues [27–29]. Furthermore, incorporating AgNPs into semi-IPN and IPN matrices based on GA can result in new composite hydrogels with improved antimicrobial properties. Because of their superior permeation and retention capabilities, AgNPs have been found to be particularly effective as an antibacterial agent, both in vivo and in vitro, when compared to bulk silver [20, 29, 30].

Zakia et al. [31] described a straightforward method for making alginate and AgNP hydrogels, as well as their antibacterial efficacy against a model bacterium (*E. coli*). They used a photo-crosslinking approach after changing the chemical structure of the polymer backbone with a methacrylate group to achieve homogeneous AgNP dispersion in the hydrogel. Mohan et al. [32] developed antibacterial hydrogel–silver nanocomposites that can be used in a variety of medical applications. Their research shows that hydrogels can be used as templates to create metal nanostructures of various sizes and morphologies. The composition of monomers or semi-IPN polymer chains, as well as the degree of cross-linking, appear to be important determinants in determining the shape and size of nanoparticles. Varaprasad et al. [33] developed poly(acrylamide)/poly(vinyl alcohol) hydrogel–silver nanocomposites to produce AgNPs with a size of 2–3 nm in gel networks. The production of nanocomposite is simple, direct, and quick, and the resulting hydrogel AgNP composites can be employed for antibacterial and wound dressing applications. Dai et al. [34] established a novel paradigm for the manufacture of silver nanoparticles (AgNPs)-containing smart hydrogels using guar gum and sodium borohydride (NaBH₄) in a simple, rapid, and cost-effective manner. They concluded that their study indicates a new class of smart metal nanoparticles/natural polymer-based hydrogels that can be made in a single step with NaBH₄. Juby et al. [20] described a one-pot synthesis of AgNPs within a polyvinyl alcohol/GA hydrogel via gamma irradiation-induced crosslinking. They discovered that adding GA to the hydrogels improved their biocompatibility and swelling properties. Against gram-negative *E. coli* bacteria, the composites demonstrate significant antibacterial activity. Singh and Dhiman [35] used GA-carbopol-crosslinked polyvinylimidazole hydrogels to wound dressings for the delivery of antibiotics and anaesthetics. Li et al. [36] developed a chronic wound healing delivery vehicle based on bioinspired alginate-gum Arabic hydrogels. GA extract has been shown to be haemostatic, non-haemolytic, and antimicrobial in recent research [37, 38].

We previously demonstrated the production of antibacterial and antifungal gum-acacia-poly(acrylamide-IPN-acrylic acid) nanocomposite hydrogels loaded with AgNPs [39]. In view of the same, it was planned to use microwave radiation to prepare GA-g-poly(MAA) and GA-g-poly(MAA-IPN-AA) hydrogels using a two-step aqueous free radical polymerization method. For the loading of AgNPs, the semi-IPN and IPN samples were employed as templates. These composite network systems provide state-of-the-art, with one portion of the polymeric network tries to improve the percentage swelling while another anchors antimicrobial characteristics. This opens a new window to tailor the property of interest. FTIR, SEM–EDX, XRD, and TGA were used to characterise the resulting semi-IPN, IPN, and their composites with AgNPs. Five strains *Escherichia coli* (*E. coli*), *Micrococcus luteus* (*M. luteus*), *Pseudomonas aeruginosa* (*P. aeruginosa*), *Rhizobium species* (*R. species*), *Staphylococcus aureus* (*S. aureus*) were used to test antibacterial properties in vitro. The main purpose was to evaluate the efficacy and antibacterial capabilities of the hydrogels and composites developed against a variety of bacteria available in our lab.

Materials and methods

Materials

Gum acacia (GA) was procured from Sisco Research Laboratories Pvt. Ltd (India). Ammonium persulfate (APS) was procured from Sigma Aldrich (USA). The monomer methacrylic acid (MAA) and acrylic acid (AA) were procured from LOBA CHEMIE PVT Ltd (India). The cross-linker N,N' methylene bisacrylamide (MBA) was procured from Central Drug House Ltd (India). On the other hand, AgNO₃ was procured from Merck, India. Distilled water was used during the synthesis process. The preparation of semi-IPN and IPN hydrogels was carried out in microwave equipment (IFB 20L) at 2450 MHz and 1200 W (India). The time for the synthesis of samples and microwave power @ 20% of the microwave oven remains the same throughout the synthesis procedure.

Preparation of semi-IPN and IPN hydrogels

Hydrogel based on GA was synthesized using microwave radiation with MAA as a monomer, APS as initiator, distilled water as a solvent, and MBA as a crosslinker. The microwave oven 17PM-MECI (IFB) utilised in the synthesis runs on 230 V- 50 Hz and operates at a frequency of 2450 MHz. 0.5 g of GA was weighed and mixed with

a known amount of dH₂O in the experiment. A known amount of APS was added to this, followed by MAA and MBA, and the solution was continually stirred in a clockwise orientation. To get the hydrogel with the best percentage swelling (%S), several reaction parameters such as initiator, cross-linker, and monomer concentrations, as well as the amount of solvent, were changed. Microwave irradiation was used for 40 s on the mixture. The synthesised hydrogel was given a repeated washing by soaking it in 10 ml distilled water for about 10 min and then drying it in a hot air oven at 60 °C. By weighing the hydrogels before and after swelling, the degree of swelling was determined gravimetrically. The conventional formulas were used to calculate the percentage swelling (%S) and grafting percentage (%G) of the produced hydrogels:

$$\text{Percentage swelling (\%S)} = \frac{W_t - W_o}{W_o}$$

where, W_t is the weight of polymer sample after swelling and W_o is its initial weight of in dry state.

$$\%G = \frac{W_f - W_b}{W_b} \times 100$$

where, W_f is the weight of functional polymer, and W_b is the weight of formed dried backbone.

GA-g-poly(MAA-IPN-AA) IPN hydrogels were prepared using preoptimized parameters used for the development of semi-IPN hydrogel under microwave radiation. A known amount of AA monomer was added drop-by-drop in optimal semi-IPN and distilled water in the experiment, and the mixture was continuously stirred. Continuous stirring was used to add the pre-optimized MBA. Optimized concentrations of APS (initiator) and MBA (cross-linker) were added to this reaction mixture. The reaction mixture was kept at room temperature for 14 h, which resulted in the formation of swollen hydrogels. Following that, the reaction mixture was agitated for a few minutes before being microwaved for two minutes to create IPN hydrogel. The IPN hydrogels were also rinsed many times with distilled water to remove homopolymers before being dried at 60 °C in a hot air oven. Table 1 displays the findings of the optimum %S and %G.

Preparation of silver nanoparticles (AgNPs)

By heating a mixture of flower extract of *Koelreuteria apiculata* under microwave radiation, silver ions were reduced to AgNPs. Our previous work [39] describes the full synthesis technique for the preparation of AgNPs.

Table 1 Optimum reaction conditions affording maximum percentage swelling for synthesis of GA-g-poly (MAA) and GA-g-poly(MAA-IPN-AA)

Sample Code	Optimized reaction parameters								%G	%S
	Back-bone (g)	Initiator (mol/L) × 10 ⁻²	Time (min.)	Amt. of solvent (ml)	pH	Monomer (mol/L) × 10 ⁻³	Cross-linker (mol/L) × 10 ⁻²	MW Power (%)		
GA-g-poly(MAA)	0.5	2.55	4	12	7	0.424	2.70	20	282	588
GA-g-poly(MAA-IPN-AA)	0.5	2.55	2.16	12	7	0.612	2.70	100	350	500

Synthesis of composite hydrogels

The ex-situ approach was used to prepare composite samples, as described in our previous work [39]. The loading of previously prepared AgNPs into the hydrogel matrices was completed by dipping 2 g of preoptimized poly(MAA) and GA-g-poly(MAA-IPN-AA) in 60 ml of AgNPs solution for 24 h each, and then placing the samples in a hot air oven at 60 °C for 24 h to achieve the preferred composites.

Characterizations

An FTIR spectrophotometer (Perkin Elmer) with diamond crystal and ZnSe for the focusing element was used to characterise the synthesised samples. SEM was used to determine the surface morphology of the composites (JEOL JSM-6490LV). Before the measurement, the samples were dried in air and sputter-coated with gold. The confirmation of AgNPs in the semi-IPN and IPN samples were confirmed by using EDX spectra. XRD pattern was measured by employing a Spinner PW3064, X'pert PRO diffractometer with Cu-K_α of 0.154056 nm at the 2θ scale. The thermal stability of prepared hydrogels was studied using a Thermal Analyzer (STA) STA7200.

In vitro microbiological assessment

The antibacterial properties of GA, GA-g-poly(MAA), GA-g-poly(MAA-IPN-AA), and their composites with AgNPs were tested using the agar well-diffusion method. For test organisms, MHA (Mueller Hinton Agar) was utilised as a supportive culture medium [40, 41]. The test organisms for the experiment (OD ≤ 0.1 at 600 nm) were gram-positive (*S. aureus* and *M. luteus*) and gram-negative (*Rhizobium species*, *E. coli*, and *P. aeruginosa*) bacterial cultures that had been cultivated overnight. A 50 mg sample of each of the test samples was used. In the assay, an antibiotic (Norfloxacin 5mcg) was employed as a control, and the zone of inhibition was determined in millimetres. The MHA plate surface is incubated by using a sterile cotton swab to disperse the necessary amount (100 µl) of

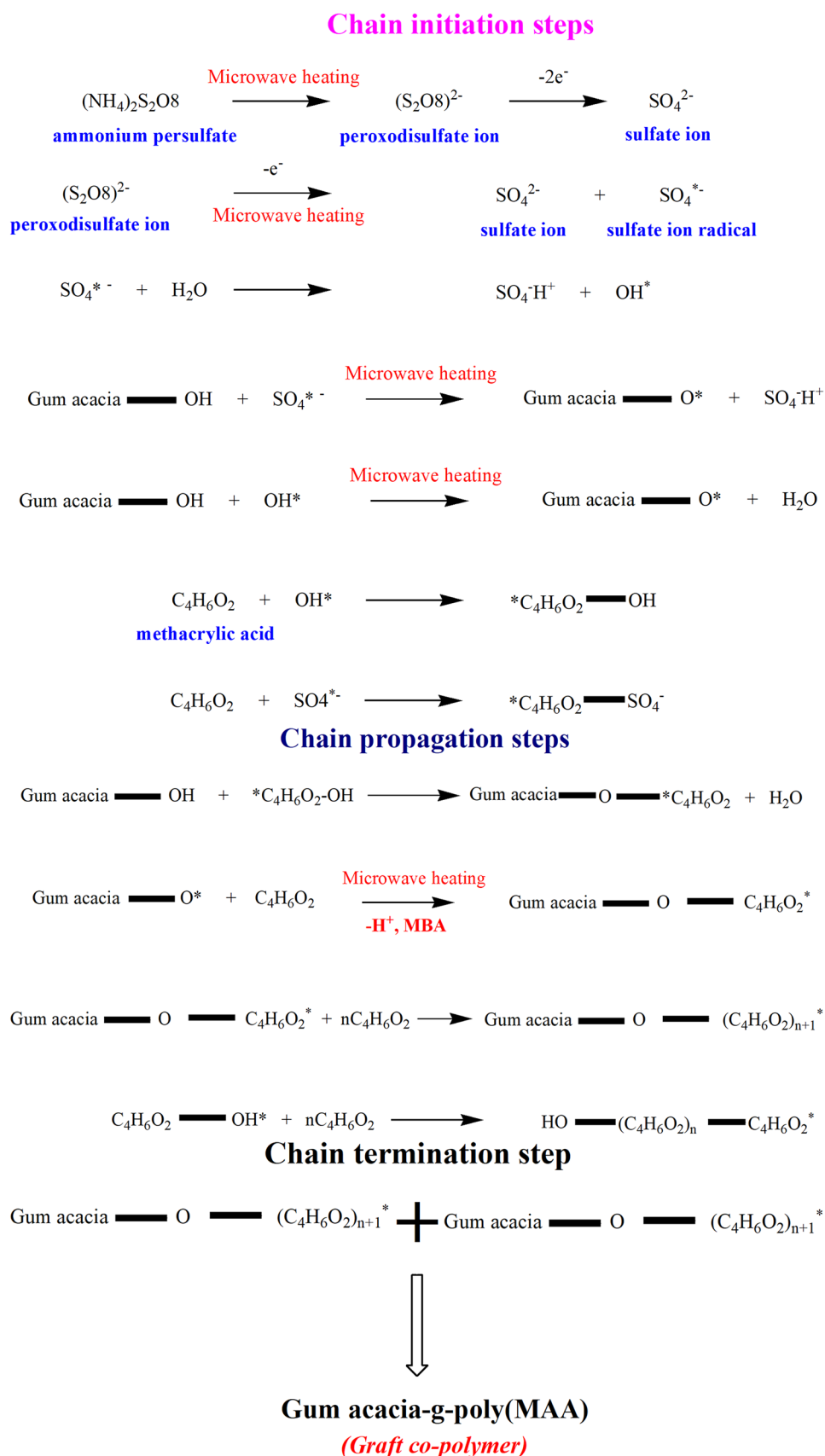
the test organism (OD ≤ 0.1 at 600 nm) across the entire agar surface. Then, using a sterile cork borer or an auto-pipette tip, a well with a diameter of 6-8 mm is punched aseptically (1 mL). The necessary amount of test sample (50 mg) is aseptically injected into the well, and the assay plates are carefully transferred to an incubator (37 °C) for overnight incubation. The test sample diffuses in the agar media and inhibits the test organism's growth, as assessed by the zone of inhibition.

Results and discussion

Reaction mechanism

The probable crosslinking mechanism is depicted in Scheme 1 and consists of three steps: chain initiation, chain propagation, and chain termination. These reactions are well known in the literature. Under the action of microwave heating, ammonium persulfate may break down to peroxydisulfate ions [(S₂O₈)²⁻], and subsequent removal of electrons from (S₂O₈)²⁻ results in the creation of sulphate ions [42]. Continuous microwave heating can produce sulphate ion radicals (SO₄^{*-}), which combine with H₂O molecules to create hydroxyl radicals (OH^{*}) [42, 43]. The -OH groups of the GA backbone and methacrylic acid (MAA) monomer chains are attacked by these free radicals (SO₄^{*-} and OH^{*}), resulting in the formation of macro radicals for the reaction. At the active sites, the stimulated MAA chains were grafted into the GA backbone to create a partially cross-linked network. To make GA-g-poly(MAA) semi-IPN hydrogels, MBA was used to cross-link the poly(MAA) and partially grafted network. The production of covalent bonds results from the recombination of macro radicals on various chains, followed by the construction of a cross-linked network [44]. AA monomer units were grafted onto GA-g-poly(MAA) semi-IPN hydrogel in a similar way. Due to the presence of SO₄^{*-} and OH^{*} radicals in the reaction mixture, active sites were produced on the AA monomer and semi-IPN hydrogel. IPN GA-g-poly(MAA-IPN-AA) hydrogel is the name given to the resulting network.

Scheme 1 Mechanism of graft copolymerization of methacrylic acid onto gum acacia



Optimized reaction conditions

The concentration of the initiator, the amount of solvent, the reaction time, the monomer concentration, and the crosslinker concentration were all optimized (Table 1). The degree of grafting and swelling capacity of hydrogels are heavily influenced by these parameters. Various reaction conditions influencing percentage swelling (%*S*) and percentage grafting (%*G*) are discussed below:

The initiator concentration was varied from 0.0182–0.0328 mol/L to study its effect on percentage swelling (Fig. 1a). The %*S* is improved with increasing initiator concentration. This is expected behavior in hydrogel networks that has been explored earlier by various authors [45, 46]. The %*S* attained a maximum value at the initiator concentration of 0.0255 mol/L. After this, the %*S* decreased with a further increase in initiator concentration.

At lower concentrations, the rise in %*S* is ascribed to the rise in the number of free radicals generated on.

The amount of the solvent was varied from 6 ml – 14 ml and its effects on the %*S* are shown in the (Fig. 1b). The increase in %*S* was observed until 12 ml of the solvent concentration and after it decreased [47]. The optimized amount of the solvent was identified to be 12 ml for attaining maximum %*S*. This may be owing to the fact that at the optimum amount of solvent there is a generation of hydroxyl radical ions giving more active sites for polymerization and thus gives rise in %*S*. Above the optimized amount of solvent, the %*S* started decreasing due to the lack of active sites for graft polymerization over the GA backbone and on monomer [48].

The changes in percentage swelling (Fig. 1c) was also studied by varying the monomer concentration. The %*S* increased with a rise in the concentration of monomer achieving maximum %*S* at 0.424×10^{-3} mol/L concentration

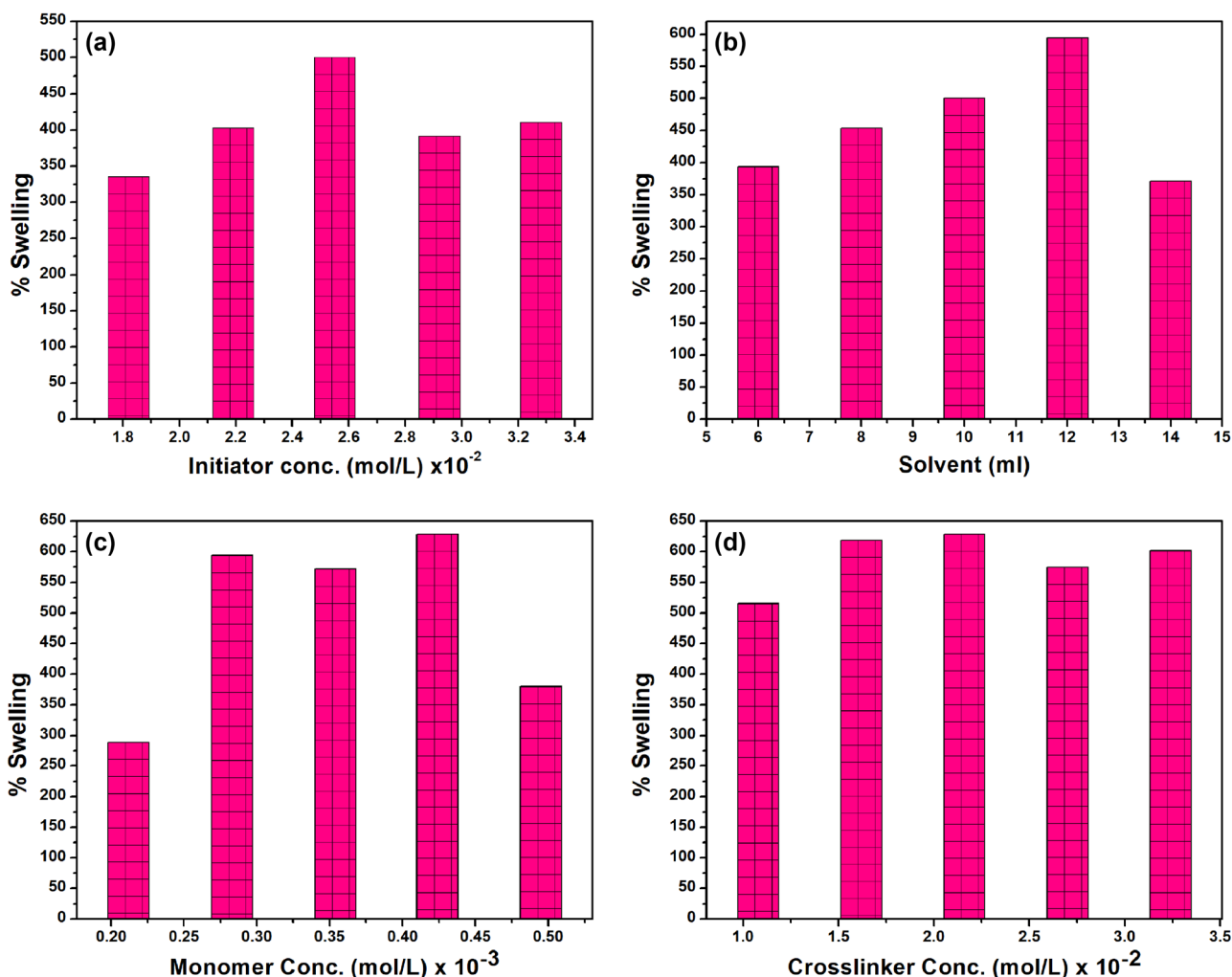


Fig. 1 Variation of percentage swelling with (a) initiator concentration, (b) amount of solvent, (c) monomer concentration, and (d) crosslinker concentration

of MAA and later it decreased. This behavior arises from the fact that in initial stages greater availability of hydrophilic monomer MAA for its cross-linking with GA backbone thus increasing the affinity of water absorption resulting in high %*S*. However, above the optimized MAA concentration the %*S* decreased because of the increased viscosity of the solution that hindered the motion of free radicals and monomer molecules and by the virtue of preferential homo polymerization than graft copolymerization [49].

The concentration of cross-linker was varied from 0.0108–0.0324 mol/L to study its effect on %*S* (Fig. 1d). The crosslinker is necessary as it makes the hydrogel insoluble in water by completely dissolving the hydrophilic polymer chains into an aqueous solution [49]. The %*S* increased by the rise in the concentration of MBA till 0.0216 mol/L where the maximum %*S* of 628% was achieved. On the basis of Flory's network theory, the fluid absorbency in hydrogels, and the density of cross-linking is determined from the concentration of the cross-linking chain [50]. Therefore, the number of network chains enhanced with a rise in the concentration of cross-linker resulting in an improved density of cross-linking. With the rise in MBA content above 0.0216 mol/L, the swelling capacity of grafted polymer decreased. It is because with increased cross-linker concentration there may be a generation of many chains in the polymeric network resulting in a highly compact and rigid structure. This suggested structure would result in low swelling as it will not succeed to uptake a large volume of water.

FTIR spectra analysis

To confirm the graft copolymerization mechanism, FTIR spectroscopy was used. Figure 2 shows the FTIR spectra of GA, semi-IPN, IPN, semi-IPN-AgNPs and IPN-AgNPs. The GA exhibited a broad peak at 3285 cm⁻¹, which is attributed to the hydrogen-bonded OH group [39]. No wide peak is found in any of the samples after grafting in the range 3300–3500 cm⁻¹, which indicates the stability of all the samples. The peak at 2925 cm⁻¹ and 2990 cm⁻¹ in all the samples is ascribed due to the C-H stretching. However, these peaks are quite prominent in semi-IPN-AgNPs and IPN. The slight shift in peak positions and variations in peak intensity indicates the impregnation of AgNPs onto the hydrogel matrices based on semi-IPN and IPN hydrogels. The sharp peaks around 1696 cm⁻¹ and 1157 cm⁻¹ attributed due to C=O stretching of carbonyl groups and C–O stretching, respectively [45]. The small peaks in the range of 1030–1070 cm⁻¹ is arises due to the stretching vibration of C–O–C and C–O–H bonds, which confirms the saccharide structure of the prepared composite samples [45]. The peak at 1258 cm⁻¹ attributed to C–N stretching vibrations [43]. It is clear from Fig. 2 that the wavenumber of prepared hydrogels is slightly shifted after

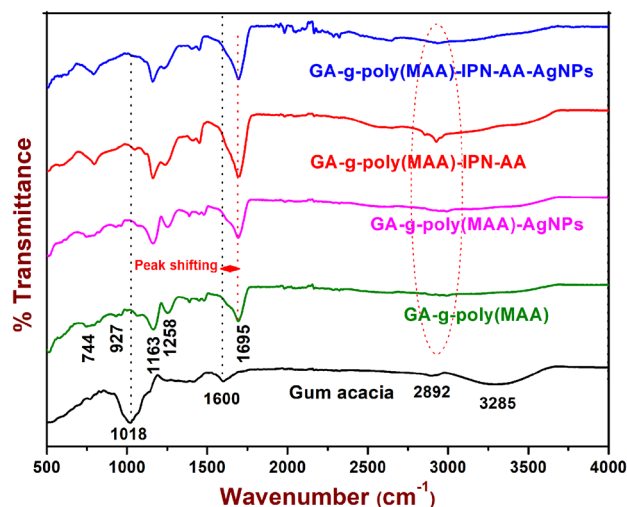


Fig. 2 FTIR spectra of semi-IPN, IPN and their nanocomposites with silver nanoparticles

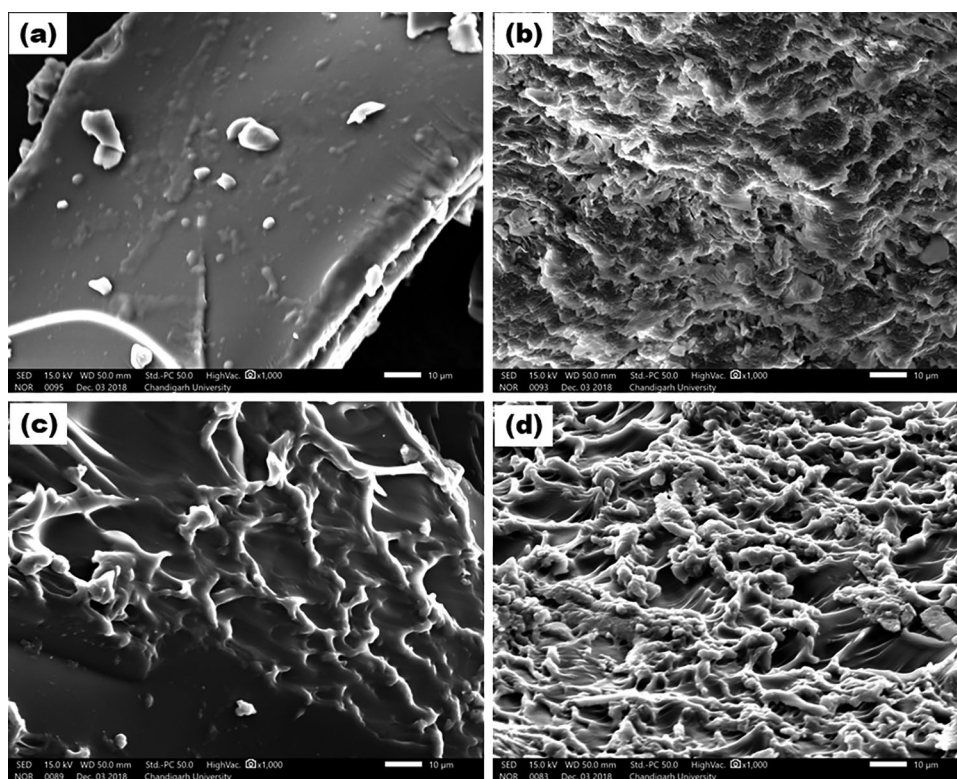
incorporation of AgNPs towards higher or lower wavenumber region which indicated the creation of covalent bonding among AgNPs and electron-rich units of the GA-g-poly(MAA) and GA-g-poly(MAA-IPN-AA) hydrogels. The observed peak shows network chain formation due to polymerization of MAA and on GA and further polymerization of AA on the GA-g-poly(MAA) network. Results of FTIR studies reflected the presence of GA, MAA, and AA in the semi-IPN and IPN hydrogels.

SEM-EDS analysis

SEM was used to analyse the surface morphology of the synthesized hydrogel systems and their composites with AgNPs, and the resulting micrographs are shown in Fig. 3. GA exhibits a uniformly homogeneous surface morphology (Fig. 3a) [39]. The surface morphology of the resultant semi-IPN is entirely modified after graft copolymerization with MAA. A rough surface with a honey beehive-like structure may be seen in the microscope (Fig. 3b). Further crosslinking with AA resulted in IPNs with a visible uniform crossing fiber-like shape, which increases roughness and agrees well with XRD results (Fig. 3c). Alternatively, the incorporation of AgNPs into the IPN matrix shows a clear formation of IPN AgNPs composite. The roughness is increased and visible globular knots are appeared over highly crisscrossed fibers like geometry (Fig. 3d).

The elemental analysis of AgNPs loaded semi-IPN and IPN composite was done by using EDS to confirm the incorporation of AgNPs in the hydrogel networks (Fig. 4). The presence of Ag peaks show that AgNPs are incorporated into the semi-IPN and IPN matrix.

Fig. 3 SEM images (a) Gum acacia, (b) semi-IPN, (c) IPN, and (d) IPN-AgNPs nanocomposite



XRD analysis

The XRD patterns of GA, semi-IPN, IPN, semi-IPN-AgNPs, and IPN-AgNPs are displayed in Fig. 5. In GA a broad peak at around 19° was obtained at 2θ scale with a comparatively lesser intensity in contrast to the prepared hydrogels [38]. The diffraction pattern of GA is semi-crystalline with a dominant amorphous nature.

The crosslinking of GA with MAA showed that the broad maxima shifted towards a higher angle and another sharp peak is obtained at an angle 15.6° . The crosslinking of MAA chains onto GA increases the relative intensity which suggested the improvement of crystallinity due to the interaction of MAA chains with different groups GA backbone [39]. In the case of AgNPs loaded semi-IPN, similar behaviour is observed with a small drop in peak intensity.

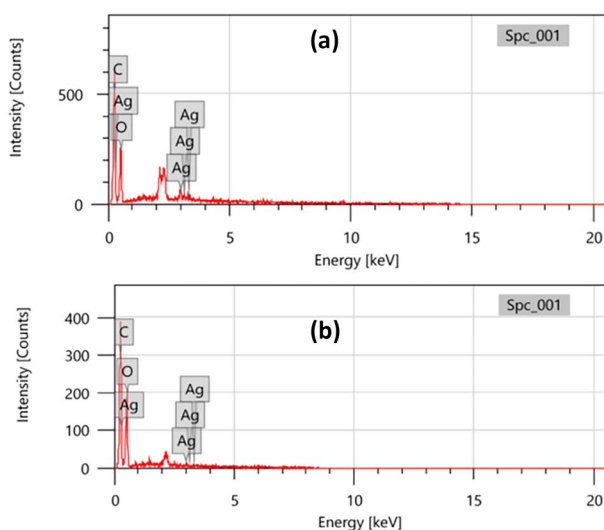


Fig. 4 EDS spectra of (a) semi-IPN-AgNPs and (b) IPN-AgNPs

Display name	Standard data	Quantification method	Result Type
Spc_001	Standardless	ZAF	Metal
Element	Line	Mass%	Atom%
C	K	43.41±0.38	54.39±0.48
O	K	47.07±1.06	44.28±1.00
Ag	L	9.52±0.63	1.33±0.09
Total		100.00	100.00
Spc_001			Fitting ratio 0.5357

Display name	Standard data	Quantification method	Result Type
Spc_001	Standardless	ZAF	Metal
Element	Line	Mass%	Atom%
C	K	47.25±0.52	56.22±0.62
O	K	48.36±1.28	43.20±1.14
Ag	L	4.39±0.99	0.58±0.13
Total		100.00	100.00
Spc_001			Fitting ratio 0.4676

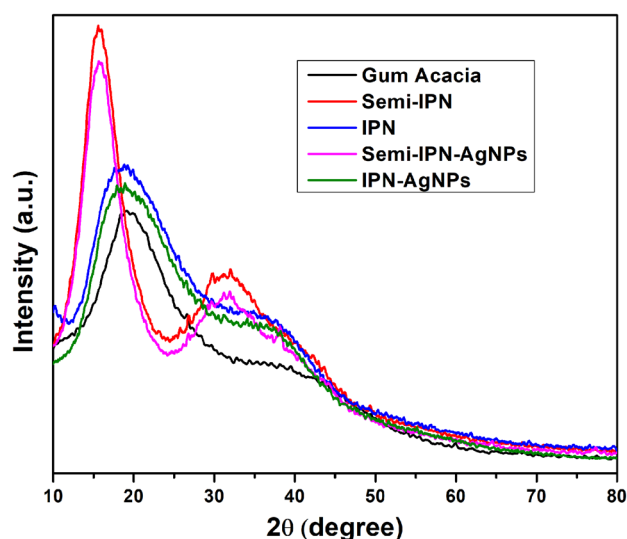


Fig. 5 XRD patterns of semi-IPN, IPN and their nanocomposites with silver nanoparticles

The observed changes in peak intensity point out the interaction of Ag ions in the semi-IPN matrix, which shows the transformation of the semi-IPN network to the amorphous system. Furthermore, adding AA to the synthesized semi-IPN matrix causes a slight shift in the intensity of diffraction peak. The loading of AgNPs into the IPN matrix, on the other hand, exhibits the same behavior, while the intensity of the diffraction peak decreases slightly. These findings are in line with the results of the SEM.

TGA analysis

The thermal behaviour of the backbone, semi-IPN, IPN, and composite with AgNPs was investigated using thermal analysis. The various degradation stages of the hydrogels and their composites with AgNPs are depicted in Fig. 6. Table 2 shows the corresponding weight loss at various temperatures. Initially, the GA shows maximum and frequent weight loss in each step as compared to its cross-linked structure. This mass loss is credited to the loss of the hydroxyl group of GA as a water molecule that leads to a fast decomposition of the backbone. The TGA spectrum of GA-g-poly(MAA) and GA-g-poly(MAA-IPN-AA) showed good thermal stability. Initial weight loss of 20–25% in both semi-IPN and IPN at the range 250 °C is attributed to the removal of free water and unreacted monomers from the hydrogels network. The 2nd stage weight loss of 25–60% from 300 °C to 400 °C was due to the removal of loosely bounded water from polymer network and breakage of physical forces involved in the hydrogel formation [35]. The last stage weight loss of 60–90% from 400 °C to 600 °C was due to complete thermal destruction of carboxylic groups,

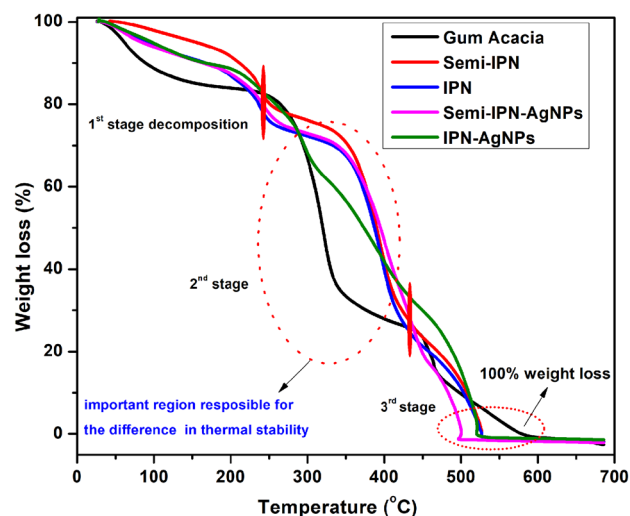


Fig. 6 TGA spectra of gum acacia, semi-IPN, IPN and their nanocomposites with silver nanoparticles

breakage of covalent bonds, splitting of backbone, and disruption of cross-linking of polymer networks [16, 43]. As expected, the thermal stability of IPN was further improved when loaded with the AgNPs as only 60–75% weight loss was only observed between 400 °C to 500 °C. Similar findings were reported by various authors [35, 51]. The improved thermal stability of the AgNPs incorporated semi-IPN and IPN may be because of two possible reasons [20]. Firstly, it is a well-known fact that AgNPs have higher thermal stability as compared to polymeric systems. Subsequently, the incorporation of AgNPs in the hydrogel matrix improves the thermal behaviour of the hydrogels. Secondly, more stability could be due to decreased mobility of the hydrogel chains on the impregnation of AgNPs. The AgNPs interact with poly(AA), poly(MAA) and GA through the -COOH and -OH groups present in the semi-IPN and IPN, as also clear by the outcomes of the FTIR spectra. This type of interaction causes weak intermolecular cross-linking between AgNPs and semi-IPN and IPN. Therefore, the degradation process can be slow down at lower temperatures, but it can be quick at higher temperatures after the AgNPs have been loaded [52]. The TGA results demonstrate that the synthesized hydrogel and their nanocomposites with AgNPs are more thermally stable.

In vitro antimicrobial studies

By using the agar well diffusion method, the antibacterial potential of GA, GA-g-poly(MAA), GA-g-poly(MAA-IPN-AA) and hydrogel composites with loaded AgNPs against *E. coli* (Gram-negative), *M. luteus* (Gram-positive) *P. aeruginosa* (Gram-negative), *R. species* (Gram-negative) and *S. aureus* (Gram-positive) bacteria was investigated

Table 2 Thermal analysis data of GA and crosslinked hydrogels

Sample name	Weight loss (%) at different temperature					
	250 °C	300 °C	350 °C	400 °C	450 °C	500 °C
GA	18	33	67	73	77	90
Semi-IPN	20	24	29	59	73	87
Semi-IPN-AgNPs	23	27	32	53	81	99
IPN	24	28	33	59	79	89
IPN-AgNPs	19	32	44	58	70	75

(Fig. 7). The inhibition zone diameter was measured in millimetres and is listed in Table 3. *M. luteus* is gram-positive bacteria that can be found in mucous membranes, water, and soil [53, 54]. This bacterium causes infections like septic shock and arthritis, endocarditis, meningitis, and cavitary pneumonia in immunosuppressed patients [54]. *E. coli* is well-known gram-negative bacterium remains present in aerobic commensal inhabitant of the large intestine and certain strains of *E. coli* cause diarrhea, and urinary tract infections [55]. *R. species* is gram-positive bacteria that is responsible for nitrogen fixation in leguminous plants [56]. Rhizobium has identified as the main contaminant of DNA extraction kit reagents and ultrapure water systems. *P. aeruginosa* is a Gram-negative, rod-shaped, asporogenous, and monoflagellated bacterium that causes plant and animal disease [57]. It causes severe infections in immunocompromised cancer patients and patients suffering from severe burns [57]. *S. aureus* is a major human pathogen causing multiple clinical infections like bacteremia and infective endocarditis, osteoarticular, skin and soft tissue infections, and pleuropulmonary [58]. The zone of bacterial growth inhibition was found to be 14 ± 0.5 mm, no inhibition, 16 ± 0.5 mm, 11 ± 0.5 mm and no inhibition against *E. coli*, *M. luteus*, *P. aeruginosa*, *R. species* and *S. aureus* with semi-IPN. In the case of AgNPs loaded semi-IPN composite, the zone of bacterial growth inhibition was bigger with a diameter of 16 ± 0.5 mm, inactive, 18 ± 0.5 mm, 13 ± 0.5 mm and 14 ± 0.5 mm against *E. coli*, *M. luteus*, *P. aeruginosa*, *R. species*, and *S. aureus* respectively. The zone of inhibition in case of IPN was found to be 20 ± 0.5 mm, 14 ± 0.5 , 23 ± 0.5 , 16 ± 0.5 mm, and 14 ± 0.5 mm against *E. coli*, *M. luteus*, *P. aeruginosa*, *R. species* and *S. aureus*, respectively. In case of AgNPs loaded IPN composite, the zone of bacterial growth inhibition was found to be more prominent and expanded i.e. 21 ± 0.5 mm, 13 ± 0.5 , 25 ± 0.5 , 17 ± 0.5 mm and 16 ± 0.5 mm against *E. coli*, *M. luteus*, *P. aeruginosa*, *R. species* and *S. aureus*, respectively. As can be seen from Fig. 6 and Table 3, after 24 h of incubation at 37 °C, all four types of samples showed an antibacterial effect on all the bacteria. The GA backbone shows antibacterial activity against *E. Coli* (11 ± 0.5), *P. aeruginosa* (8 ± 0.5) and *R. species* (8 ± 0.5). GA contains chiefly arabin, a mixture of calcium, magnesium,

and potassium salts of Arabic acid [19]. The Arabic acid on hydrolysis produces Lrhamnopyranose, Larabofuranose, galactopyranose, and taldobionic acid 6-dglucuronosido-d-galactose. The gum has also contained enzymes like oxidases, peroxidases, and pectinases. GA boosts the antimicrobial activities due to the presence of these bioactive agents like tannins, oxidases, peroxidases, pectinases, alkaloids, cardiac glycosides, flavonoids, volatile oils, saponins, and anthraquinone. The semi-IPN and IPN show excellent antibacterial activity against +ve and –ve strains.

The possible mechanism is explained in Scheme 2. The resulting semi-IPN and IPN produced charged hydrogels having stereo complexation and these complexes give strong fiber-like morphology as confirmed by SEM images that significantly enhanced antimicrobial activity against both Gram-positive/Gram-negative bacteria through interfacial stereochemistry [59, 60]. The antimicrobial mechanism involves lysis of cell walls by a blockage in the biosynthesis of peptidoglycan, making bacterial cells more susceptible to osmotic lysis. The GA and acrylates present in the hydrogels interfere with the nucleic acid metabolism, protein synthesis, and repair of bacterial cells leading to bactericidal effect. The cellular contents of both +ve and –ve bacterium, which is surrounded by an inner peptidoglycan cell wall as well an inner plasma membrane. Gram-negative bacteria possess outer lipid bilayer. The semi-IPN and IPN block the biosynthesis of the cell wall through weakening the peptidoglycan and compromising the structural integrity that causes disruption of bacterial cells [61, 62].

The incorporation of AgNPs to semi-IPN and IPN resulting in a composite having enhanced antibacterial activities against all five types of +ve and –ve bacterial strains. The AgNPs showed good dispersion ability throughout the hydrogel network. The bactericidal properties of AgNPs against bacteria have been a well-known fact for centuries. The Ag^+ ion can easily bind to the bacterial cell membrane due to its small size [63, 64]. This creates interaction between Ag^+ and thiol group of proteins on the bacterial cell membrane [25]. Hence, disturbing the bacterial cell's viability by obstructing the replication of DNA [64, 65]. AgNPs get easily diffused through bacterial cell membranes cause hindrance in the functions of respiratory

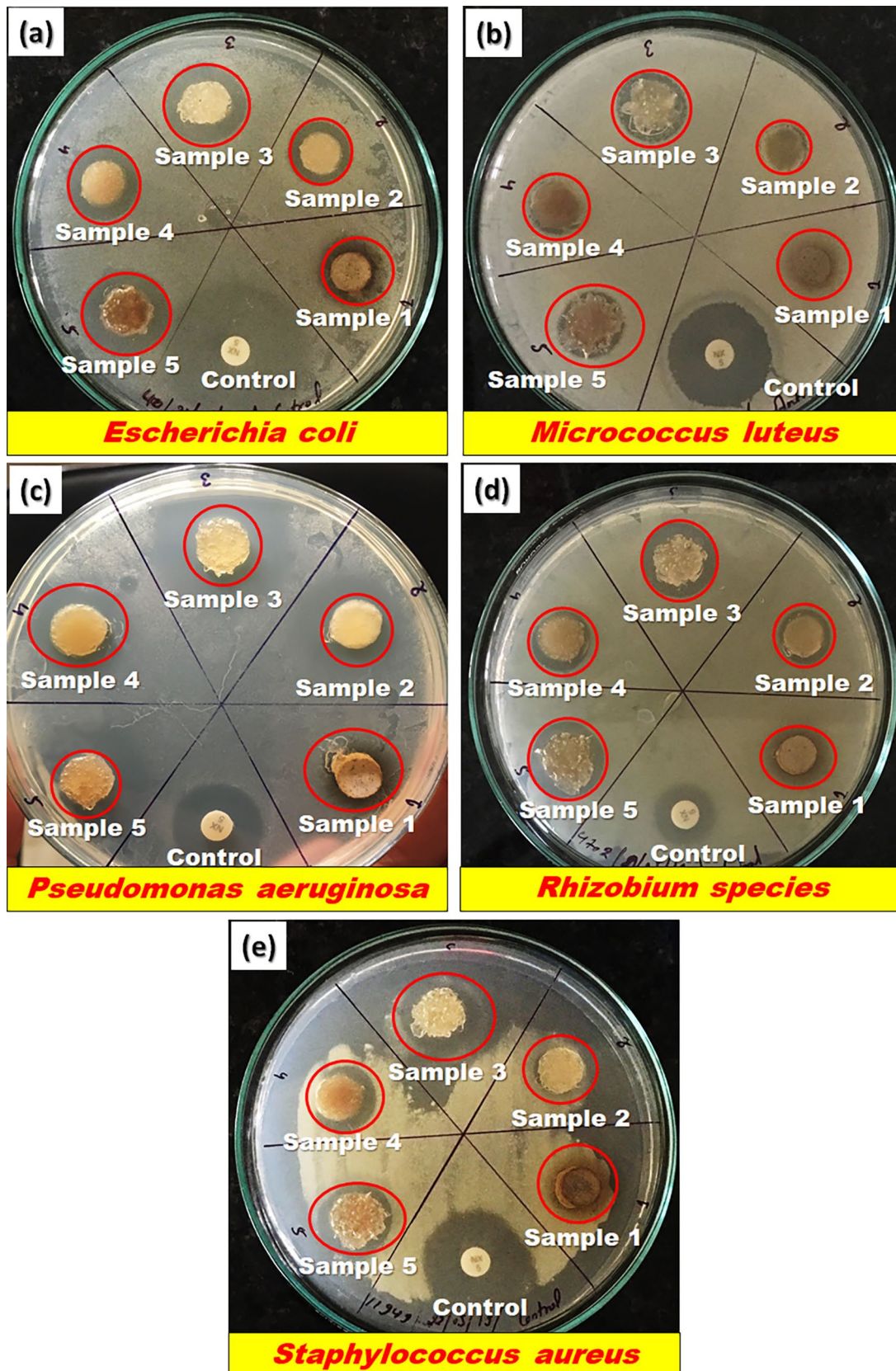
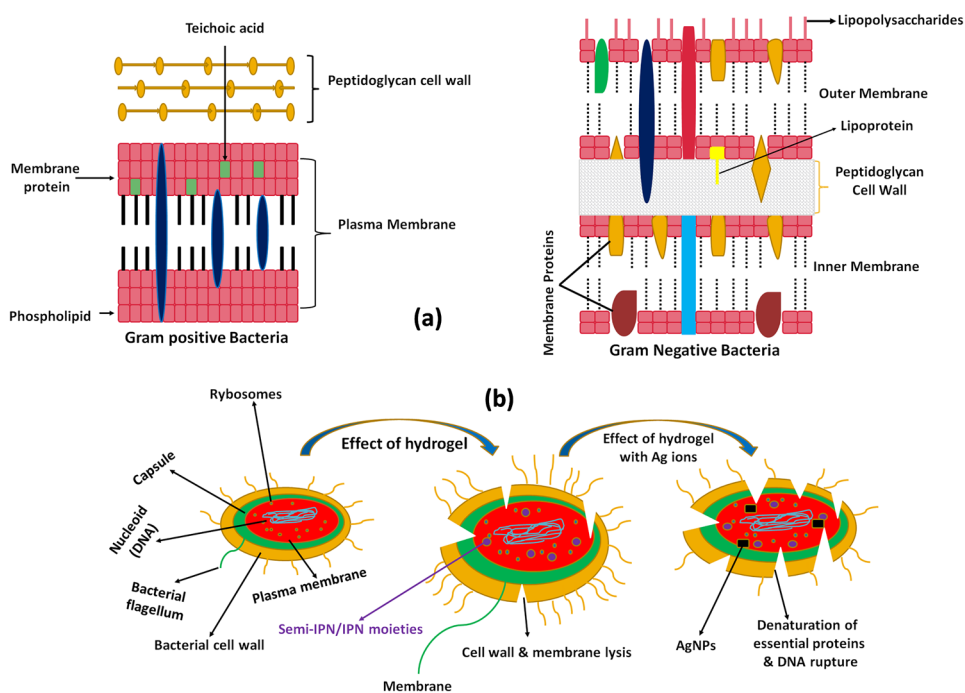


Fig. 7 Antimicrobial activities of the prepared samples with various bacterial strains

Table 3 Antibacterial activity of synthesized hydrogels against bacterium *Escherichia coli*, *Micrococcus luteus*, *Pseudomonas aeruginosa*, *Rhizobium species* and *Staphylococcus aureus*

S. No	Name of Bacteria	Inhibition zone diameter (mm)				
		Gum Acacia	Semi-IPN	IPN	Semi-IPN-AgNPs	IPN-AgNPs
1	<i>Escherichia coli</i> (Gram-negative)	11 ± 0.5	14 ± 0.5	20 ± 0.5	16 ± 0.5	21 ± 0.5
2	<i>Micrococcus luteus</i> (Gram-positive)	0 (inactive)	0 (inactive)	14 ± 0.5	0 (inactive)	13 ± 0.5
3	<i>Pseudomonas aeruginosa</i> (Gram-positive)	8 ± 0.5	16 ± 0.5	23 ± 0.5	18 ± 0.5	25 ± 0.5
4	<i>Rhizobium species</i> (Gram-negative)	8 ± 0.5	11 ± 0.5	16 ± 0.5	13 ± 0.5	17 ± 0.5
5	<i>Staphylococcus aureus</i> (Gram-positive)	0 (inactive)	0 (inactive)	14 ± 0.5	14 ± 0.5	16 ± 0.5

Scheme 2 Probable mechanism of antibacterial activity (a) cell membrane of gram positive and gram negative bacteria, (b) lysis of the cell membrane through its interaction with hydrogels and AgNPs**Table 4** Comparison of antibacterial activity of natural polysaccharide based hydrogels against various bacteria

S. No	Hydrogels	Bacteria	Inhibition Zone	References
1	Polyvinyl alcohol/gum karaya/ silver nanoparticles	<i>E. coli</i>	14.0	[69]
		<i>P. aeruginosa</i>	14.5	
		<i>S. aureus</i>	14.5	
2	Guar Gum-2/curcumin-AgNPs 3.14 nM hydrogel	<i>E.coli</i>	13.5	[70]
		<i>P. aeruginosa</i>	13.0	
		<i>S.aureus</i>	15.0	
3	PVA-GG hydrogels with 4 µg/mL AgNPs	<i>E. Coli</i>	11.5	[71]
4	Guar gum/P70) containing cinnamaldehyde-loaded chitosan nanoparticles	<i>S. aureus</i>	14.3 ± 1.12	[72]
5	PVA/PANI/Ag nanocomposites (at 15% AgNPs)	<i>E. coli</i>	12	[73]
		<i>Staph. aureus</i>	15	
6	Bacterial cellulose/silver nanocomposite hydrogels	<i>S. aureus</i>	26 ± 0.35	[74]
		<i>E. coli</i>	28 ± 0.16	

chain proteins and transport proteins [66]. The AgNPs even in small concentrations produces excess proton leakage throughout bacterial cell membranes causing loss of proton motive force [67]. The interaction of AgNPs with the dissolved oxygen produced reactive oxygen species that resulted in oxidative stress and free radical attacks onto the membrane lipids leads to the breakdown of the cell membrane that results in final death of bacteria [66, 68]. Table 4 compares the antibacterial activity of similar hydrogels systems against various bacteria.

Conclusion

Antimicrobial materials have been studied extensively in response to rising microbial resistance to traditional antibiotics and disinfectants. Natural polysaccharide-based hydrogels and their composites containing metal nanoparticles could be suitable for antibacterial applications in some cases. Using a microwave irradiation approach, we were able to successfully synthesis GA-g-poly(MAA) and GA-g-poly(MAA-IPN-AA) based hydrogels from GA. APS and MBA being used as initiator and cross-linker, respectively. To achieve maximum %S of semi-IPN (588%) and IPN (500%), different parameters were optimized. The formation of cross-linked hydrogels and hydrogel composites with AgNPs was successfully demonstrated using FTIR analysis. The SEM investigation of hydrogel networks reveals a variety of morphologies, which could be attributed to structural differences between semi-IPN, IPN, and their composites with AgNPs. Semi-IPN, IPN and their composite showed superb antibacterial potential against *E. coli*, *M. luteus*, *P. aeruginosa*, *Rhizobium species*, and *S. aureus*, and may be used to treat a variety of infected effluents. Such interconnected hydrogels will give superior material properties, particularly in biomedical research, to prevent biomaterial-associated infections as well as general pathogenic invasions into the body. The synthesized hydrogel samples will lead to enhanced applications in the future, as well as a deeper knowledge of material interactions. The efforts to improve the property profile of cross-linked hydrogels are still on, with the hope of improving overall performance in terms of water uptake capacity, biocompatibility, and biodegradability.

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Declarations

Conflicts of interest The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this article.

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