

CONTINUING EDUCATION PROGRAM: FOCUS...

Functional imaging of cerebral perfusion



A. Krainik^{a,b,c,*}, M. Villien^b, I. Troprès^c, A. Attyé^{a,b,c}, L. Lamalle^c, J. Bouvier^b, J. Pietras^c, S. Grand^{a,b,c}, J.-F. Le Bas^{a,b,c}, J. Warnking^b

^a Clinique universitaire de neuroradiologie et IRM, CHU de Grenoble, CS 10217, 38043 Grenoble cedex, France

^b Inserm U836, université Joseph-Fourier, site santé, chemin Fortuné-Ferrini, 38706 La Tronche cedex, France

^c UMS IRMaGe, unité IRM 3T recherche, CHU de Grenoble, CS 10217, 38043 Grenoble cedex 9, France

KEYWORDS

Cerebral perfusion;
Neurovascular
coupling;
Cerebral
vasoreactivity;
Autoregulation;
Functional BOLD MRI

Abstract The functional imaging of perfusion enables the study of its properties such as the vasoreactivity to circulating gases, the autoregulation and the neurovascular coupling. Downstream from arterial stenosis, this imaging can estimate the vascular reserve and the risk of ischemia in order to adapt the therapeutic strategy. This method reveals the hemodynamic disorders in patients suffering from Alzheimer's disease or with arteriovenous malformations revealed by epilepsy. Functional MRI of the vasoreactivity also helps to better interpret the functional MRI activation in practice and in clinical research.

© 2013 Éditions françaises de radiologie. Published by Elsevier Masson SAS. All rights reserved.

Reminders about cerebral perfusion

The study of cerebral perfusion provides critical information to understand the functioning of the central nervous system and apprehend its dysfunction, among the main causes of morbidity and mortality in the West. In neurology and psychiatry, the identification of these microvascular pathophysiological disorders may provide information that may help to better characterize several diseases, or even assess individual vulnerability.

Above all, cerebral perfusion allows for the transfer of appropriate quantities of glucose and oxygen for the functional needs of the brain, while eliminating heat and some catabolites such as CO₂ [1]. Perfusion is a dynamic physiological phenomenon able to respond to changes in the homoeostasis of the vascularized organ and the entire body. Like any biological function, general and local factors are likely to not only modify its state of equilibrium but also its adaptive properties. This adjustment is both passive, due to the

Abbreviations: ASL, arterial spin labeling; BOLD, blood oxygenation level dependent; CBF, cerebral blood flow; CBV, cerebral blood volume; CVR, cerebral vasoreactivity; fMRI, functional MRI; MRA, magnetic resonance angiography; P_aCO₂, arterial pressure in CO₂; P_eCO₂, expiratory pressure in CO₂.

* Corresponding author. Clinique universitaire de neuroradiologie et IRM, CHU de Grenoble, CS 10217, 38043 Grenoble cedex, France.

E-mail address: akrainik@chu-grenoble.fr (A. Krainik).

mechanical characteristics of blood vessels, and active by the arteriolar vasoconstriction. The vasoconstriction controls the caliber of the arterioles, so as to maintain the blood supply during variations in neuronal activity (neurovascular coupling), cerebral perfusion pressure (autoregulation), capnia, oxygenation and pH (vasoreactivity) [1].

Cerebral perfusion may simply be characterized by the cerebral blood flow (CBF), defined by the volume of blood transiting by the mass of the cerebral parenchyma per unit of time (classically expressed in ml/100 g of cerebral parenchyma/min). Since the density of brain tissue is close to that of water, the mass is often converted into volume, allowing the CBF to be expressed as a percentage of the parenchyma perfused per second (s^{-1}) (Table 1). The direct measurement of this dynamic property is especially difficult. This has given rise to the development of multiple, more or less invasive, methods that can, more or less, be considered in man. Numerous indicators and analytic models, with their advantages and disadvantages, have thereby been proposed [1,2]. The initial methods, such as those developed by Kety and Schmidt with the inhalation of NO_2 [3], measured a global CBF based on the analysis of the concentration of the marker at the entry and exit from the vascular system. In imaging, the CBF is measured on the scale of a cerebral region (rCBF), or even the pixel of a digital image and the voxel with tomography techniques. The success, over the last decades, of the imaging that has been established due to a major reduction in the invasiveness of the procedures and an increase in the temporal and spatial resolutions, has led to assimilate the rCBF with the CBF.

The cerebral blood flow and the cerebral blood volume (CBV) are two physiologically, closely related parameters, since they depend on the variations in arteriolar resistance. The mechanics of the fluids proportionally links variations in the volume with the square root of variations in flow. In the neurosciences, this ratio is estimated by: $([V/V_0] = [F/F_0])^\alpha$

V_0 and F_0 representing the volume and flow at the initial state, and V and F the volume and flow at the final state. With an experimental modulation of the cerebral perfusion by CO_2 , α was estimated as being close to 0.40 in the animal [4–6]. In man, values of α between 0.29 and 0.73 have been reported with high regional disparities [7,8]. These heterogeneities are likely to reflect differences between the methods of measurement [9], the regional variability of the capillary density [10,11], the physiological mechanism used to modulate the perfusion [7,12,13], the time interval between the early variations in the CBF that are mainly based on the arteriolar and capillary sectors, and later variations in the CBV that better reflect the venous sector [6,14,15].

Physiological variations in cerebral perfusion

At rest, the cerebral perfusion decreases with age [16] and is significantly higher in women [17]. The cerebral perfusion is closely related to the activity of the brain. It is often measured in the neurosciences and in medicine since it reflects the interaction between the vessels and the neurons through neurovascular coupling. Moreover, the cerebral perfusion is maintained roughly constant in order to deal with variations in the blood pressure and intracranial pressure by autoregulation. The cerebral perfusion is also sensitive to variations in the arterial concentration in CO_2 and O_2 by the vasoreactivity (Fig. 1). All of these physiological functions are based on the vasoconstriction that enables, through the dilation and contraction of the vessels, to adjust the cerebral blood flow in order to guarantee the cerebral activity by dealing with the general and local physiological constraints [18].

Innervation of cerebral vascularization

Extrinsic innervation

The extrinsic vascular innervation of the pial arteries arrives from the peripheral nervous system by the upper cervical, sphenopalatine, trigeminal and optical ganglia that relay the information from peripheral baroreceptors. The extrinsic innervation is mainly sympathetic vasoconstrictor (noradrenalin, serotonin, neuropeptide Y) and parasympathetic vasodilator (acetylcholine, nitric oxide, VIP). The arteries progressively divide into arterioles that enter the cerebral parenchyma. They consist of an internal layer of endothelial cells, smooth muscle cells and an outer layer of leptomeningeal cells that form the external tunic. The arteriole is separated from the parenchyma by the Virchow-Robin space that contains the cerebrospinal fluid and is bordered by astrocytes on the outside. As the arterioles enter the parenchyma, the fluid space disappears and the arterioles and then the capillaries are in direct contact with the feet of the astrocytes that form the glia limitans [19–21].

Intrinsic innervation and the neurovascular unit

The intrinsic innervation of the intra-parenchymatous arterioles by the central nervous system relies on cortical interneurons and the efferences of subcortical neurons

Table 1 Basal cerebral perfusion values usually admitted in man [1].

Mean CBF	$\geq 60 \text{ ml}/100 \text{ g}/\text{min}$ $\geq 60 \text{ ml}/100 \text{ ml}/\text{min}$ (parenchyma density = 1.04 g/ml) $\geq 1 \text{ ml}/100 \text{ ml}/\text{s}$ $\geq 0.01/\text{s}$ (or 1% of the parenchyma corresponds to the fresh blood each second)
CBF grey matter	80 ml/100 g/min
CBF white matter	20 ml/100 g/min
Density grey matter / Density white matter	≥ 1
CBV	4 ml/100 g $\geq 4\%$ of the parenchyma $\geq 60 \text{ ml}$ of blood for a 1500 g brain
Arterial / capillary / venous CBV [280]	21% / 33% / 46%

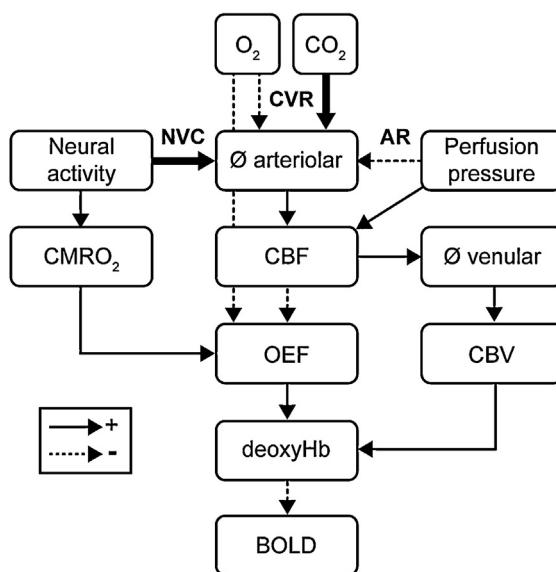


Figure 1. Functional variations in cerebral perfusion and BOLD signal. The full arrows represent a positive relationship and the dotted arrows a negative relationship. The arteriolar caliber is modulated by the neurovascular coupling induced by the neuronal activity, the vasoreactivity to O_2 and, above all, CO_2 , the autoregulation of the perfusion pressure (that is, the difference between the mean arterial pressure and the intracranial pressure). The increase in the cerebral blood flow (CBF), higher than the cell needs, reduces the oxygen extraction fraction (OEF) and increases the cerebral blood volume (CBV) which reduces the deoxyhemoglobin content (deoxyHb) in the voxel. Since the deoxyHb is paramagnetic, it shortens the T2. This reduction increases the signal in imaging sensitive to the BOLD effect.

coming from the nucleus basalis of Meynert and the frontobasal region (acetylcholine), the locus coeruleus (noradrenalin) and the raphe nuclei (serotonin). The action of the GABAergic cortical interneurons may contract or dilate the vessels. Whereas GABA is a vasodilator, the activity of these interneurons is modulated by a specific subcortical innervation. The subcortical efferences directly innervate the vascular wall and the astrocytes. Other vasomotor neurotransmitters such as dopamine coming from the ventral tegmental area (black matter) may also act by direct vascular innervation or through the innervation of the astrocytes (Table 2) [22,23].

At the capillary level, the structural association of the pericytes edged by the feet of the astrocytes innervated by cortical interneurons and subcortical neurons form a "neuron-astrocyte-vasculature tripartite functional unit", more commonly called "neurovascular unit" [19]. The essential role of the astrocytes in the vasodilation induced by neuronal activity has recently been demonstrated [24–26]. In addition, the integrity of the glia limitans is required for arteriolar vasodilation [27].

Neurovascular coupling

Cerebral perfusion is closely related to neuronal activity. This global and local adaptation of cerebral perfusion extensively relies on the perivascular innervation and the neurovascular unit. Therefore, the perfusion is currently

Table 2 Main vasoactive substances [20].

	Vasomotor effect
Ions	
K^+	Dilation
H^+	Dilation
Metabolites	
CO_2	Dilation
O_2	Constriction
Adenosine	Dilation
Neurotransmitters	
Acetylcholine	Dilation
Noradrenalin	Constriction
Serotonin	Constriction
Dopamine	Dilation (D1/D5 receptor) Constriction (D2/D3/D4 receptor)
GABA	Dilation
Other	
NO	Dilation

measured in the neurosciences and in medicine since it is the direct reflection of the interaction between the vessels and the neurons.

The increase in perfusion is related to the energy activity of the brain. In man, 75% of the energy intake is consumed by post-synaptic activity, 7% by pre-synaptic activity, 10% by the neuron action potential, 6% by the astrocyte activity and 2% by the transmembrane resting potential [28]. The neuron energy metabolism is based on the use of the oxygen and glucose carried by the blood to the capillaries [29]. The increase in synaptic activity results in a reduction in the oxyhemoglobin (HbO_2) concentration and an increase in that of deoxyhemoglobin (deoxyHb). This initial dip in blood oxygenation before the hemodynamic response may be the most reliable marker of the neuronal activity [30], although its demonstration using BOLD fMRI is unpredictable.

Functional hyperemia occurs 1 to 2 s after the beginning of the neuronal activity. Although the local increase in CBF is proportional to the glucose consumption [31], the oxygen intake is abnormally high compared with the moderate increase in its capillary extraction. In addition, the hyperemia covers a wider zone than that of the neuronal activity [32]. The time before the hemodynamic response, the inadequacy between the needs in glucose and the oxygen intake and the early release of lactate have suggested that the glycolysis may initially be anaerobic by the use of the glycogen delivered by the astrocytes, and more quickly useable than the capillary glucose that will be used subsequently. Therefore, the release of glutamate in the synaptic space would lead to an astrocyte recapture that metabolizes it into glutamine by anaerobic glycolysis, then delivered to the neuron. As for the lactate, it would be aerobically metabolized by the neuron to supply the ATP required for ionic re-equilibrium following the neuronal depolarization [29,33].

These elements underline the close anatomo-functional relationship between the neurons, vessels and astrocytes in neurovascular coupling. Its proper operation relies on the structural integrity of the constituents (anatomic,

histological and cellular), an appropriate release and concentration of vasoactive agents, as well as the integrity of their action mechanisms.

From this, the idea progressively emerges that there may be diseases of the neurovascular unit, such as Alzheimer's disease [19,20,34–37].

Autoregulation of cerebral pressure

Autoregulation helps maintain the cerebral pressure significantly constant by adjusting the arteriolar dilation according to the variations in the perfusion pressure (PP), defined by the difference between the mean arterial pressure (MAP) and the intracranial pressure (ICP), $PP = MAP - ICP$. When the perfusion pressure increases due to an increase in arterial pressure, the autoregulation induces a vasoconstriction, in order to avoid the risk of the rupture of the brain-blood barrier (BBB) and tissue edema, or even hemorrhage. When the perfusion pressure decreases due to a drop in the arterial pressure, the autoregulation induces vasodilation in order to avoid the risk of ischemia. When the perfusion pressure decreases due to an increase in the intracranial pressure, the autoregulation induces vasodilation to avoid the risk of ischemia.

The autoregulation that adjusts the arteriolar resistances according to the transmural pressure gradient is maintained by the vascular muscle response to the perfusion pressure by the myotatic reflex and the extrinsic vascular innervation. The limits of the autoregulation may be modified by the sympathetic activity, the arterial pressure in CO_2 , chronic arterial hypertension and certain drugs [38–40].

The monitoring of the autoregulation is very important in medicine, in particular in traumatic disorders, since the loss of autoregulation and vasoreactivity to CO_2 are elements pointing to a poor prognosis [38,39,41–45].

Vasoreactivity to circulating gases

Vasoreactivity to CO_2

The effects of CO_2 on cerebral vascularization have been observed for a long time and the inhalation of 5 and 10% volume fraction CO_2 are accompanied by a 50 and 100% increase in CBF, respectively [46]. The CBF variation curve as a function of $P_a\text{CO}_2$ is a sigmoid with an approximately linear portion for the physiological values of the CBF. As of normocapnia ($P_a\text{CO}_2$ = about 40 mmHg), the hypercapnia is accompanied by an increase in the CBF and CBV of 6% and 2%/mmHg de $P_a\text{CO}_2$, respectively, while the hypocapnia is accompanied by a reduction in the CBF and CBV of 3% and 1%/mmHg $P_a\text{CO}_2$ respectively [7,47–49]. The transcranial Doppler measurement of the arterial speed is used to calculate the CO_2 vasoreactivity index, which corresponds to the slope of the curve that describes the mean arterial speed as a function of the expiratory pressure of CO_2 . In healthy subjects, this index is close to $1.5 \text{ cm.s}^{-1}.\text{mmHg}^{-1}$ [50]. This vascular response predominates on the small caliber arterioles – 40–100 μ [51,52] – but all vessels, including the capillaries and venules are involved [51,53].

Several mechanisms have been proposed to account for the CO_2 vasoreactivity. It seems that the modification in the perivascular pH, induced by the free passage of CO_2

through the BBB and the production of H^+ ions ($\text{CO}_2 + \text{H}_2\text{O} \rightarrow \text{H}^+ + \text{HCO}_3^-$), plays a preponderant role [54,55], since HCO_3^- is used for the clearance of excess lactic acid produced by the neurons and glial cells [54]. The role of NO in the CO_2 vasoreactivity has also been proposed [56]. In physiological conditions, its action is modest [18]. However, in pathological conditions (diabetes, AHT), the endothelial dysfunction and the poor distribution of NO account for the alteration in the CO_2 vasoreactivity of patients. The administration of an "NO donor", sodium nitroprussiate, would restore the CO_2 vasoreactivity [50]. The role of the prostaglandins has been demonstrated in certain animal species, but not in man [18].

Physiological variations have been described with a reduced vasoreactivity in women [57], the elderly subject [58] and during the early hours of the day [59,60].

O_2 vasoreactivity

The vascular effects of O_2 , as opposed to those of CO_2 , have also been known for several decades [61]. Hypoxemia accompanies an increase in the CBF, whether due to hypoxia (by a reduction in the O_2 intake), anoxia (by the inhalation of carbon monoxide (CO) that saturates the Hb in HbCO) or anemia (by a reduction in the functional Hb concentration). However, this effect is observed for pathological values of $P_a\text{O}_2$ under 60 mmHg (threshold of respiratory insufficiency), while the normal $P_a\text{O}_2$ is close to 90 mmHg [18,49,62]. The mechanisms responsible for the cerebral vasodilation with hypoxia are still poorly known. Vascular innervation and the peripheral baro or chemoreceptors are not involved. However, the ventrolateral region of the bulb is sensitive to hypoxia and its destruction alters the vasodilation at hypoxia without modifying the CO_2 vasoreactivity [63]. Another analogous, more caudal region has also been described. Adenosine may be the mediator of this vasodilation during mild hypoxia while the H^+ and K^+ ions may play a role in case of severe hypoxia [18,64,65]. Finally, chronic hypoxia provokes structural modifications in the microvascular walls and angiogenesis that reduces the intercapillary distance [66]. In a recent experimental study carried out in adults exposed to prolonged hypoxia by residing at 4400 m, we observed an increase in the cerebral perfusion associated with an alteration in the vasoreactivity [67].

Normobaric hyperoxia, obtained by the inhalation of 85 to 100% O_2 , induces a vasoconstriction responsible for a 7 to 31% reduction in the CBF [61,64,68–71], or even an increase in the CBF [72]. The effects of hyperoxia on cerebral perfusion are significant for a $P_a\text{O}_2$ superior to 300 mmHg [61]. The reduction in the $P_a\text{CO}_2$ concomitant with the inhalation of O_2 may increase the reduction in CBF. The vascular effects of hyperoxia would be due to the inactivation of the NO-dependent vasodilation by an increase in the O_2^- free radicals [73].

Functional imaging of cerebral perfusion

The functional imaging of cerebral perfusion may be defined as the study in imaging of the functional variations in cerebral perfusion induced by vasomotor stimuli. We have seen that the perfusion was physiologically modified by the neuronal activity (neurovascular coupling), the intracranial

perfusion pressure (autoregulation), the CO₂ level in the blood and, to a lesser extent, the O₂ level (vasoreactivity) (Fig. 1). According to the temporal resolution of the techniques, we have measurements at the state of equilibrium, or of the hemodynamic response to the vasomotor stimulus over time.

Influence of the initial conditions

The functional variations of cerebral perfusion may be influenced by the initial conditions. Insofar as the basal cerebral perfusion reflects the initial dilation of the arterioles and, as long as the structural and functional integrity of the arteriolar system is preserved, we can intuitively think that the variation in perfusion induced by a vasomotor stimulus is inversely proportional to the initial perfusion. This functional aspect of the perfusion was above all studied with neurovascular coupling. The polemics that opposes the proportional model [74–79] with the additive model that defends the independence of the variations in perfusion of the initial conditions [80–85] has not been solved [72,86]. Increases in the BOLD signal and the CBF tend towards maximum values. The extreme experimental conditions required to reach them are far from the physiological conditions. This does not guarantee the validity of the physiological models usually used in such conditions [72]. Even pharmacological challenge using neuromediator (e.g., L-DOPA a regional vasodilator) failed to demonstrate significant change of the hemodynamic response to hypercapnic stimuli using BOLD contrast [87].

Although, the relationship between the basal conditions of perfusion and the hemodynamic response to the CO₂ vasoreactivity has not been studied as extensively as that of the neurovascular coupling, it seems necessary to have a good assessment of the basal cerebral perfusion. Ideally quantitative, this measurement may be semi-quantitative in patients with anatomic hypotheses.

Imaging of the neurovascular coupling

Imaging of neurovascular coupling is one of the main methods in functional neuroimaging. Among other techniques, BOLD fMRI contrast is based on the modifications in cerebral oxygenation following that of the synaptic activity and, above all, that of the perfusion that is induced by the neurovascular coupling (Fig. 1). This approach privileges the control of the neuronal activity that determines the regressor of an observed vascular signal. The changes in the hemodynamic response are therefore interpreted as the consequence of those of the neuronal activity. The close relationship between the synaptic activity and BOLD signal, demonstrated in the animal, its non-invasive nature and reproducibility, have made the fMRI become, within several years, the most used technique in functional neuroimaging [88,89].

The neuronal activity associated with a cognitive task is accompanied by an arteriolar vasodilation and an increase in the regional cerebral blood flow (rCBF), leading to vasodilation of the venules and veins due to their elasticity (balloon model) [90,91]. In addition, the exaggerated increase in oxygen intake in the blood flow induces hyperoxygenation of the venous blood, reflected by an increase in the oxyhemoglobin

concentration and a reduction in the deoxyhemoglobin concentration. Now, deoxyhemoglobin is paramagnetic. By altering the homogeneity of the intra and perivascular magnetic field, the deoxyhemoglobin decreases the intensity of the free induction decay observed in T₂^{*}-weighted gradient echo that is a true endogenous contrast agent. Thereby, in response to neuronal activity, the reduction in the deoxyhemoglobin concentration is accompanied by an increase of several percentages in the T₂^{*}-weighted signal (Fig. 1) [89,92].

The estimate of the neuronal activity from the analysis of the BOLD signal is based on the hypothesis that their relationship is approximately linear, which does not exclude complex non-linear relationships and interactions between the different elements in this relationship. This hypothesis was backed up by several experiments coupling fMRI with different electrophysiological methods [88].

In spite of many approximations, the location of the neuronal activity by BOLD fMRI was experimentally validated in the animal [88] and in clinical practice by the comparison with the evoked potentials [93], the magnetoencephalography [94], the pre- and per-surgical stimulations [95–97] and lesion studies [98–100].

However, the relationship between the neuronal activity and the BOLD signal is very indirect and is influenced by multiple, often interdependent, physiological factors [101,102]. The BOLD signal has been shown to be sensitive to the quality and intensity of the neuronal response to the stimulus [103,104], to the basal conditions of perfusion [74–76,80,85], to the blood oxygenation [105–107], to the capnia [108], to the vasoreactivity [109–112]... In addition, it is advisable to add variations in the general factors to these sources, that may influence them such as age [58,75,113–115], sex [17], the consumption of substances such as caffeine [116–118], alcohol [119], neuroleptics [120,121], apomorphine in patients with Parkinson's disease s [122], anesthetic products [123,124] or products modulating the activity of neurotransmitters [125].

In pathological conditions, a modification in the BOLD signal was observed in the presence of transient arterial hypertension [126,127], arterial stenosis [111,128–130], micro-angiopathy [110,128], migraine [131], multiple sclerosis [132], Alzheimer's disease [75,133] and near a macroscopic lesion such as a stroke [109,110,134], tumor [112,135–140], arteriovenous malformation [137,141,142]. The pathophysiology of these BOLD signal variations is highly varied, insofar as these disorders may have an overall or locoregional influence on the basal CBF, neurovascular coupling and vasomotricity.

Several alternatives have been proposed for a better estimate of the neuronal activity in MRI. However, they are based on the measurement of the vascular effects of the neuronal activity. Perfusion imaging by arterial spin labeling allows for the dynamic and repeated measurement of the variations in perfusion induced by cognitive activity, with an excellent topographic characterization. However, the signal-noise ratio remains low [143–145]. The quantitative BOLD fMRI allows for the calculation of the oxygen consumption by the neurons, by associating the conventional BOLD fMRI with a conventional and simultaneous measurement of the BOLD and CBF signals modulated by a vasodilation test using CO₂ inhalation [81,146,147].

Therefore, to interpret the BOLD fMRI of the neurovascular coupling, it's important to have an estimate of the basal value and the functional variations in the cerebral perfusion (Fig. 2). This aspect becomes especially essential in EEG-fMRI. In fact, the modifications in the hemodynamic response to abnormal neuronal activity in epileptogenic zones should be taken into account in order to identify these regions. Without these adjustments, the risk of localization error would be too high to consider the clinical use of the EEG-fMRI [148].

Imaging of autoregulation

The autoregulation of perfusion pressure is an important function to monitor when there is intracranial hypertension, in particular during a serious traumatic brain injury. Taking into account the risks related to the transport of heavily

equipped subjects outside of intensive care, the autoregulation may be assessed with the transcranial Doppler [38,41,45,149]. To experimentally modulate the perfusion pressure, compression cuffs may be used on the thighs. A systemic hypotension is provoked by suddenly releasing the pressure. The autoregulation then induces vasodilatation in reaction to the drop in perfusion pressure [150].

However, studies on the imaging of autoregulation are rare and remain experimental [126].

Imaging of vasoreactivity

The imaging of cerebral vasoreactivity is another approach for the functional imaging of cerebral perfusion. As opposed to the imaging of neurovascular coupling that privileges the study of the neuronal activity, the imaging of the cerebral

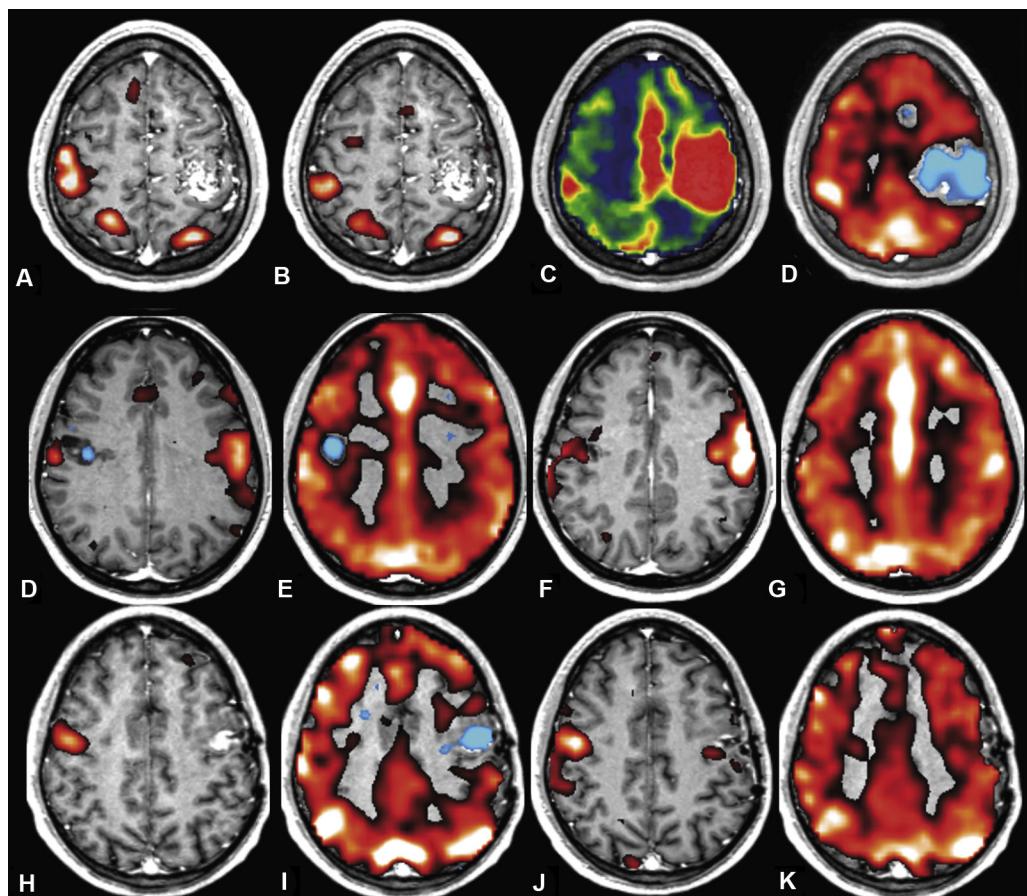


Figure 2. False negatives in fMRI activation induced by modifications in perfusion. In fMRI, an increase in the BOLD contrast is expected near the functional cortex. If absent, an alteration in the perfusion and its functional properties may be the cause and lead to negative results. A–D. Patient with a left precentral arteriovenous malformation. The movements of the left hand are accompanied by a right primary sensorimotor activation (A), while the movements of the right hand are not accompanied by a left primary sensorimotor activation (B). The mapping of the cerebral blood volume reveals locoregional hyperperfusion due to the lesion (C), with inverted vasoreactivity in projection of the precentral cortex (D). E–H. Patient with a ganglioglioma of the right precentral gyrus without facial palsy. Before surgery (E, F), inverted activation is detected in projection from the primary motor cortex next to the tumor (E), related to an inversion of the vasoreactivity (F). After the surgery, there is a pseudo-recovery of the activation (G), with normalization of the vasoreactivity (H). I–L. Patient with a recurrence of xanthoastrocytoma of the left precentral gyrus, manifested by seizures of the right hemiface. No pre-surgical facial palsy. Before surgery (I, J), the movements of the lips did not allow the activation of the ipsitumoral primary sensorimotor cortex, as opposed to the healthy hemisphere (I). The mapping of the vasoreactivity to hypercapnia shows an extensive positive vascular response in red, with an inverted response around the lesion in blue, masking the functional cortex (J). The post-surgical fMRI shows pseudo-recovery of the ipsitumoral primary sensorimotor activation (K), associated with normalization of the vasoreactivity (L) [139].

vasoreactivity privileges the study of a non-neuronal vaso-motor stimulus, such as CO₂ or acetazolamide.

In normal conditions, the vasoreactivity is influenced by sex [151], age [58, 152], arterial pressure [153], basal perfusion conditions [106, 152, 154, 155], anesthetic [106, 156], modulation of the action of vasoconstrictor neuromediators, such as that shown for acetylcholine [157, 158], but not for noradrenalin [159] or dopamine [87].

The vasoreactivity is greater in the grey matter than in the white matter [9, 84, 108, 160–166]. Within the grey matter, there are regional variations that may be under the influence of variations in the microvascular density [10, 11] and the heterogeneous distribution of vasoconstrictor neuromediators [101]. However, the published results often differ and do not provide an exact idea about these regional variations. Therefore, the vasoreactivity of the cerebral cortex is greater than that of the cerebellum [163, 164] or inversely [7], than that of the basal ganglia [163, 164, 167] except for the thalamus [166] or inversely [168]... Differences have also been reported within the cerebral cortex, with a more marked vasoreactivity in the occipital and frontal lobes [163, 164] although this is not constant [166, 169].

In pathological conditions, global alterations in the hemodynamic response have been observed with chronic arterial hypertension [50, 170], diabetes [50, 171–173], sepsis [174], the administration of certain antihypertensive drugs [175], opioid agonists [176], in the presence of diffuse intracerebral factors such as microangiopathy [128, 177–179], transient global amnesia [154], CADASIL [180, 181], normal pressure hydrocephalus [182], traumatic brain injury [38, 39, 41–45, 183, 184], Alzheimer's disease [185–189]... Locoregional variations are also observed downstream from arterial stenosis [111, 128, 129, 190–198], in focal epilepsy [199–201] and near a macroscopic lesion, such as a stroke [109, 110, 202], an arteriovenous malformation (Fig. 2) [141, 203] or a tumor [112, 139, 140].

Value of the measurement of the cerebral vasoreactivity

Value of neurovascular coupling for functional imaging

The different functional imaging techniques for cognition, such as BOLD fMRI, are mainly based on neurovascular coupling. However, in spite of obvious modifications in the BOLD signal in patients, almost all of the studies do not take into account possible intra- and inter-individual variations in the physiological mechanism at the origin of the measured signal. During the interpretation of the BOLD signal, it is therefore impossible to distinguish the specific effects of the variations in neuronal activity from variations in the hemodynamic response or a possible interaction between these two factors. This represents an obvious limit to the technique that has to be overcome in order to better interpret fMRI results [101, 109, 112, 139, 140].

The evaluation of the hemodynamic response is obtained by the test of vasoreactivity to CO₂, even though the mechanisms involved differ. In spite of this physiological approximation, the normalization of the neurovascular

BOLD signal by a hypercapnia test is at the base of the quantitative fMRI [76, 162, 204]. This approach consists of simultaneously measuring the BOLD signal and the cerebral blood flow. These parameters provide, by hypercapnia calibration of a biophysical model of the BOLD signal, the relative quantification of the consumption of oxygen by the cerebral parenchyma [81, 146, 147, 205]. From then, this method, in the cognitive neurosciences, allows for comparison of the intra- and inter-individual data to better estimate the modifications induced by aging, neurodegenerative disease, disabling focal cerebral lesions (epilepsy, tumor and removal, ischemic stroke...).

In clinical practice, this approach would allow for a better interpretation of the functional mapping, in particular in patients before surgery, by taking into account alterations in the vasoreactivity near the lesions (Fig. 2) [112, 139, 140].

Prognostic value in steno-occlusive disease

In patients monitored for steno-occlusive disease of the cervical arteries, the measurement of the vascular reserve in the territory of the damaged artery is used to assess the risk of an ischemic stroke (CVA) or a transient ischemic attack (TIA). Downstream from an occlusion or severe stenosis, the reduction in perfusion pressure is compensated by the autoregulation and vasodilation that help maintain the constant perfusion. The absence of additional vasodilation attests to the absence of vascular reserve to deal with an increase in perfusion required to compensate the reduction in the blood supply (increase in the stenosis, reduction in the cardiac output or the arterial blood pressure), thereby exposing the patient to arterial ischemia [190, 206–215]. Therefore, the study of the vascular reserve offers a functional evaluation of the functionality of the vascular network downstream from the stenosis and the available collaterality [216, 217].

The clinical value of this imaging may be determinant in patients with tight intracranial arterial stenosis in which the treatment is highly debated. In fact, following an initial ischemic episode, the risk of recurrence under medical treatment may attain 30 to 40% after 2 years, or even 60% when there are clinical signs of hemodynamic disorders [218]. Currently, endovascular treatment by stent is considered only following a new ischemic episode. This being so, the morbidity related to this treatment has called its justification into question [219]. Thereby, the imaging of the vascular reserve may better characterize the nature of a CVA/TIA in these patients. A reserve maintained downstream from a stenosis would be in favor of a thromboembolic CVA/TIA, while an altered reserve may more reveal a CVA/TIA of hemorrhagic origin [214]. In patients with tight intracranial arterial stenosis, this imaging may identify patients at risk of a failure of a well carried out medical treatment and help better select patients likely to benefit from endovascular treatment (Figs. 3 and 4) [198].

The imaging of cerebral vasoreactivity shows an improvement, or even normalization, of the vascular reserve after the surgical treatment of a carotid stenosis [197, 220, 221], moyamoya disease [195, 222–226] or tight intracranial arterial stenosis (Fig. 4).

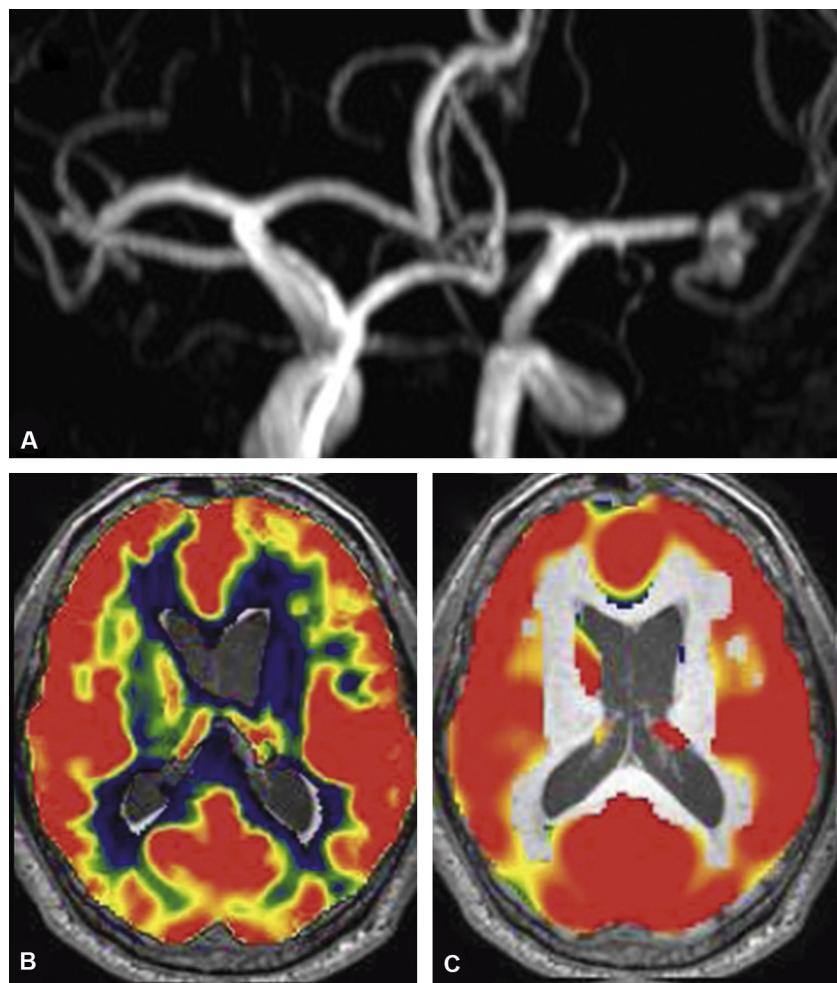


Figure 3. Tight intracranial arterial stenosis without reduction of the vascular reserve. A. A 44-year-old man treated for left capsulo-lenticular infarction revealing tight stenosis of the termination of the first portion of the left middle cerebral artery, accompanied by a post-stenotic aneurism. B. The basal perfusion is normal. C. The vasoreactivity does not show an alteration in the vascular reserve in the territory downstream from the stenosis. Most likely, it consists of a thromboembolic ischemia. A deficiency recurrence was not noted under well carried out anti-platelet therapy. The aneurism was surgically treated.

Towards imaging of the vasoreactivity for diagnostic purposes

The global and regional variations in the cerebral vasoreactivity observed in patients may reveal modifications in the basal cerebral perfusion [5,8,74], mechanical vascular properties [180,227], concentration of vasomotor substances in the perivascular environment, relations between the vessels, astrocytes and neighboring neurons [101]. However, a great many cerebral and cerebrovascular disorders are likely to be accompanied by global and locoregional alterations in vasoreactivity, as shown in patients [82,109,111,112,128,177,186,192,228] and experimental animal models [27,158,180,229–233].

The potential fields for the application of the imaging of the vasoreactivity for diagnostic purposes are extremely vast. However, the imaging of the vasoreactivity is only used in case of occlusive arterial disease, in order to assess the vascular reserve of the cerebral parenchyma and the risk of ischemic stroke. The cerebral imaging of the vasoreactivity may be used to search, in the entire brain,

infracentimetric spatial anomalies of the functional modulation of the cerebral perfusion. It would be possible to estimate the sensitivity and specificity of these anomalies by comparing the results with those obtained in controls. For diagnostic purposes, this imaging of the microvascular function and its interactions with the metabolic, neuronal and glial perivascular environments may offer a better phenotypic characterization of neurological and psychiatric diseases, or even an estimate of the individual vulnerability to these disorders even before the appearance of clinical signs or macroscopically identifiable lesions in imaging. In the same way that the injection of contrast product offers a functional contrast, the imaging of the vasoreactivity may offer a functional contrast that should be put to use for medical purposes.

In neurodegenerative disease, three recent studies have shown alterations in the vasoreactivity in Alzheimer's disease and patients at risk [186,188,189], by underlining the importance of microvascular disorders in the physiopathology of the disease, as suggested by the data in the animal [234–237]. On the other hand, in an analogous study carried

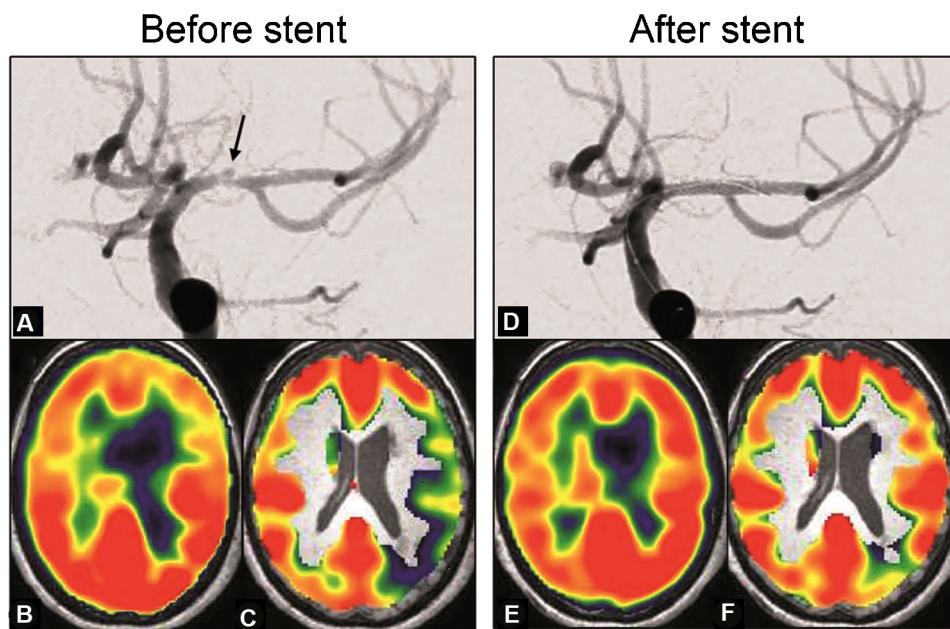


Figure 4. Tight intracranial arterial stenosis with reduction of the vascular reserve [198]. A. A 63-year-old woman treated for tight stenosis of the M1 portion of the left middle cerebral artery (arrow), revealed by a deep ischemic stroke. A–C. Pre-surgical examinations. D–F. Post-surgical examinations. A and D. The angiography shows tight stenosis before (A) and after dilation by stenting (D). B and E. Basal perfusion MRI measured by arterial spin labeling. The basal perfusion is normal and globally symmetrical, both after and before the intervention. C and F. BOLD-fMRI of the vasoreactivity to hypercapnia. The fMRI of the vasoreactivity to CO₂ shows a major alteration in the vascular reserve in the territory of the arterial stenosis (C). It obviously is a hemodynamic ischemia. Except for the posterior junctional territory, the intervention normalized the vascular reserve (F).

out in patients monitored for idiopathic Parkinson's disease, we did not find a significant alteration in the cerebral vasoreactivity (Fig. 5) [87]. In this context, the normality of this examination may be a diagnostic criterion to distinguish it from vascular parkinsonism.

In epileptology, perfusion anomalies have been described outside of seizures, both in the irritative cortical zones and in the non-epileptic zones [200,238]. In focal epilepsy, interictal basal hypoperfusion has been reported with a cortico-thalamic diaschisis [239]. More surprisingly, an increase in the perfusion of the epileptogenic cortex and hypoperfusion of the healthy cortex has been identified up to 10 min before the occurrence of a seizure [240,241]. Beside

these basal perfusion anomalies, a reduction in the perinidal vasoreactivity has been detected in patients with arteriovenous malformations and epilepsy whereas the vasoreactivity was normal in the absence of epilepsy (Fig. 6) [242].

In neuro-oncology, the BOLD MRI, sensitive to variations in tissue perfusion and blood oxygenation, illustrate the considerable inter-individual and intra-tumoral variability [243–246]. Therefore, the perfusion imaging can show functional differences in microvascularization according to the type of tumor. The crude angiogenesis of certain tumors is without vasoreactivity, while the angiogenesis of others is closer to normal cerebral vascularization and have vasoreactivity [247].

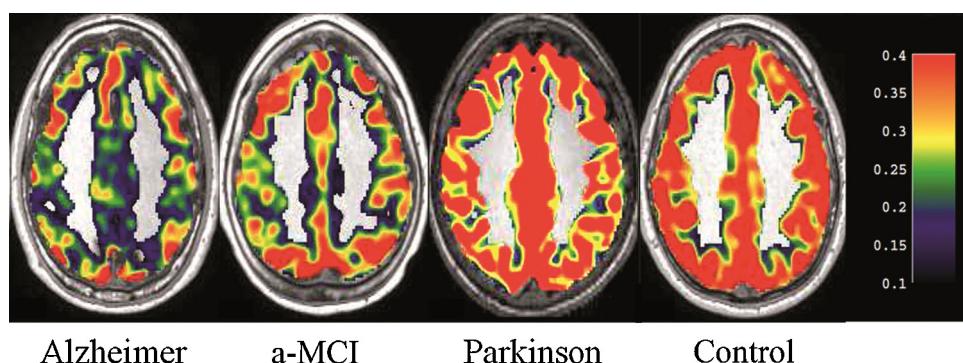


Figure 5. Vasoreactivity and neurodegenerative diseases. These illustrative cases of subjects about 70 years old, without microangiopathy, demonstrate that the study of cerebral vasoreactivity to CO₂ using BOLD fMRI detects diffuse abnormalities in a patient treated for beginning Alzheimer's disease and in a subject at risk with amnestic mild cognitive impairment (a-MCI) [186], while the vasoreactivity is normal in patients with Parkinson's disease when compared with a healthy subject [87].

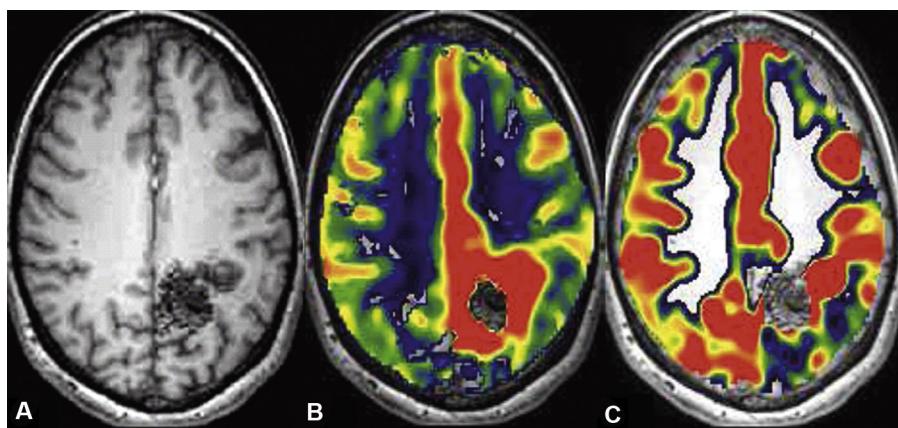


Figure 6. Alteration in the vasoreactivity in a patient with an arteriovenous malformation revealed by an epileptic seizure. A. Left parietal internal arteriovenous malformation (AVM). B. Local hyperperfusion that spares the parietal cortex. C. However, the BOLD-fMRI of the vasoreactivity to CO_2 reveals an alteration in the inferior perinidal parietal cortex (C). This presentation is described in patients with AMV revealed by epilepsy, while the perilesional vasoreactivity in patients without epilepsy was normal [203].

In traumatology, the monitoring of functional perfusion disorders is very important since their occurrence is a poor prognostic factor [38,39,41–45,183,184]. Several rare experimental trials have been carried out by perfusion CT-scan with Xenon 133 [43,248] or by positron emission tomography [249]. However, it is legitimate to wonder about the pertinence of having cerebral mapping when the expected anomalies are usually diffuse. This is why these barely ambulatory patients are usually explored in bed by transcranial Doppler during a modulation of the capnia. Nevertheless, several difficult cases may be solved by the regional anomalies detected by perfusion MRI or perfusion CT-scan during a short hypercapnia test.

Techniques to measure cerebral vasoreactivity

Choice of imaging method

Initially studied in transcranial echo Doppler, then in nuclear medicine (PET, SPECT), the imaging of the vasoreactivity appeared in MRI several years ago, with the development of rapid acquisition sequences. Perfusion tomodensitometry has also been proposed [250].

Compared with other imaging techniques, MRI is available and non-invasive while providing an image with an infra-centimetric spatial resolution and a temporal resolution of about one second, which make clinical use conceivable. In addition, cerebral MRI is currently used in neurology, neurosurgery and psychiatry, providing directly superimposable functional and morphological data during the same examination.

Several MRI methods have been proposed: BOLD contrast [109,161,225,228,251,252], ASL [170,195,215,216,253–255], susceptibility contrast by the injection of gadolinium [9,256,257], magnetic resonance angiography [217,258].

In addition to their availability, the multiplicity of methods attests to the limits of each of them. Excluding macrovascular imaging (Doppler, MRA), the calculation of

the CVR is based on the same principle with, for variations in the capnia:

$$\text{CVR} = \Delta \text{signal} (\%) / \Delta P_e \text{CO}_2 (\text{mmHg})^*$$

Theoretically, ASL is the best method since it allows the cerebral blood flow to be quantified and the variation in its signal may be expressed as a flow variation. Nevertheless, ASL requires careful methodological rigor with a correction of the T1 effects, the choice of post-saturation times and a 3T MRI to have a sufficient signal-noise ratio to detect a variation of several percent of this already very low signal [259]. Moreover, ASL with a fixed inversion time is sensitive to major variations in the arterial transit time of the labeled blood bolus, distorting the quality of the measurement in steno-occlusive disease. The current temporal resolution of multi-T1 ASL sequences does not allow for dynamic ASL. This being the case, the acquisitions with pseudo-continuous labeling cover the entire brain with a resolution under 10 s, allowing for the study of the dynamic variations in perfusion [67,216]. Now and in the absence of steno-occlusive disease, ASL seems to be the choice method to study healthy subjects in physiology [67] and the cognitive neurosciences, patients presenting a neurodegenerative disease or even children. However and in spite of these theoretical advantages, the use of ASL in this context remains very marginal compared with the increasing number of BOLD contrast studies.

Although, the BOLD signal is influenced by a great many parameters (Fig. 1), this reproducible method is correlated with variations in the cerebral blood flow [57,260]. We have already seen that variations in the signal were sensitive to the basal perfusion conditions. As a result, a measurement of basal perfusion either by ASL or susceptibility contrast is required to make sure that the differences in the BOLD signal variations are really due to modifications in the cerebral vasoreactivity and not simply the consequence of a modification in the basal perfusion [112,186,189].

Perfusion imaging by susceptibility contrast during the first pass of a gadolinium bolus may also be proposed. The main limit is the need to inject two boluses of contrast product, one before the vasomotor stimulus and the other after. In addition, the quantification of the results is still difficult, in particular in the presence of a rupture of the

brain-blood barrier. It also lacks the measurement of hemodynamic changes that might be relevant [112] because such imaging approach is performed at two different steady-states.

The perfusion CT-scan is also based on the study of the transit of a bolus of contrast product. The limits for this approach are the need to carry out two examinations, one before the vasomotor stimulus and the other after, thereby doubling the irradiation and the quantity of iodine injected [2,250]. The cover volume may also be limited, although it increases with the optimization of the detectors. The quantification of the perfusion, easier than with MRI, and its great availability may be determining elements if, in the future, the clinical applications of the imaging of the vasoreactivity becomes necessary.

Choice of vasomotor stimulus

In clinical practice, MRI of the vasoreactivity is above all obtained by modulating the capnia either by hyperventilation, apnea, inhalation of CO₂ or injection of acetazolamide. The study of the vasoreactivity with oxygen is much less developed since the vasomotor effects of oxygen are very low (see above) [64,68–71].

Acetazolamide (Diamox®) is a carbonic anhydrase inhibitor that converts CO₂ into bicarbonates. It provokes hypercapnia and acidosis responsible for a vasodilation and an increase in the CBF of about 20 to 30% [9,80,261]. In imaging, 15 mg/kg of the product is administered by intravenous route with a maximum of 1200 mg. The onset of action is after several minutes. Its efficacy lasts for several hours before a return to the initial state. This product has several contraindications (kidney failure, liver failure, allergy...) and many secondary effects (hydro-electrolytic disorders, diabetes, nephritic colic...). The pharmacokinetics and the adverse effects of acetazolamide limit its use in imaging and in particular in fMRI [80]. This stimulus requires two imaging examinations, one before the injection and the other at least 20 minutes after the injection. The intravenous administration of the product and its contraindications limits its use in healthy volunteers.

The sustained hyperventilation for at least 1 min induces hypocapnia with a P_eCO₂ under 30 mmHg, that is a reduction of 10 mmHg or more [8,50,74,82,109,164]. The hyperventilation is accompanied by a vasoconstriction responsible for a 25% reduction in the CBF [8,262] and a 1 to 5% drop in the BOLD signal [74,82,109,164]. Moreover, the hyperventilation is responsible for tachycardia without a change in the arterial pressure [50].

On the contrary, an apnea of at least 20 s is required to observe an increase in the CBF, which may attain 60% and a 1 to 5% increase in the BOLD signal [168,251,263–266]. The 20 s apnea allows for a 7 mmHg rise in the P_eCO₂ de 7 mmHg and a 45% increase in the arterial velocity, but it also provokes an increase in blood pressure, secondary hyperventilation and transient hypoxia. The return to equilibrium takes 45 s. However, the hypoxia does not seem to affect the increase in perfusion [267].

The hyperventilation and apnea are easy and reliable tasks. However, they induce movements of the head and are highly dependent on the sometimes-altered performances of the subjects and, a fortiori of the patients [109].

Moreover, except for sensory stimulations (visual for example), it's very difficult to carry out an additional cognitive task to study neurovascular coupling in the functional regions of motor skills or language.

The vasodilatation induced by the inhalation of 5 to 10% CO₂ for 2 to 3 min is accompanied by an increase in the cerebral blood flow (CBF) of 2 to 11%, 6% per mmHg, and the cerebral blood volume of 1.8% per mmHg [7]. Similar MRI results have been observed with ASL [254]. In BOLD fMRI, the increase in cerebral perfusion is accompanied by an increase in signal ranging from 0.2 to 0.3% per mmHg or 2–4% for an increase in expiratory pressure in CO₂ close to 10–15 mmHg [87,112,161,186,251,252,268]. The signal progressively varies to reach a plateau after 30–60 s. A return to the state of equilibrium also requires 30–60 s [58,76,162,166,167,251]. The amplitude of the variation in signal induced by the inhalation of CO₂ is roughly superior than that induced by the neurovascular coupling that is about 0.5 to 3%. This difference is not due to a sudden increase in neuronal activity for a moderate hypercapnia test [46,269].

Among the disadvantages of the inhalation of CO₂, several minor adverse and rapidly reversible effects should be noted. They have been reported during respiratory function tests and include mild headache, shortage of air, anxiety or fatigue [270–272]. The anxiogenic effects of the inhalation of CO₂ are experimentally used to provoke anxiety disorders, or even panic attacks, although the appearance of such effects requires much longer inhalation protocols (15–20 min) or the use of mixtures more concentrated in CO₂ (20–35% CO₂) [273,274]. The mean arterial pressure and the pulse are stable for several minutes of inhalation [50], whereas an increase in these parameters may be observed with an inhalation of over 10 min [275].

The inhalation of carbogen a 7/93% mixture of CO₂/O₂, is a potential alternative, since there are no reported contraindications or adverse effects. This aspect is readily used in a clinical context in brain-damaged subjects [50,111,182,246]. Carbogen is currently used in clinical practice, in cardiorespiratory fitness tests and to test the hyperventilation reflex to hypercapnia. In spite of the high oxygen content, the inhalation of carbogen with over 3% CO₂ provokes a vasodilation that in turn induces an increase in the arterial speed of about 1.5 cm.s⁻¹.mmHg P_eCO₂⁻¹ [50]. The increase in the BOLD signal is 2 to 6% in the grey matter [87,112,161,186,252]. With its high oxygen content, carbogen also induces tissue hyperoxia. This hyperoxia may provoke an increase in the BOLD signal of up to 3% in the grey matter [69,246], in spite of a potential reduction of several percent in the CBF [70]. This effect is accounted for by the intake of freely diffusible O₂ that is directly used by the neurons without having to extract the O₂ carried by the hemoglobin. This results in a reduction in the oxygen extraction factor and the deoxyhemoglobin concentration. This increase in oxygenation has to be taken into account now that the effects of carbogen are studied in BOLD imaging since, as with the interpretation of neurovascular coupling, it's difficult to distinguish the increase in perfusion from the local increase in oxygenation (Fig. 1). Moreover, the properties of carbogen have been used in neuro-oncology to increase the oxygenation of brain tumors and their radiosensitivity [245,246,276–278]. In spite of the encouraging experimental results [279], the evaluation of

the effects of radiosensitization by carbogen and nicotinamide have been disappointing [276,277]. However, we can debate the possibility of a better selection of patients, in particular on the basis of imaging of the tumor response to radiosensitizing agents [276].

Conclusion

To conclude, the functional imaging of cerebral perfusion is used for the mapping of the three main properties of brain microvascularization: neurovascular coupling, vasoreactivity to circulating gases and autoregulation of the perfusion pressure. BOLD fMRI of the cerebral vasoreactivity to CO₂ is increasingly used. fMRI of the cerebral vasoreactivity may be used to estimate the vascular reserve and risk of a stroke in steno-occlusive disease in order to improve the treatment. fMRI of cerebral vasoreactivity detects early alterations in Alzheimer's disease and the subjects at risk. fMRI of cerebral vasoreactivity may be used to better interpret the results in fMRI activation, in particular before neurosurgery.

Therefore, it's only a question of time, access and propagation of techniques. Most likely, the coming decade will confirm the development of the clinical applications of the functional imaging of perfusion for brain and heart diseases, as well as for other organs.

TAKE-HOME MESSAGES

- Functional imaging of cerebral perfusion is used in the mapping of the 3 main properties of brain microvascularization: neurovascular coupling, vasoreactivity to circulating gases and autoregulation of the perfusion pressure.
- BOLD fMRI of cerebral vasoreactivity to CO₂ is increasingly used.
- fMRI of cerebral vasoreactivity may be used to estimate the vascular reserve and the risk of stroke in steno-occlusive disease in order to improve the treatment.
- fMRI of cerebral vasoreactivity detects early alterations in Alzheimer's disease and subjects at risk.
- fMRI of cerebral vasoreactivity may be used to better interpret the results in fMRI activation, in particular before neurosurgery.

Disclosure of interest

The authors declare that they have no conflicts of interest concerning this article.

Acknowledgements

We thank doctors Florence Tahon, Kamel Boubagra, Olivier Detante, Katia Garambois and Marianne Barbeux for their comments. We also thank Patrice Jousse for the creation of the figures.

References

- [1] Buxton RB. Cerebral blood flow. In: Buxton RB, editor. Introduction to functional magnetic resonance imaging: principles and techniques. Cambridge: Cambridge university press; 2002. p. 22–40.
- [2] Wintermark M, Sesay M, Barbier E, Borbely K, Dillon WP, Eastwood JD, et al. Comparative overview of brain perfusion imaging techniques. *Stroke* 2005;36:e83–99.
- [3] Kety S, Schmidt C. Nitrous oxide method for quantitative determination of cerebral blood flow in man: theory, procedure and normal values. *J Clin Invest* 1948;27:475–83.
- [4] Grubb Jr RL, Raichle ME, Eichling JO, Ter-Pogossian MM. The effects of changes in PaCO₂ on cerebral blood volume, blood flow, and vascular mean transit time. *Stroke* 1974;5:630–9.
- [5] Lee SP, Duong TQ, Yang G, Iadecola C, Kim SG. Relative changes of cerebral arterial and venous blood volumes during increased cerebral blood flow: implications for BOLD fMRI. *Magn Reson Med* 2001;45:791–800.
- [6] Mandeville JB, Marota JJ, Ayata C, Zaharchuk G, Moskowitz MA, Rosen BR, et al. Evidence of a cerebrovascular postarterial windkessel with delayed compliance. *J Cereb Blood Flow Metab* 1999;19:679–89.
- [7] Ito H, Kanno I, Ibaraki M, Hatazawa J, Miura S. Changes in human cerebral blood flow and cerebral blood volume during hypercapnia and hypoxia measured by positron emission tomography. *J Cereb Blood Flow Metab* 2003;23:665–70.
- [8] Rostrup E, Knudsen GM, Law I, Holm S, Larsson HB, Paulson OB. The relationship between cerebral blood flow and volume in humans. *Neuroimage* 2005;24:1–11.
- [9] Grandin CB, Bol A, Smith AM, Michel C, Cosnard G. Absolute CBF and CBV measurements by MRI bolus tracking before and after acetazolamide challenge: Repeatability and comparison with PET in humans. *Neuroimage* 2005;26:525–35.
- [10] Cavaglia M, Dombrowski SM, Drazba J, Vasanji A, Bokesch PM, Janigro D. Regional variation in brain capillary density and vascular response to ischemia. *Brain Res* 2001;910:81–93.
- [11] Klein B, Kuschinsky W, Schrock H, Vetterlein F. Interdependency of local capillary density, blood flow, and metabolism in rat brains. *Am J Physiol* 1986;251:H1333–40.
- [12] Ito H, Kanno I, Shimosegawa E, Tamura H, Okane K, Hatazawa J. Hemodynamic changes during neural deactivation in human brain: a positron emission tomography study of crossed cerebellar diaschisis. *Ann Nucl Med* 2002;16:249–54.
- [13] Ito H, Takahashi K, Hatazawa J, Kim SG, Kanno I. Changes in human regional cerebral blood flow and cerebral blood volume during visual stimulation measured by positron emission tomography. *J Cereb Blood Flow Metab* 2001;21:608–12.
- [14] Ito H, Ibaraki M, Kanno I, Fukuda H, Miura S. Changes in cerebral blood flow and cerebral oxygen metabolism during neural activation measured by positron emission tomography: comparison with blood oxygenation level-dependent contrast measured by functional magnetic resonance imaging. *J Cereb Blood Flow Metab* 2005;25:371–7.
- [15] Kim T, Hendrich KS, Masamoto K, Kim SG. Arterial versus total blood volume changes during neural activity-induced cerebral blood flow change: implication for BOLD fMRI. *J Cereb Blood Flow Metab* 2007;27:1235–47.
- [16] Leenders KL, Perani D, Lammertsma AA, Heather JD, Buckingham P, Healy MJ, et al. Cerebral blood flow, blood volume and oxygen utilization. Normal values and effect of age. *Brain* 1990;113(Pt 1):27–47.
- [17] Kastrup A, Li TQ, Glover GH, Kruger G, Moseley ME. Gender differences in cerebral blood flow and oxygenation response during focal physiologic neural activity. *J Cereb Blood Flow Metab* 1999;19:1066–71.

- [18] Hurn P, Traystman R. Changes in arterial gaz tension. In: Edvinsson L, Krause D, editors. *Cerebral blood flow and metabolism*. Philadelphia: Lippincott Williams & Wilkins; 2002. p. 384–94.
- [19] Hamel E. Perivascular nerves and the regulation of cerebrovascular tone. *J Appl Physiol* 2006;100:1059–64.
- [20] Girouard H, Iadecola C. Neurovascular coupling in the normal brain and in hypertension, stroke, and Alzheimer disease. *J Appl Physiol* 2006;100:328–35.
- [21] Edvinsson L, Hamel E. Perivascular nerves in brain vessels. In: Edvinsson L, Krause D, editors. *Cerebral blood flow and metabolism*. Philadelphia: Lippincott Williams & Wilkins; 2002. p. 43–67.
- [22] Choi JK, Chen YI, Hamel E, Jenkins BG. Brain hemodynamic changes mediated by dopamine receptors: Role of the cerebral microvasculature in dopamine-mediated neurovascular coupling. *Neuroimage* 2006;30:700–12.
- [23] Krimer LS, Muly 3rd EC, Williams GV, Goldman-Rakic PS. Dopaminergic regulation of cerebral cortical microcirculation. *Nat Neurosci* 1998;1:286–9.
- [24] Takano T, Tian GF, Peng W, Lou N, Libionka W, Han X, et al. Astrocyte-mediated control of cerebral blood flow. *Nat Neurosci* 2006;9:260–7.
- [25] Zonta M, Angulo MC, Gobbo S, Rosengarten B, Hossmann KA, Pozzan T, et al. Neuron-to-astrocyte signaling is central to the dynamic control of brain microcirculation. *Nat Neurosci* 2003;6:43–50.
- [26] Lecrux C, Hamel E. The neurovascular unit in brain function and disease. *Acta Physiol (Oxf)* 2011;203:47–59.
- [27] Xu HL, Koenig HM, Ye S, Feinstein DL, Pelligrino DA. Influence of the glia limitans on pial arteriolar relaxation in the rat. *Am J Physiol Heart Circ Physiol* 2004;287:H331–9.
- [28] Attwell D, Iadecola C. The neural basis of functional brain imaging signals. *Trends Neurosci* 2002;25:621–5.
- [29] Drake CT, Iadecola C. The role of neuronal signaling in controlling cerebral blood flow. *Brain Lang* 2007;102:141–52.
- [30] Duong TQ, Kim DS, Ugurbil K, Kim SG. Spatiotemporal dynamics of the BOLD fMRI signals: toward mapping submillimeter cortical columns using the early negative response. *Magn Reson Med* 2000;44:231–42.
- [31] Fox PT, Raichle ME, Mintun MA, Dence C. Nonoxidative glucose consumption during focal physiologic neural activity. *Science* 1988;241:462–4.
- [32] Malonek D, Grinvald A. Interactions between electrical activity and cortical microcirculation revealed by imaging spectroscopy: implications for functional brain mapping. *Science* 1996;272:551–4.
- [33] Pellerin L, Magistretti PJ. Food for thought: challenging the dogmas. *J Cereb Blood Flow Metab* 2003;23:1282–6.
- [34] Farkas E, Luiten PG. Cerebral microvascular pathology in aging and Alzheimer's disease. *Prog Neurobiol* 2001;64:575–611.
- [35] de la Torre JC. Alzheimer disease as a vascular disorder: nosological evidence. *Stroke* 2002;33:1152–62.
- [36] Iadecola C. Rescuing troubled vessels in Alzheimer disease. *Nat Med* 2005;11:923–4.
- [37] Zlokovic BV. Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders. *Nat Rev Neurosci* 2011;12:723–38.
- [38] Panerai RB. Assessment of cerebral pressure autoregulation in humans—a review of measurement methods. *Physiol Meas* 1998;19:305–38.
- [39] Lang EW, Lagopoulos J, Griffith J, Yip K, Yam A, Mudaliar Y, et al. Cerebral vasomotor reactivity testing in head injury: the link between pressure and flow. *J Neurol Neurosurg Psychiatry* 2003;74:1053–9.
- [40] Chillon J-M, Baumbach G. Autoregulation: arterial and intracranial pressure. In: Edvinsson L, Krause D, editors. *Cerebral blood flow and metabolism*. Philadelphia: Lippincott Williams & Wilkins; 2002. p. 395–412.
- [41] Czosnyka M, Brady K, Reinhard M, Smielewski P, Steiner LA. Monitoring of cerebrovascular autoregulation: facts, myths, and missing links. *Neurocrit Care* 2009;10:373–86.
- [42] Jackson SA, Piper M, Dunn L, Leffler C, Daley M. Assessment of the variation in cerebrovascular reactivity in head injured patients. *Acta Neurochir Suppl* 2000;76:445–9.
- [43] Kelly DF, Kordestani RK, Martin NA, Nguyen T, Hovda DA, Bergsneider M, et al. Hyperemia following traumatic brain injury: relationship to intracranial hypertension and outcome. *J Neurosurg* 1996;85:762–71.
- [44] Lang EW, Lagopoulos J, Griffith J, Yip K, Mudaliar Y, Mehndorn HM, et al. Noninvasive cerebrovascular autoregulation assessment in traumatic brain injury: validation and utility. *J Neurotrauma* 2003;20:69–75.
- [45] Secher NH, van Lieshout JJ. Dynamic cerebral autoregulation and monitoring cerebral perfusion. *Hypertension* 2010;56:189–90.
- [46] Kety S, Schmidt C. The effects of the altered gas tensions of carbon dioxide and oxygen on cerebral blood flow and cerebral oxygen consumption of normal young men. *J Clin Invest* 1948;27:484–92.
- [47] Ito H, Yokoyama I, Iida H, Kinoshita T, Hatazawa J, Shimosegawa E, et al. Regional differences in cerebral vascular response to PaCO₂ changes in humans measured by positron emission tomography. *J Cereb Blood Flow Metab* 2000;20:1264–70.
- [48] Reivich M. Arterial PCO₂ and cerebral hemodynamics. *Am J Physiol* 1964;206:25–35.
- [49] Brugniaux JV, Hodges AN, Hanly PJ, Poulin MJ. Cerebrovascular responses to altitude. *Respir Physiol Neurobiol* 2007;158:212–23.
- [50] Lavi S, Gaitini D, Milloul V, Jacob G. Impaired cerebral CO₂ vasoactivity: association with endothelial dysfunction. *Am J Physiol Heart Circ Physiol* 2006;291:H1856–61.
- [51] Morii S, Ngai AC, Winn HR. Reactivity of rat pial arterioles and venules to adenosine and carbon dioxide: with detailed description of the closed cranial window technique in rats. *J Cereb Blood Flow Metab* 1986;6:34–41.
- [52] Wei EP, Kontos HA, Patterson Jr JL. Dependence of pial arteriolar response to hypercapnia on vessel size. *Am J Physiol* 1980;238:697–703.
- [53] Atkinson JL, Anderson RE, Sundt Jr TM. The effect of carbon dioxide on the diameter of brain capillaries. *Brain Res* 1990;517:333–40.
- [54] Kontos HA, Raper AJ, Patterson JL. Analysis of vasoactivity of local pH, PCO₂ and bicarbonate on pial vessels. *Stroke* 1977;8:358–60.
- [55] Dulla CG, Dobelis P, Pearson T, Frenguelli BG, Staley KJ, Masino SA. Adenosine and ATP link PCO₂ to cortical excitability via pH. *Neuron* 2005;48:1011–23.
- [56] Sandor P, Komjati K, Reivich M, Nyary I. Major role of nitric oxide in the mediation of regional CO₂ responsiveness of the cerebral and spinal cord vessels of the cat. *J Cereb Blood Flow Metab* 1994;14:49–58.
- [57] Kassner A, Winter JD, Poublanc J, Mikulis DJ, Crawley AP. Blood-oxygen level dependent MRI measures of cerebrovascular reactivity using a controlled respiratory challenge: reproducibility and gender differences. *J Magn Reson Imaging* 2010;31:298–304.
- [58] Riecker A, Grodd W, Klose U, Schulz JB, Groschel K, Erb M, et al. Relation between regional functional MRI activation and vascular reactivity to carbon dioxide during normal aging. *J Cereb Blood Flow Metab* 2003;23:565–73.
- [59] Ainslie PN, Murrell C, Peebles K, Swart M, Skinner MA, Williams MJ, et al. Early morning impairment in cerebral

- autoregulation and cerebrovascular CO₂ reactivity in healthy humans: relation to endothelial function. *Exp Physiol* 2007;92:769–77.
- [60] Cummings KJ, Swart M, Ainslie PN. Morning attenuation in cerebrovascular CO₂ reactivity in healthy humans is associated with a lowered cerebral oxygenation and an augmented ventilatory response to CO₂. *J Appl Physiol* 2007;102:1891–8.
- [61] Bes A, Geraud G. Circulation cérébrale: physiologie. Reuil-Malmaison: Sandoz éditions; 1974.
- [62] Prisman E, Slessarev M, Han J, Poublanc J, Mardimae A, Crawley A, et al. Comparison of the effects of independently-controlled end-tidal PCO₂) and PO₂) on blood oxygen level-dependent (BOLD) MRI. *J Magn Reson Imaging* 2008;27:185–91.
- [63] Golovan EV, Reis DJ. Contribution of oxygen-sensitive neurons of the rostral ventrolateral medulla to hypoxic cerebral vasodilatation in the rat. *J Physiol* 1996;495(Pt 1):201–16.
- [64] Johnston AJ, Steiner LA, Gupta AK, Menon DK. Cerebral oxygen vasoreactivity and cerebral tissue oxygen reactivity. *Br J Anaesth* 2003;90:774–86.
- [65] Xu K, Lamanna JC. Chronic hypoxia and the cerebral circulation. *J Appl Physiol* 2006;100:725–30.
- [66] Patt S, Sampaolo S, Theallier-Janko A, Tschairkin I, Cervos-Navarro J. Cerebral angiogenesis triggered by severe chronic hypoxia displays regional differences. *J Cereb Blood Flow Metab* 1997;17:801–6.
- [67] Villien M, Bouzat P, Rupp T, Robach P, Lamalle L, Tropres I, et al. Changes in cerebral blood flow and vasoreactivity to CO₂ measured by arterial spin labeling after 6 days at 4350 m. *Neuroimage* 2013;72:272–9.
- [68] Watson NA, Beards SC, Altaf N, Kassner A, Jackson A. The effect of hyperoxia on cerebral blood flow: a study in healthy volunteers using magnetic resonance phase-contrast angiography. *Eur J Anaesthesiol* 2000;17:152–9.
- [69] Chiarelli PA, Bulte DP, Wise R, Gallichan D, Jezzard P. A calibration method for quantitative BOLD fMRI based on hyperoxia. *Neuroimage* 2007;37:808–20.
- [70] Bulte DP, Chiarelli PA, Wise RG, Jezzard P. Cerebral perfusion response to hyperoxia. *J Cereb Blood Flow Metab* 2007;27:69–75.
- [71] Rostrup E, Larsson HB, Toft PB, Garde K, Henriksen O. Signal changes in gradient echo images of human brain induced by hypo- and hyperoxia. *NMR Biomed* 1995;8:41–7.
- [72] Sicard KM, Duong TQ. Effects of hypoxia, hyperoxia, and hypercapnia on baseline and stimulus-evoked BOLD, CBF, and CMRO₂ in spontaneously breathing animals. *Neuroimage* 2005;25:850–8.
- [73] Demchenko IT, Oury TD, Crapo JD, Piantadosi CA. Regulation of the brain's vascular responses to oxygen. *Circ Res* 2002;91:1031–7.
- [74] Cohen ER, Ugurbil K, Kim SG. Effect of basal conditions on the magnitude and dynamics of the blood oxygenation level-dependent fMRI response. *J Cereb Blood Flow Metab* 2002;22:1042–53.
- [75] Restom K, Bangen KJ, Bondi MW, Perthen JE, Liu TT. Cerebral blood flow and BOLD responses to a memory encoding task: a comparison between healthy young and elderly adults. *Neuroimage* 2007;37:430–9.
- [76] Bandettini PA, Wong EC. A hypercapnia-based normalization method for improved spatial localization of human brain activation with fMRI. *NMR Biomed* 1997;10:197–203.
- [77] Kemna LJ, Posse S, Tellmann L, Schmitz T, Herzog H. Interdependence of regional and global cerebral blood flow during visual stimulation: an O-15-butanol positron emission tomography study. *J Cereb Blood Flow Metab* 2001;21:664–70.
- [78] Kemna LJ, Posse S. Effect of respiratory CO₂ changes on the temporal dynamics of the hemodynamic response in functional MR imaging. *Neuroimage* 2001;14:642–9.
- [79] Posse S, Kemna LJ, Elghahwagi B, Wiese S, Kiselev VG. Effect of graded hypo- and hypercapnia on fMRI contrast in visual cortex: quantification of T(*)₂ changes by multiecho EPI. *Magn Reson Med* 2001;46:264–71.
- [80] Brown GG, Elyer Zorrilla LT, Georgy B, Kindermann SS, Wong EC, Buxton RB. BOLD and perfusion response to finger-thumb apposition after acetazolamide administration: differential relationship to global perfusion. *J Cereb Blood Flow Metab* 2003;23:829–37.
- [81] Hoge RD, Atkinson J, Gill B, Crelier GR, Marrett S, Pike GB. Investigation of BOLD signal dependence on cerebral blood flow and oxygen consumption: the deoxyhemoglobin dilution model. *Magn Reson Med* 1999;42:849–63.
- [82] Weckesser M, Posse S, Olthoff U, Kemna L, Dager S, Muller-Gartner HW. Functional imaging of the visual cortex with bold-contrast MRI: hyperventilation decreases signal response. *Magn Reson Med* 1999;41:213–6.
- [83] Ramsay SC, Murphy K, Shea SA, Friston KJ, Lammertsma AA, Clark JC, et al. Changes in global cerebral blood flow in humans: effect on regional cerebral blood flow during a neural activation task. *J Physiol* 1993;471:521–34.
- [84] Corfield DR, Murphy K, Josephs O, Adams L, Turner R. Does hypercapnia-induced cerebral vasodilation modulate the hemodynamic response to neural activation? *Neuroimage* 2001;13:1207–11.
- [85] Stefanovic B, Warnking JM, Rylander KM, Pike GB. The effect of global cerebral vasodilation on focal activation hemodynamics. *Neuroimage* 2006;30:726–34.
- [86] Uludag K, Dubowitz DJ, Yoder EJ, Restom K, Liu TT, Buxton RB. Coupling of cerebral blood flow and oxygen consumption during physiological activation and deactivation measured with fMRI. *Neuroimage* 2004;23:148–55.
- [87] Krainik A, Maillet A, Fleury V, Sahin M, Tropres I, Lamalle L, et al. Levodopa does not change cerebral vasoreactivity in Parkinson's disease. *Mov Disord* 2013;28:469–75.
- [88] Logothetis NK, Wandell BA. Interpreting the BOLD signal. *Annu Rev Physiol* 2004;66:735–69.
- [89] Kim SG, Ogawa S. Biophysical and physiological origins of blood oxygenation level-dependent fMRI signals. *J Cereb Blood Flow Metab* 2012;32:1188–206.
- [90] Belle V, Delon-Martin C, Massarelli R, Decety J, Le Bas JF, Benabid AL, et al. Intracranial gradient-echo and spin-echo functional MR angiography in humans. *Radiology* 1995;195:739–46.
- [91] Buxton RB, Wong EC, Frank LR. Dynamics of blood flow and oxygenation changes during brain activation: the balloon model. *Magn Reson Med* 1998;39:855–64.
- [92] Ogawa S, Tank DW, Menon R, Ellermann JM, Kim SG, Merkle H, et al. Intrinsic signal changes accompanying sensory stimulation: functional brain mapping with magnetic resonance imaging. *Proc Natl Acad Sci U S A* 1992;89:5951–5.
- [93] Yamane F, Muragaki Y, Maruyama T, Okada Y, Iseki H, Ikeda A, et al. Preoperative mapping for patients with supplementary motor area epilepsy: multimodality brain mapping. *Psychiatry Clin Neurosci* 2004;58:S16–21.
- [94] Stippich C, Freitag P, Kassubek J, Soros P, Kamada K, Kober H, et al. Motor, somatosensory and auditory cortex localization by fMRI and MEG. *Neuroreport* 1998;9:1953–7.
- [95] Krings T, Schreckenberger M, Rohde V, Folts H, Spetzger U, Sabri O, et al. Metabolic and electrophysiological validation of functional MRI. *J Neurol Neurosurg Psychiatry* 2001;71:762–71.
- [96] Lehericy S, Duffau H, Cornu P, Capelle L, Pidoux B, Carpentier A, et al. Correspondence between functional magnetic resonance imaging somatotopy and individual brain anatomy of the central region: comparison with intraoperative stimulation in patients with brain tumors. *J Neurosurg* 2000;92:589–98.

- [97] Hanakawa T, Ikeda A, Sadato N, Okada T, Fukuyama H, Nagamine T, et al. Functional mapping of human medial frontal motor areas. The combined use of functional magnetic resonance imaging and cortical stimulation. *Exp Brain Res* 2001;138:403–9.
- [98] Krainik A, Lehericy S, Duffau H, Vlaicu M, Poupon F, Capelle L, et al. Role of the supplementary motor area in motor deficit following medial frontal lobe surgery. *Neurology* 2001;57:871–8.
- [99] Krainik A, Lehericy S, Duffau H, Capelle L, Chainay H, Cornu P, et al. Postoperative speech disorder after medial frontal surgery: role of the supplementary motor area. *Neurology* 2003;60:587–94.
- [100] Krainik A, Duffau H, Capelle L, Cornu P, Boch AL, Mangin JF, et al. Role of the healthy hemisphere in recovery after resection of the supplementary motor area. *Neurology* 2004;62:1323–32.
- [101] D'Esposito M, Deouell LY, Gazzaley A. Alterations in the BOLD fMRI signal with ageing and disease: a challenge for neuroimaging. *Nat Rev Neurosci* 2003;4:863–72.
- [102] Arthurs OJ, Boniface S. How well do we understand the neural origins of the fMRI BOLD signal? *Trends Neurosci* 2002;25:27–31.
- [103] Hoge RD, Atkinson J, Gill B, Crelier GR, Marrett S, Pike GB. Linear coupling between cerebral blood flow and oxygen consumption in activated human cortex. *Proc Natl Acad Sci U S A* 1999;96:9403–8.
- [104] Logothetis NK, Pauls J, Augath M, Trinath T, Oeltermann A. Neurophysiological investigation of the basis of the fMRI signal. *Nature* 2001;412:150–7.
- [105] Kannurpatti SS, Biswal BB, Hudetz AG. Baseline physiological state and the fMRI-BOLD signal response to apnea in anesthetized rats. *NMR Biomed* 2003;16:261–8.
- [106] Sicard K, Shen Q, Brevard ME, Sullivan R, Ferris CF, King JA, et al. Regional cerebral blood flow and BOLD responses in conscious and anesthetized rats under basal and hypercapnic conditions: implications for functional MRI studies. *J Cereb Blood Flow Metab* 2003;23:472–81.
- [107] Tuunanen PI, Kauppinen RA. Effects of oxygen saturation on BOLD and arterial spin labelling perfusion fMRI signals studied in a motor activation task. *Neuroimage* 2006;30:102–9.
- [108] Wise RG, Ide K, Poulin MJ, Tracey I. Resting fluctuations in arterial carbon dioxide induce significant low frequency variations in BOLD signal. *Neuroimage* 2004;21:1652–64.
- [109] Krainik A, Hund-Georgiadis M, Zysset S, von Cramon DY. Regional impairment of cerebrovascular reactivity and BOLD signal in adults after stroke. *Stroke* 2005;36:1146–52.
- [110] Rossini PM, Altamura C, Ferretti A, Vernieri F, Zappasodi F, Caulo M, et al. Does cerebrovascular disease affect the coupling between neuronal activity and local haemodynamics? *Brain* 2004;127:99–110.
- [111] Hamzei F, Knab R, Weiller C, Rother J. The influence of extra- and intracranial artery disease on the BOLD signal in fMRI. *Neuroimage* 2003;20:1393–9.
- [112] Jiang Z, Krainik A, David O, Salon C, Tropres I, Hoffmann D, et al. Impaired fMRI activation in patients with primary brain tumors. *Neuroimage* 2010;52:538–48.
- [113] Hesselmann V, Zaro Weber O, Wedekind C, Krings T, Schulte O, Kugel H, et al. Age related signal decrease in functional magnetic resonance imaging during motor stimulation in humans. *Neurosci Lett* 2001;308:141–4.
- [114] Mehagnoul-Schipper DJ, van der Kallen BF, Colier WN, van der Sluijs MC, van Erning LJ, Thijssen HO, et al. Simultaneous measurements of cerebral oxygenation changes during brain activation by near-infrared spectroscopy and functional magnetic resonance imaging in healthy young and elderly subjects. *Hum Brain Mapp* 2002;16:14–23.
- [115] D'Esposito M, Zarahn E, Aguirre GK, Rypma B. The effect of normal aging on the coupling of neural activity to the bold hemodynamic response. *Neuroimage* 1999;10:6–14.
- [116] Mulderink TA, Gitelman DR, Mesulam MM, Parrish TB. On the use of caffeine as a contrast booster for BOLD fMRI studies. *Neuroimage* 2002;15:37–44.
- [117] Laurienti PJ, Field AS, Burdette JH, Maldjian JA, Yen YF, Moody DM. Dietary caffeine consumption modulates fMRI measures. *Neuroimage* 2002;17:751–7.
- [118] Liu TT, Behzadi Y, Restom K, Uludag K, Lu K, Buracas GT, et al. Caffeine alters the temporal dynamics of the visual BOLD response. *Neuroimage* 2004;23:1402–13.
- [119] Seifritz E, Bilecen D, Hanggi D, Haselhorst R, Radu EW, Wetzel S, et al. Effect of ethanol on BOLD response to acoustic stimulation: implications for neuropharmacological fMRI. *Psychiatry Res* 2000;99:1–13.
- [120] Braus DF, Ende G, Weber-Fahr W, Sartorius A, Krier A, Hubrich-Ungureanu P, et al. Antipsychotic drug effects on motor activation measured by functional magnetic resonance imaging in schizophrenic patients. *Schizophr Res* 1999;39:19–29.
- [121] Brassen S, Tost H, Hoehn F, Weber-Fahr W, Klein S, Braus DF. Haloperidol challenge in healthy male humans: a functional magnetic resonance imaging study. *Neurosci Lett* 2003;340:193–6.
- [122] Peters S, Suchan B, Rusin J, Daum I, Koster O, Przuntek H, et al. Apomorphine reduces BOLD signal in fMRI during voluntary movement in Parkinsonian patients. *Neuroreport* 2003;14:809–12.
- [123] Marcar VL, Schwarz U, Martin E, Loenneker T. How depth of anesthesia influences the blood oxygenation level-dependent signal from the visual cortex of children. *AJNR Am J Neuroradiol* 2006;27:799–805.
- [124] Born AP, Law I, Lund TE, Rostrup E, Hanson LG, Wildschiodtz G, et al. Cortical deactivation induced by visual stimulation in human slow-wave sleep. *Neuroimage* 2002;17:1325–35.
- [125] Takahashi H, Yahata N, Koeda M, Takano A, Asai K, Suhara T, et al. Effects of dopaminergic and serotonergic manipulation on emotional processing: a pharmacological fMRI study. *Neuroimage* 2005;27:991–1001.
- [126] Qiao M, Rushforth D, Wang R, Shaw RA, Tomanek B, Dunn JF, et al. Blood-oxygen-level-dependent magnetic resonance signal and cerebral oxygenation responses to brain activation are enhanced by concurrent transient hypertension in rats. *J Cereb Blood Flow Metab* 2007;27:1280–9.
- [127] Wang R, Foniok T, Wamsteeker JL, Qiao M, Tomanek B, Vivanco RA, et al. Transient blood pressure changes affect the functional magnetic resonance imaging detection of cerebral activation. *Neuroimage* 2006;31:1–11.
- [128] Hund-Georgiadis M, Mildner T, Georgiadis D, Weih K, von Cramon DY. Impaired hemodynamics and neural activation? A fMRI study of major cerebral artery stenosis. *Neurology* 2003;61:1276–9.
- [129] Rother J, Knab R, Hamzei F, Fiehler J, Reichenbach JR, Buchel C, et al. Negative dip in BOLD fMRI is caused by blood flow-oxygen consumption uncoupling in humans. *Neuroimage* 2002;15:98–102.
- [130] Roc AC, Wang J, Ances BM, Liebeskind DS, Kasner SE, Detre JA. Altered hemodynamics and regional cerebral blood flow in patients with hemodynamically significant stenoses. *Stroke* 2006;37:382–7.
- [131] Rocca MA, Colombo B, Pagani E, Falini A, Codella M, Scotti G, et al. Evidence for cortical functional changes in patients with migraine and white matter abnormalities on conventional and diffusion tensor magnetic resonance imaging. *Stroke* 2003;34:665–70.
- [132] Faro SH, Mohamed FB, Tracy JL, Elfant RM, Pinus AB, Lublin FD, et al. Quantitative functional MR imaging of the visual cortex at 1.5 T as a function of luminance contrast in healthy

- volunteers and patients with multiple sclerosis. *AJNR Am J Neuroradiol* 2002;23:59–65.
- [133] Rombouts SA, Goekoop R, Stam CJ, Barkhof F, Scheltens P. Delayed rather than decreased BOLD response as a marker for early Alzheimer's disease. *Neuroimage* 2005;26:1078–85.
- [134] Pineiro R, Pendlebury S, Johansen-Berg H, Matthews PM. Altered hemodynamic responses in patients after subcortical stroke measured by functional MRI. *Stroke* 2002;33:103–9.
- [135] Holodny AI, Schulder M, Liu WC, Wolko J, Maldjian JA, Kalnin AJ. The effect of brain tumors on BOLD functional MR imaging activation in the adjacent motor cortex: implications for image-guided neurosurgery. *AJNR Am J Neuroradiol* 2000;21:1415–22.
- [136] Schreiber A, Hubbe U, Ziyeh S, Hennig J. The influence of gliomas and nonglial space-occupying lesions on blood-oxygen-level-dependent contrast enhancement. *AJNR Am J Neuroradiol* 2000;21:1055–63.
- [137] Ulmer JL, Hacein-Bey L, Mathews VP, Mueller WM, DeYoe EA, Prost RW, et al. Lesion-induced pseudo-dominance at functional magnetic resonance imaging: implications for preoperative assessments. *Neurosurgery* 2004;55:569–79 [discussion 80–1].
- [138] Fujiwara N, Sakatani K, Katayama Y, Murata Y, Hoshino T, Fukaya C, et al. Evoked-cerebral blood oxygenation changes in false-negative activations in BOLD contrast functional MRI of patients with brain tumors. *Neuroimage* 2004;21:1464–71.
- [139] Krainik A. Functional MRI. In: Duffau H, editor. *Brain mapping: from neural basis of cognition to surgical applications*. Wien: Springer-Verlag; 2011. p. 45–59.
- [140] Zaca D, Hua J, Pillai JJ. Cerebrovascular reactivity mapping for brain tumor presurgical planning. *World J Clin Oncol* 2011;2:289–98.
- [141] Schaller C, Schramm J, Haun D, Meyer B. Patterns of cortical oxygen saturation changes during CO₂ reactivity testing in the vicinity of cerebral arteriovenous malformations. *Stroke* 2003;34:938–44.
- [142] Lehericy S, Biondi A, Sourour N, Vlaicu M, du Montcel ST, Cohen L, et al. Arteriovenous brain malformations: is functional MR imaging reliable for studying language reorganization in patients? Initial observations. *Radiology* 2002;223:672–82.
- [143] Liu TT, Brown GG. Measurement of cerebral perfusion with arterial spin labeling: part 1. Methods. *J Int Neuropsychol Soc* 2007;13:517–25.
- [144] Raoult H, Petr J, Bannier E, Stamm A, Gauvrit JY, Barillot C, et al. Arterial spin labeling for motor activation mapping at 3T with a 32-channel coil: reproducibility and spatial accuracy in comparison with BOLD fMRI. *Neuroimage* 2011;58:157–67.
- [145] Pimentel MA, Vilela P, Sousa I, Figueiredo P. Localization of the hand motor area by arterial spin labeling and blood oxygen level-dependent functional magnetic resonance imaging. *Hum Brain Mapp* 2013;34(1):96–108.
- [146] Davis TL, Kwong KK, Weisskoff RM, Rosen BR. Calibrated functional MRI: mapping the dynamics of oxidative metabolism. *Proc Natl Acad Sci U S A* 1998;95:1834–9.
- [147] Blockley NP, Griffith VE, Simon AB, Buxton RB. A review of calibrated blood oxygenation level-dependent (BOLD) methods for the measurement of task-induced changes in brain oxygen metabolism. *NMR Biomed* 2013;26(8):987–1003.
- [148] Grouiller F, Vercueil L, Krainik A, Segebarth C, Kahane P, David O. Characterization of the hemodynamic modes associated with interictal epileptic activity using a deformable model-based analysis of combined EEG and functional MRI recordings. *Hum Brain Mapp* 2010;31:1157–73.
- [149] Saqqur M, Zygun D, Demchuk A. Role of transcranial Doppler in neurocritical care. *Crit Care Med* 2007;35:S216–23.
- [150] Vokatch N, Grotzsch H, Mermilliod B, Burkhard PR, Sztajzel R. Is cerebral autoregulation impaired in Parkinson's disease? A transcranial Doppler study. *J Neurol Sci* 2007;254:49–53.
- [151] Kastrup A, Thomas C, Hartmann C, Schabet M. Sex dependency of cerebrovascular CO₂ reactivity in normal subjects. *Stroke* 1997;28:2353–6.
- [152] Rogers RL, Meyer JS, Mortel KF, Mahurin RK, Thornby J. Age-related reductions in cerebral vasomotor reactivity and the law of initial value: a 4-year prospective longitudinal study. *J Cereb Blood Flow Metab* 1985;5:79–85.
- [153] Kannurpatti SS, Biswal BB. Effect of anesthesia on CBF, MAP and fMRI-BOLD signal in response to apnea. *Brain Res* 2004;1011:141–7.
- [154] Sakashita Y, Kanai M, Sugimoto T, Taki S, Takamori M. Changes in cerebral blood flow and vasoreactivity in response to acetazolamide in patients with transient global amnesia. *J Neurol Neurosurg Psychiatry* 1997;63:605–10.
- [155] van Osch MJ, Rutgers DR, Vonken EP, van Huffelen AC, Klijn CJ, Bakker CJ, et al. Quantitative cerebral perfusion MRI and CO₂ reactivity measurements in patients with symptomatic internal carotid artery occlusion. *Neuroimage* 2002;17:469–78.
- [156] Brevard ME, Duong TQ, King JA, Ferris CF. Changes in MRI signal intensity during hypercapnic challenge under conscious and anesthetized conditions. *Magn Reson Imaging* 2003;21:995–1001.
- [157] Dauphin F, Lacombe P, Sercombe R, Hamel E, Seylaz J. Hypercapnia and stimulation of the substantia innominata increase rat frontal cortical blood flow by different cholinergic mechanisms. *Brain Res* 1991;553:75–83.
- [158] Lacombe P, Sercombe R, Vaucher E, Seylaz J. Reduced cortical vasodilatory response to stimulation of the nucleus basalis of Meynert in the aged rat and evidence for a control of the cerebral circulation. *Ann N Y Acad Sci* 1997;826:410–5.
- [159] Harik SI, Prado R, Bustó R, Ginsberg MD. Increased cerebral blood flow during hypercapnia is not affected by lesion of the nucleus locus ceruleus. *Stroke* 1986;17:1235–8.
- [160] Kastrup A, Engelhorn T, Beaulieu C, de Crespigny A, Moseley ME. Dynamics of cerebral injury, perfusion, and blood-brain barrier changes after temporary and permanent middle cerebral artery occlusion in the rat. *J Neurol Sci* 1999;166:91–9.
- [161] Vesely A, Sasano H, Volgyesi G, Somogyi R, Tesler J, Fedorko L, et al. MRI mapping of cerebrovascular reactivity using square wave changes in end-tidal PCO₂. *Magn Reson Med* 2001;45:1011–3.
- [162] Macey PM, Macey KE, Kumar R, Harper RM. A method for removal of global effects from fMRI time series. *Neuroimage* 2004;22:360–6.
- [163] Posse S, Olthoff U, Weckesser M, Jancke L, Muller-Gartner HW, Dager SR. Regional dynamic signal changes during controlled hyperventilation assessed with blood oxygen level-dependent functional MR imaging. *AJNR Am J Neuroradiol* 1997;18:1763–70.
- [164] Naganawa S, Norris DG, Zysset S, Mildner T. Regional differences of fMR signal changes induced by hyperventilation: comparison between SE-EPI and GE-EPI at 3-T. *J Magn Reson Imaging* 2002;15:23–30.
- [165] Nishimura S, Suzuki A, Hatazawa J, Nishimura H, Shirane R, Yasui N, et al. Cerebral blood-flow responses to induced hypotension and to CO₂ inhalation in patients with major cerebral artery occlusive disease: a positron-emission tomography study. *Neuroradiology* 1999;41:73–9.
- [166] Rostrup E, Law I, Blenkinsberg M, Larsson HB, Born AP, Holm S, et al. Regional differences in the CBF and BOLD responses to hypercapnia: a combined PET and fMRI study. *Neuroimage* 2000;11:87–97.
- [167] Kim SG, Rostrup E, Larsson HB, Ogawa S, Paulson OB. Determination of relative CMRO₂ from CBF and BOLD changes:

- significant increase of oxygen consumption rate during visual stimulation. *Magn Reson Med* 1999;41:1152–61.
- [168] Liu HL, Huang JC, Wu CT, Hsu YY. Detectability of blood oxygenation level-dependent signal changes during short breath hold duration. *Magn Reson Imaging* 2002;20:643–8.
- [169] Blockley NP, Driver ID, Francis ST, Fisher JA, Gowland PA. An improved method for acquiring cerebrovascular reactivity maps. *Magn Reson Med* 2011;65:1278–86.
- [170] Hajjar I, Zhao P, Alsop D, Novak V. Hypertension and cerebral vasoreactivity: a continuous arterial spin labeling magnetic resonance imaging study. *Hypertension* 2010;56:859–64.
- [171] Dandona P, James IM, Newbury PA, Woollard ML, Beckett AG. Cerebral blood flow in diabetes mellitus: evidence of abnormal cerebrovascular reactivity. *Br Med J* 1978;2:325–6.
- [172] Kadoi Y, Hinohara H, Kunimoto F, Saito S, Ide M, Hiraoka H, et al. Diabetic patients have an impaired cerebral vasodilatory response to hypercapnia under propofol anesthesia. *Stroke* 2003;34:2399–403.
- [173] Novak V, Zhao P, Manor B, Sejdic E, Alsop D, Abduljalil A, et al. Adhesion molecules, altered vasoreactivity, and brain atrophy in type 2 diabetes. *Diabetes care* 2011;34:2438–41.
- [174] Terborg C, Schummer W, Albrecht M, Reinhart K, Weiller C, Rother J. Dysfunction of vasomotor reactivity in severe sepsis and septic shock. *Intensive Care Med* 2001;27:1231–4.
- [175] Walters M, Muir S, Shah I, Lees K. Effect of perindopril on cerebral vasomotor reactivity in patients with lacunar infarction. *Stroke* 2004;35:1899–902.
- [176] Pattinson KT, Rogers R, Mayhew SD, Tracey I, Wise RG. Pharmacological fMRI: measuring opioid effects on the BOLD response to hypercapnia. *J Cereb Blood Flow Metab* 2007;27:414–23.
- [177] Marstrand JR, Garde E, Rostrup E, Ring P, Rosenbaum S, Mortensen EL, et al. Cerebral perfusion and cerebrovascular reactivity are reduced in white matter hyperintensities. *Stroke* 2002;33:972–6.
- [178] Terborg C, Gora F, Weiller C, Rother J. Reduced vasomotor reactivity in cerebral microangiopathy: a study with near-infrared spectroscopy and transcranial Doppler sonography. *Stroke* 2000;31:924–9.
- [179] Bakker SL, de Leeuw FE, de Groot JC, Hofman A, Koudstaal PJ, Breteler MM. Cerebral vasomotor reactivity and cerebral white matter lesions in the elderly. *Neurology* 1999;52:578–83.
- [180] Lacombe P, Oligo C, Domenga V, Tournier-Lasserve E, Joutel A. Impaired cerebral vasoreactivity in a transgenic mouse model of cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy arteriopathy. *Stroke* 2005;36:1053–8.
- [181] Pfefferkorn T, von Stuckrad-Barre S, Herzog J, Gasser T, Hamann GF, Dichgans M. Reduced cerebrovascular CO₂ reactivity in CADASIL: a transcranial Doppler sonography study. *Stroke* 2001;32:17–21.
- [182] Lee EJ, Hung YC, Chang CH, Pai MC, Chen HH. Cerebral blood flow velocity and vasomotor reactivity before and after shunting surgery in patients with normal pressure hydrocephalus. *Acta Neurochir (Wien)* 1998;140:599–604 [discussion -5].
- [183] Martin NA, Patwardhan RV, Alexander MJ, Africk CZ, Lee JH, Shalmon E, et al. Characterization of cerebral hemodynamic phases following severe head trauma: hypoperfusion, hyperemia, and vasospasm. *J Neurosurg* 1997;87:9–19.
- [184] Carmona Suazo JA, Maas AI, van den Brink WA, van Santbrink H, Steyerberg EW, Avezaat CJ. CO₂ reactivity and brain oxygen pressure monitoring in severe head injury. *Crit Care Med* 2000;28:3268–74.
- [185] Oishi M, Mochizuki Y, Takasu T. Regional differences in cerebrovascular reactivity to acetazolamide in Alzheimer's disease. *J Clin Neurosci* 1999;6:380–1.
- [186] Cantin S, Villien M, Moreaud O, Tropres I, Keignart S, Chipon E, et al. Impaired cerebral vasoreactivity to CO₂ in Alzheimer's disease using BOLD fMRI. *Neuroimage* 2011;58:579–87.
- [187] Silvestrini M, Pasqualetti P, Baruffaldi R, Bartolini M, Handouk Y, Matteis M, et al. Cerebrovascular reactivity and cognitive decline in patients with Alzheimer disease. *Stroke* 2006;37:1010–5.
- [188] Glodzik L, Rusinek H, Brys M, Tsui WH, Switalski R, Mosconi L, et al. Framingham cardiovascular risk profile correlates with impaired hippocampal and cortical vasoreactivity to hypercapnia. *J Cereb Blood Flow Metab* 2011;31:671–9.
- [189] Yezhuvath US, Uh J, Cheng Y, Martin-Cook K, Weiner M, Diaz-Arrastia R, et al. Forebrain-dominant deficit in cerebrovascular reactivity in Alzheimer's disease. *Neurobiol Aging* 2012;33:75–82.
- [190] Vernieri F, Pasqualetti P, Matteis M, Passarelli F, Troisi E, Rossini PM, et al. Effect of collateral blood flow and cerebral vasomotor reactivity on the outcome of carotid artery occlusion. *Stroke* 2001;32:1552–8.
- [191] Lythgoe D, Simmons A, Pereira A, Cullinane M, Williams S, Markus HS. Magnetic resonance markers of ischaemia: their correlation with vasodilatory reserve in patients with carotid artery stenosis and occlusion. *J Neurol Neurosurg Psychiatry* 2001;71:58–62.
- [192] Ziyyeh S, Rick J, Reinhard M, Hetzel A, Mader I, Speck O. Blood oxygen level-dependent MRI of cerebral CO₂ reactivity in severe carotid stenosis and occlusion. *Stroke* 2005;36:751–6.
- [193] Conklin J, Fierstra J, Crawley AP, Han JS, Poublanc J, Silver FL, et al. Mapping white matter diffusion and cerebrovascular reactivity in carotid occlusive disease. *Neurology* 2011;77:431–8.
- [194] Mandell DM, Han JS, Poublanc J, Crawley AP, Fierstra J, Tymianski M, et al. Quantitative measurement of cerebrovascular reactivity by blood oxygen level-dependent MR imaging in patients with intracranial stenosis: preoperative cerebrovascular reactivity predicts the effect of extracranial-intracranial bypass surgery. *AJNR Am J Neuroradiol* 2011;32:721–7.
- [195] Mandell DM, Han JS, Poublanc J, Crawley AP, Stainsby JA, Fisher JA, et al. Mapping cerebrovascular reactivity using blood oxygen level-dependent MRI in Patients with arterial steno-occlusive disease: comparison with arterial spin labeling MRI. *Stroke* 2008;39:2021–8.
- [196] Bernier AV, Correia CE, Haller MJ, Theriaque DW, Shuster JJ, Weinstein DA. Vascular dysfunction in glycogen storage disease type I. *J Pediatr* 2009;154:588–91.
- [197] Haller S, Bonati LH, Rick J, Klarhofer M, Speck O, Lyrer PA, et al. Reduced cerebrovascular reserve at CO₂ BOLD MR imaging is associated with increased risk of periinterventional ischemic lesions during carotid endarterectomy or stent placement: preliminary results. *Radiology* 2008;249:251–8.
- [198] Attye A, Villien M, Tahon F, Warnking J, Detante O, Krainik A. Normalization of cerebral vasoreactivity using BOLD MRI after intravascular stenting. *Hum Brain Mapp* 2013, <http://dx.doi.org/10.1002/hbm.22255>.
- [199] Weinand ME, Carter LP, Oommen KJ, Hutzler R, Labiner DM, Talwar D, et al. Response of human epileptic temporal lobe cortical blood flow to hyperventilation. *Epilepsy Res* 1995;21:221–6.
- [200] Tae WS, Joo EY, Kim JH, Han SJ, Suh YL, Kim BT, et al. Cerebral perfusion changes in mesial temporal lobe epilepsy: SPM analysis of ictal and interictal SPECT. *Neuroimage* 2005;24:101–10.
- [201] Katayama S, Momose T, Sano I, Nakashima Y, Nakajima T, Niwa S, et al. Temporal lobe CO₂ vasoreactivity in patients with complex partial seizures. *Jpn J Psychiatry Neurol* 1992;46:379–85.

- [202] Zhao P, Alsop DC, Abduljalil A, Selim M, Lipsitz L, Novak P, et al. Vasoreactivity and peri-infarct hyperintensities in stroke. *Neurology* 2009;72:643–9.
- [203] Fierstra J, Conklin J, Krings T, Slessarev M, Han JS, Fisher JA, et al. Impaired peri-nidal cerebrovascular reserve in seizure patients with brain arteriovenous malformations. *Brain* 2011;134:100–9.
- [204] Cohen ER, Rostrup E, Sidaros K, Lund TE, Paulson OB, Ugurbil K, et al. Hypercapnic normalization of BOLD fMRI: comparison across field strengths and pulse sequences. *Neuroimage* 2004;23:613–24.
- [205] Stefanovic B, Warnking JM, Pike GB. Hemodynamic and metabolic responses to neuronal inhibition. *Neuroimage* 2004;22:771–8.
- [206] Kuroda S, Kamiyama H, Abe H, Houkin K, Isobe M, Mitsumori K. Acetazolamide test in detecting reduced cerebral perfusion reserve and predicting long-term prognosis in patients with internal carotid artery occlusion. *Neurosurgery* 1993;32:912–8 [discussion 8–9].
- [207] Cao B, Hasegawa Y, Yokota C, Minematsu K, Yamaguchi T. Spontaneous improvement in reduced vasodilatory capacity in major cerebral arterial occlusive disease. *Neuroradiology* 2000;42:19–25.
- [208] Ogasawara K, Ogawa A, Yoshimoto T. Cerebrovascular reactivity to acetazolamide and outcome in patients with symptomatic internal carotid or middle cerebral artery occlusion: a xenon-133 single-photon emission computed tomography study. *Stroke* 2002;33:1857–62.
- [209] Gur AY, Bova I, Bornstein NM. Is impaired cerebral vasomotor reactivity a predictive factor of stroke in asymptomatic patients? *Stroke* 1996;27:2188–90.
- [210] Markus H, Cullinane M. Severely impaired cerebrovascular reactivity predicts stroke and TIA risk in patients with carotid artery stenosis and occlusion. *Brain* 2001;124:457–67.
- [211] Kuroda S, Houkin K, Kamiyama H, Mitsumori K, Iwasaki Y, Abe H. Long-term prognosis of medically treated patients with internal carotid or middle cerebral artery occlusion: can acetazolamide test predict it? *Stroke* 2001;32:2110–6.
- [212] Muller M, Voges M, Piepras U, Schimrigk K. Assessment of cerebral vasomotor reactivity by transcranial Doppler ultrasound and breath-holding. A comparison with acetazolamide as vasodilatory stimulus. *Stroke* 1995;26:96–100.
- [213] Silvestrini M, Vernieri F, Pasqualetti P, Matteis M, Passarelli F, Troisi E, et al. Impaired cerebral vasoreactivity and risk of stroke in patients with asymptomatic carotid artery stenosis. *JAMA* 2000;283:2122–7.
- [214] King A, Serena J, Bornstein NM, Markus HS. Does impaired cerebrovascular reactivity predict stroke risk in asymptomatic carotid stenosis? A prospective substudy of the asymptomatic carotid emboli study. *Stroke* 2011;42:1550–5.
- [215] Leoni RF, Mazzetto-Betti KC, Silva AC, Dos Santos AC, de Araujo DB, Leite JP, et al. Assessing cerebrovascular reactivity in carotid steno-occlusive disease using MRI BOLD and ASL techniques. *Radiol Res Pract* 2012, <http://dx.doi.org/10.1155/2012/268483>.
- [216] Bokkers RP, van Osch MJ, Klijn CJ, Kappelle LJ, Hendrikse J. Cerebrovascular reactivity within perfusion territories in patients with an internal carotid artery occlusion. *J Neurol Neurosurg Psychiatry* 2011;82:1011–6.
- [217] Hartkamp NS, Hendrikse J, van der Worp HB, de Borst GJ, Bokkers RP. Time course of vascular reactivity using repeated phase-contrast MR angiography in patients with carotid artery stenosis. *Stroke* 2012;43:553–6.
- [218] Mazighi M, Tanasescu R, Ducrocq X, Vicaut E, Bracard S, Houdart E, et al. Prospective study of symptomatic atherothrombotic intracranial stenoses: the GESICA study. *Neurology* 2006;66:1187–91.
- [219] Chimowitz MI, Lynn MJ, Derdeyn CP, Turan TN, Fiorella D, Lane BF, et al. Stenting versus aggressive medical therapy for intracranial arterial stenosis. *N Engl J Med* 2011;365:993–1003.
- [220] Goode SD, Altaf N, Auer DP, MacSweeney ST. Carotid endarterectomy improves cerebrovascular reserve capacity preferentially in patients with preoperative impairment as indicated by asymmetric BOLD response to hypercapnia. *Eur J Vasc Endovasc Surg* 2009;38:546–51.
- [221] Chang TY, Liu HL, Lee TH, Kuan WC, Chang CH, Wu HC, et al. Change in cerebral perfusion after carotid angioplasty with stenting is related to cerebral vasoreactivity: a study using dynamic susceptibility-weighted contrast-enhanced MR imaging and functional MR imaging with a breath-holding paradigm. *AJR Am J Neuroradiol* 2009;30:1330–6.
- [222] So Y, Lee HY, Kim SK, Lee JS, Wang KC, Cho BK, et al. Prediction of the clinical outcome of pediatric moyamoya disease with postoperative basal/acetazolamide stress brain perfusion SPECT after revascularization surgery. *Stroke* 2005;36:1485–9.
- [223] Conklin J, Fierstra J, Crawley AP, Han JS, Poublanc J, Mandell DM, et al. Impaired cerebrovascular reactivity with steal phenomenon is associated with increased diffusion in white matter of patients with Moyamoya disease. *Stroke* 2010;41:1610–6.
- [224] Han JS, Abou-Hamden A, Mandell DM, Poublanc J, Crawley AP, Fisher JA, et al. Impact of extracranial-intracranial bypass on cerebrovascular reactivity and clinical outcome in patients with symptomatic moyamoya vasculopathy. *Stroke* 2011;42:3047–54.
- [225] Mikulis DJ, Krolczyk G, Desal H, Logan W, Deveber G, Dirks P, et al. Preoperative and postoperative mapping of cerebrovascular reactivity in moyamoya disease by using blood oxygen level-dependent magnetic resonance imaging. *J Neurosurg* 2005;103:347–55.
- [226] Calviere L, Ssi Yan Kai G, Catalaa I, Marlats F, Bonneville F, Larrue V. Executive dysfunction in adults with moyamoya disease is associated with increased diffusion in frontal white matter. *J Neurol Neurosurg Psychiatry* 2012;83:591–3.
- [227] Kalaria RN. Cerebral vessels in ageing and Alzheimer's disease. *Pharmacol Ther* 1996;72:193–214.
- [228] Lythgoe DJ, Williams SC, Cullinane M, Markus HS. Mapping of cerebrovascular reactivity using BOLD magnetic resonance imaging. *Magn Reson Imaging* 1999;17:495–502.
- [229] Ono Y, Morikawa S, Inubushi T, Shimizu H, Yoshimoto T. T2*-weighted magnetic resonance imaging of cerebrovascular reactivity in rat reversible focal cerebral ischemia. *Brain Res* 1997;744:207–15.
- [230] Harris NG, Lythgoe MF, Thomas DL, Williams SR. Cerebrovascular reactivity following focal brain ischemia in the rat: a functional magnetic resonance imaging study. *Neuroimage* 2001;13:339–50.
- [231] Csete K, Vezekenyi Z, Doczi T, Papp JG, Bodosi M, Barzo P. Comparison of regional vasomotor responses to acetazolamide and CO₂ in rabbit cerebrum and cerebellum, measured by a hydrogen clearance method. *Acta Physiol Scand* 2004;182:287–94.
- [232] Julien C, Payen JF, Tropres I, Farion R, Grillon E, Montigon O, et al. Assessment of vascular reactivity in rat brain glioma by measuring regional blood volume during graded hypoxic hypoxia. *Br J Cancer* 2004;91:374–80.
- [233] Niwa K, Kazama K, Younkin L, Younkin SG, Carlson GA, Iadecola C. Cerebrovascular autoregulation is profoundly impaired in mice overexpressing amyloid precursor protein. *Am J Physiol Heart Circ Physiol* 2002;283:H315–23.
- [234] Iadecola C. Neurovascular regulation in the normal brain and in Alzheimer's disease. *Nat Rev Neurosci* 2004;5:347–60.

- [235] Mueggler T, Sturchler-Pierrat C, Baumann D, Rausch M, Staufenbiel M, Rudin M. Compromised hemodynamic response in amyloid precursor protein transgenic mice. *J Neurosci* 2002;22:7218–24.
- [236] Princz-Kranz FL, Mueggler T, Knobloch M, Nitsch RM, Rudin M. Vascular response to acetazolamide decreases as a function of age in the arca beta mouse model of cerebral amyloidosis. *Neurobiol Dis* 2010;40:284–92.
- [237] Suga Y, Ogasawara K, Saito H, Kobayashi M, Inoue T, Kondo R, et al. Endarterectomy for cervical internal carotid artery stenosis accompanied with severe aortic valve stenosis – case report. *Brain Nerve* 2007;59:1377–81.
- [238] Weinand ME, Labiner DM, Ahern GL. Temporal lobe seizure interhemispheric propagation time depends on non-epileptic cortical cerebral blood flow. *Epilepsy Res* 2001;44:33–9.
- [239] Yune MJ, Lee JD, Ryu YH, Kim DJ, Lee BI, Kim SJ. Ipsilateral thalamic hypoperfusion on interictal SPECT in temporal lobe epilepsy. *J Nucl Med* 1998;39:281–5.
- [240] Weinand ME, Carter LP, el-Saadany WF, Sioutos PJ, Labiner DM, Ommen KJ. Cerebral blood flow and temporal lobe epileptogenicity. *J Neurosurg* 1997;86:226–32.
- [241] Baumgartner C, Serles W, Leutmezer F, Pataria E, Aull S, Czech T, et al. in temporal lobe epilepsy: regional cerebral blood flow is increased prior to electroencephalography-seizure onset. *J Nucl Med* 1998;39:978–82.
- [242] Fierstra J, Spieth S, Tran L, Conklin J, Tymianski M, ter Brugge KG, et al. Severely impaired cerebrovascular reserve in patients with cerebral proliferative angiopathy. *J Neurosurg Pediatr* 2011;8:310–5.
- [243] Landuyt W, Hermans R, Bosmans H, Sunaert S, Beatse E, Farina D, et al. BOLD contrast fMRI of whole rodent tumour during air or carbogen breathing using echo-planar imaging at 1.5 T. *Eur Radiol* 2001;11:2332–40.
- [244] Dunn JF, O'Hara JA, Zaim-Wadghiri Y, Lei H, Meyerand ME, Grinberg OY, et al. Changes in oxygenation of intracranial tumors with carbogen: a BOLD MRI and EPR oximetry study. *J Magn Reson Imaging* 2002;16:511–21.
- [245] Taylor NJ, Baddeley H, Goodchild KA, Powell ME, Thoumine M, Culver LA, et al. BOLD MRI of human tumor oxygenation during carbogen breathing. *J Magn Reson Imaging* 2001;14: 156–63.
- [246] Rauscher A, Sedlacik J, Barth M, Haacke EM, Reichenbach JR. Nonvasive assessment of vascular architecture and function during modulated blood oxygenation using susceptibility weighted magnetic resonance imaging. *Magn Reson Med* 2005;54:87–95.
- [247] Lemasson B, Pannetier N, Christen T, Warnking J, Krainik A, Farion R, et al. Assessment of vascular reactivity in two rat brain gliomas (C6 and RG2) by blood volume fraction MRI during CO₂ challenge and correlation to mature vessels. 2009.
- [248] Schalen W, Messeter K, Nordstrom CH. Cerebral vasoreactivity and the prediction of outcome in severe traumatic brain lesions. *Acta Anaesthesiol Scand* 1991;35:113–22.
- [249] Steiner LA, Coles JP, Johnston AJ, Chatfield DA, Smielewski P, Fryer TD, et al. Assessment of cerebrovascular autoregulation in head-injured patients: a validation study. *Stroke* 2003;34:2404–9.
- [250] Smith LM, Elkins JS, Dillon WP, Schaeffer S, Wintermark M. Perfusion-CT assessment of the cerebrovascular reserve: a revisit to the acetazolamide challenges. *J Neuroradiol* 2008;35:157–64.
- [251] Kastrup A, Kruger G, Neumann-Haefelin T, Moseley ME. Assessment of cerebrovascular reactivity with functional magnetic resonance imaging: comparison of CO₍₂₎ and breath holding. *Magn Reson Imaging* 2001;19:13–20.
- [252] van der Zande FH, Hofman PA, Backes WH. Mapping hypercapnia-induced cerebrovascular reactivity using BOLD MRI. *Neuroradiology* 2005;47:114–20.
- [253] Bokkers RP, van Osch MJ, van der Worp HB, de Borst GJ, Mali WP, Hendrikse J. Symptomatic carotid artery stenosis: impairment of cerebral autoregulation measured at the brain tissue level with arterial spin-labeling MR imaging. *Radiology* 2010;256:201–8.
- [254] Noth U, Kotajima F, Deichmann R, Turner R, Corfield DR. Mapping of the cerebral vascular response to hypoxia and hypercapnia using quantitative perfusion MRI at 3 T. *NMR Biomed* 2008;21:464–72.
- [255] Noth U, Meadows GE, Kotajima F, Deichmann R, Corfield DR, Turner R. Cerebral vascular response to hypercapnia: determination with perfusion MRI at 1.5 and 3.0 Tesla using a pulsed arterial spin labeling technique. *J Magn Reson Imaging* 2006;24:1229–35.
- [256] Berthezene Y, Nighoghossian N, Meyer R, Damien J, Cinotti L, Adeleine P, et al. Can cerebrovascular reactivity be assessed by dynamic susceptibility contrast-enhanced MRI? *Neuroradiology* 1998;40:1–5.
- [257] Calviere L, Catalaa I, Marlats F, Viguer A, Bonneville F, Cognard C, et al. Correlation between cognitive impairment and cerebral hemodynamic disturbances on perfusion magnetic resonance imaging in European adults with moyamoya disease. Clinical article. *J Neurosurg* 2010;113: 753–9.
- [258] Bokkers RP, Wessels FJ, van der Worp HB, Zwanenburg JJ, Mali WP, Hendrikse J. Vasodilatory capacity of the cerebral vasculature in patients with carotid artery stenosis. *AJR Am J Neuroradiol* 2011;32:1030–3.
- [259] Villien M, Chipon E, Tropres I, Bouvier J, Cantin S, Chechin D, et al. Per-subject characterization of bolus width in pulsed arterial spin labeling using bolus turbo sampling. *Magn Reson Med* 2013;69:1677–82.
- [260] Goode SD, Krishan S, Alexakis C, Mahajan R, Auer DP. Precision of cerebrovascular reactivity assessment with use of different quantification methods for hypercapnia functional MR imaging. *AJR Am J Neuroradiol* 2009;30:972–7.
- [261] Okazawa H, Yamauchi H, Sugimoto K, Takahashi M. Differences in vasodilatory capacity and changes in cerebral blood flow induced by acetazolamide in patients with cerebrovascular disease. *J Nucl Med* 2003;44:1371–8.
- [262] Rostrup E, Law I, Pott F, Ide K, Knudsen GM. Cerebral hemodynamics measured with simultaneous PET and near-infrared spectroscopy in humans. *Brain Res* 2002;954:183–93.
- [263] Kannurpatti SS, Biswal BB, Hudetz AG. Differential fMRI-BOLD signal response to apnea in humans and anesthetized rats. *Magn Reson Med* 2002;47:864–70.
- [264] Kastrup A, Kruger G, Glover GH, Neumann-Haefelin T, Moseley ME. Regional variability of cerebral blood oxygenation response to hypercapnia. *Neuroimage* 1999;10:675–81.
- [265] Kastrup A, Kruger G, Glover GH, Moseley ME. Assessment of cerebral oxidative metabolism with breath holding and fMRI. *Magn Reson Med* 1999;42:608–11.
- [266] Thomason ME, Burrows BE, Gabrieli JD, Glover GH. Breath holding reveals differences in fMRI BOLD signal in children and adults. *Neuroimage* 2005;25:824–37.
- [267] Przybylowski T, Bangash MF, Reichmuth K, Morgan BJ, Skatrud JB, Dempsey JA. Mechanisms of the cerebrovascular response to apnoea in humans. *J Physiol* 2003;548:323–32.
- [268] Yezhuvath US, Lewis-Amezua K, Varghese R, Xiao GH, Lu HZ. On the assessment of cerebrovascular reactivity using hypercapnia BOLD MRI. *Nmr in Biomedicine* 2009;22: 779–86.
- [269] Jones M, Berwick J, Hewson-Stoate N, Gias C, Mayhew J. The effect of hypercapnia on the neural and hemodynamic responses to somatosensory stimulation. *Neuroimage* 2005;27:609–23.
- [270] Jensen D, Wolfe LA, Slatkovska L, Webb KA, Davies GA, O'Donnell DE. Effects of human pregnancy on the ventilatory

- chemoreflex response to carbon dioxide. *Am J Physiol Regul Integr Comp Physiol* 2005;288:R1369–75.
- [271] Jensen D, Wolfe LA, O'Donnell DE, Davies GA. Chemoreflex control of breathing during wakefulness in healthy men and women. *J Appl Physiol* 2005;98:822–8.
- [272] Spano VR, Mandell DM, Poublanc J, Sam K, Battistin-Charbonney A, Pucci O, et al. CO₂ blood oxygen level-dependent MR mapping of cerebrovascular reserve in a clinical population: safety, tolerability, and technical feasibility. *Radiology* 2013;266:592–8.
- [273] Bailey JE, Kendrick A, Diaper A, Potokar J, Nutt DJ. A validation of the 7.5% CO₂ model of GAD using paroxetine and lorazepam in healthy volunteers. *J Psychopharmacol* 2007;21(1):42–9.
- [274] Coryell W, Pine D, Fyer A, Klein D. Anxiety responses to CO₂ inhalation in subjects at high-risk for panic disorder. *J Affect Disord* 2006;92:63–70.
- [275] Bailey JE, Argyropoulos SV, Kendrick AH, Nutt DJ. Behavioral and cardiovascular effects of 7.5% CO₂ in human volunteers. *Depress Anxiety* 2005;21:18–25.
- [276] Kaanders JH, Bussink J, van der Kogel AJ. Clinical studies of hypoxia modification in radiotherapy. *Semin Radiat Oncol* 2004;14:233–40.
- [277] Simon JM, Noel G, Chiras J, Hoang-Xuan K, Delattre JY, Baillet F, et al. Radiotherapy and chemotherapy with or without carbogen and nicotinamide in inoperable biopsy-proven glioblastoma multiforme. *Radiother Oncol* 2003;67:45–51.
- [278] Griffiths JR, Taylor NJ, Howe FA, Saunders MI, Robinson SP, Hoskin PJ, et al. The response of human tumors to carbogen breathing, monitored by Gradient-recalled echo magnetic resonance imaging. *Int J Radiat Oncol Biol Phys* 1997;39:697–701.
- [279] Evelhoch JL, Gillies RJ, Karczmar GS, Koutcher JA, Maxwell RJ, Nalcioglu O, et al. Applications of magnetic resonance in model systems: cancer therapeutics. *Neoplasia* 2000;2:152–65.
- [280] van Zijl PC, Eleff SM, Ulatowski JA, Oja JM, Ulug AM, Traystman RJ, et al. Quantitative assessment of blood flow, blood volume and blood oxygenation effects in functional magnetic resonance imaging. *Nat Med* 1998;4:159–67.